



2019 台灣胸腔暨重症加護醫學會

2019 Taiwan Society of Pulmonary and Critical Care Medicine

藉阻斷鈣離子釋放激活鈣離子通道降低大鼠動物呼吸器
引起內質網壓力與改善肺損傷

Blocking Calcium Release-activated Calcium Channel Attenuates Ventilator-induced Endoplasmic Reticulum Stress and Lung Injury in Rats

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A Ventilation at low lung volume

End expiration



Atelectrauma

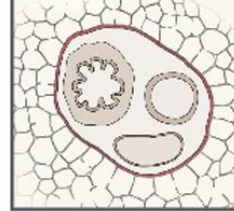
End inspiration



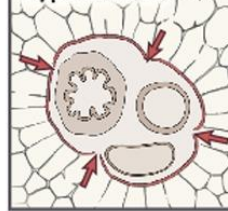
Lung inhomogeneity

B Ventilation at high lung volume

Normal



Hyperinflation

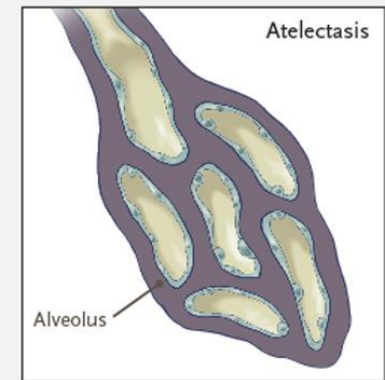
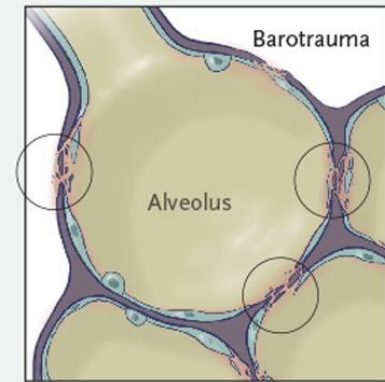
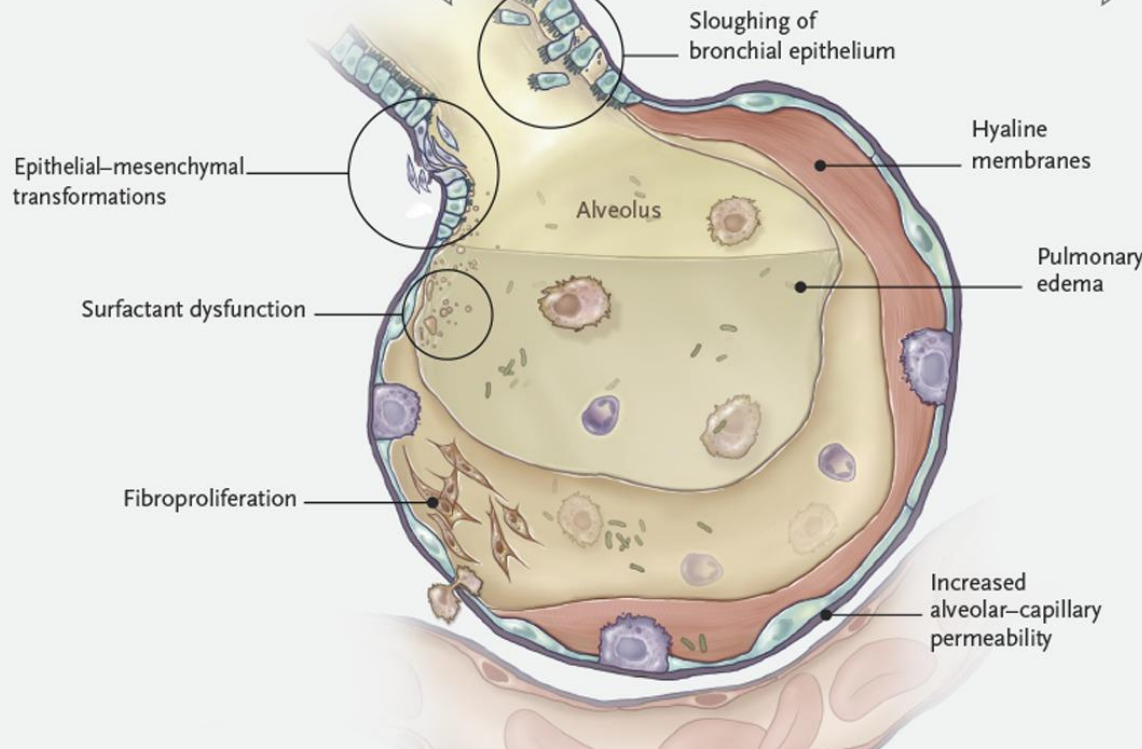


Air leaks



Overdistention

C Structural consequences



Biologic alterations

Increased concentrations of:
Hydroxyproline
Transforming growth factor- β

TNF- α

PMN

Increased physiological dead space

N Engl J Med 2013; 369:2126-2136

Fibroproliferation

Increased alveolar-capillary permeability

Alveolus

Biologic alterations

- Increased concentrations of:
 - Hydroxyproline
 - Transforming growth factor- β
 - Interleukin-8
- Release of mediators:
 - Tumor necrosis factor α (TNF- α)
 - β -catenin
 - Interleukin-6 (IL-6)
 - Interleukin-1 β (IL-1 β)
- Recruitment of:
 - Pulmonary alveolar macrophages (PAMs)
 - Neutrophils
- Activation of epithelium and endothelium

Physiological abnormalities

- Increased physiological dead space
- Decreased compliance
- Decreased Pao₂
- Increased Paco₂

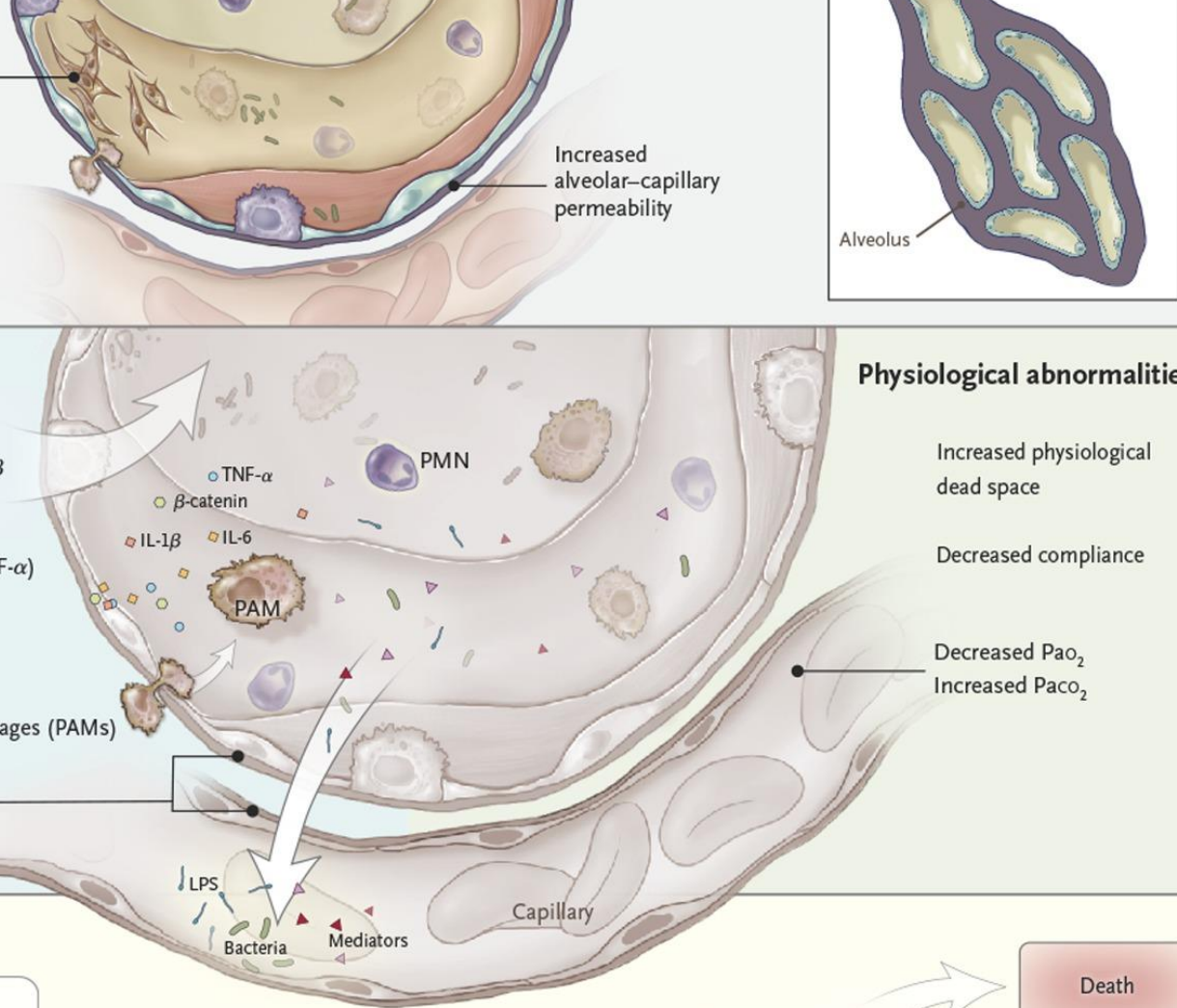
Systemic effects

Translocation of:
Lipopolysaccharides (LPS)
Bacteria
Various mediators

Multiple mechanisms
(e.g., increased apoptosis)

Multiorgan dysfunction

Death

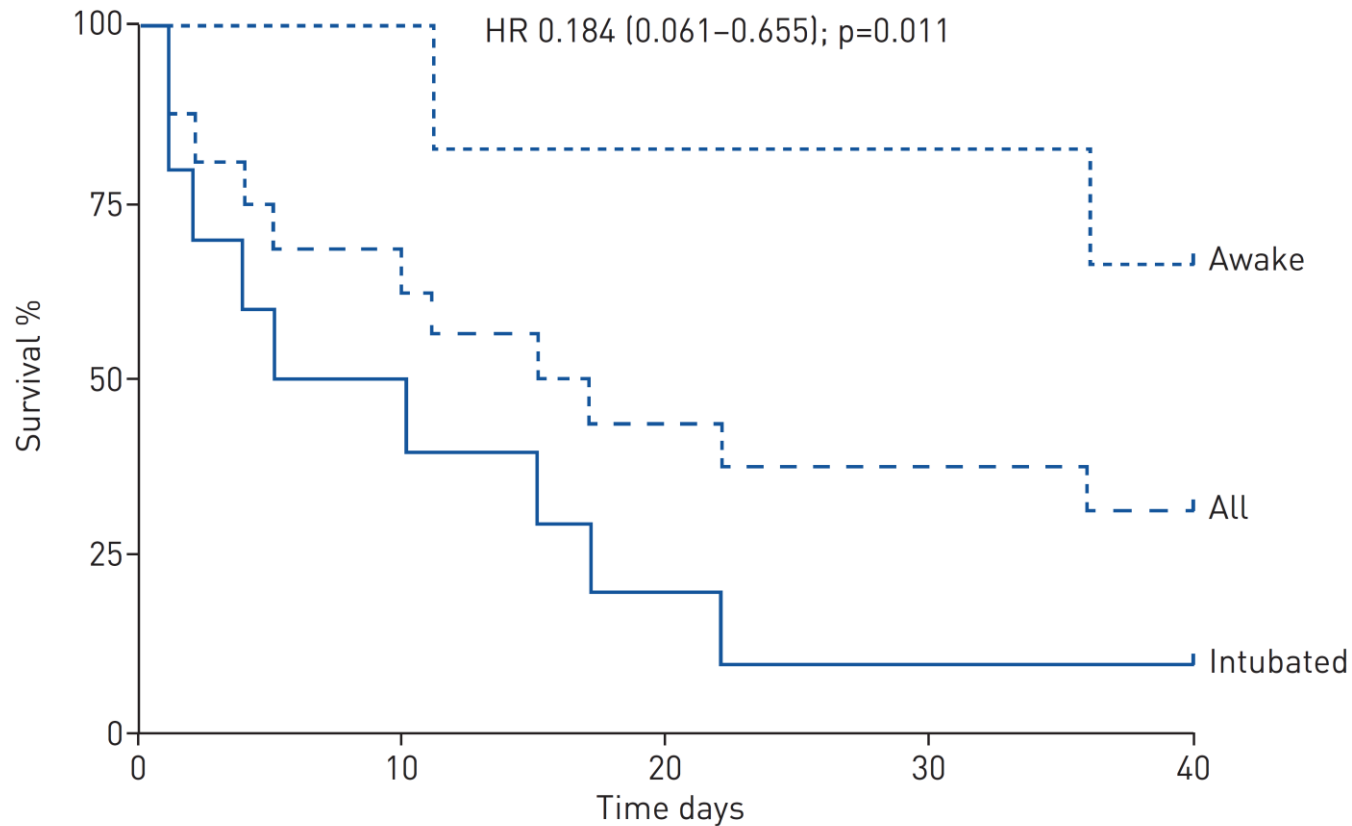


ECMO in a non-intubated patient with ARDS (Awake ECMO)



European Respiratory Journal 2012 40: 1296-1298

Awake ECMO for ARDS due to *Pneumocystis pneumonia*



Introduction

Acute respiratory distress syndrome (ARDS)

Unfortunately, there is **no specific therapy for ALI/ARDS.**

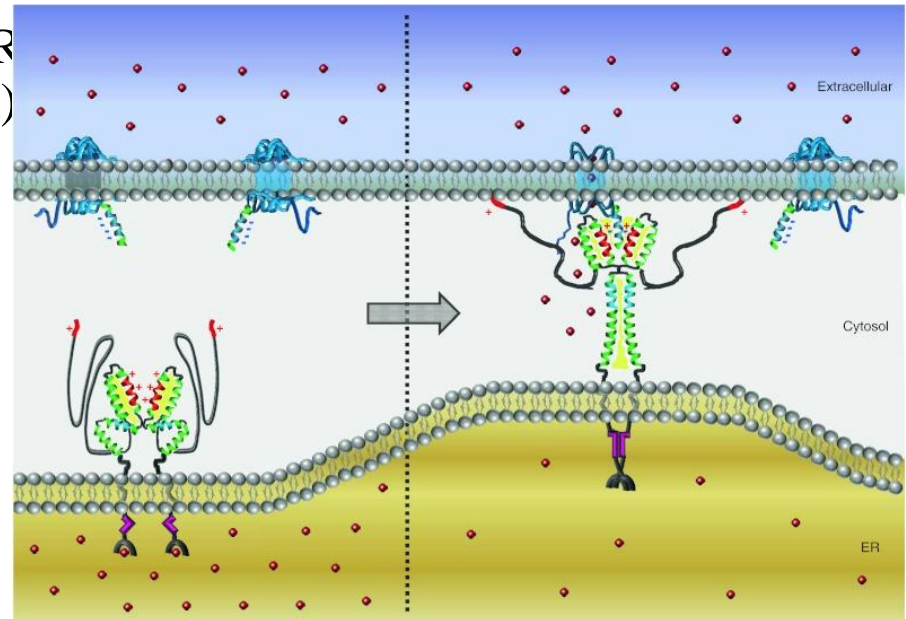
Hypothesis

Calcium release-activated Calcium Channel participate in the pathogenesis of ventilator-induced lung injury.

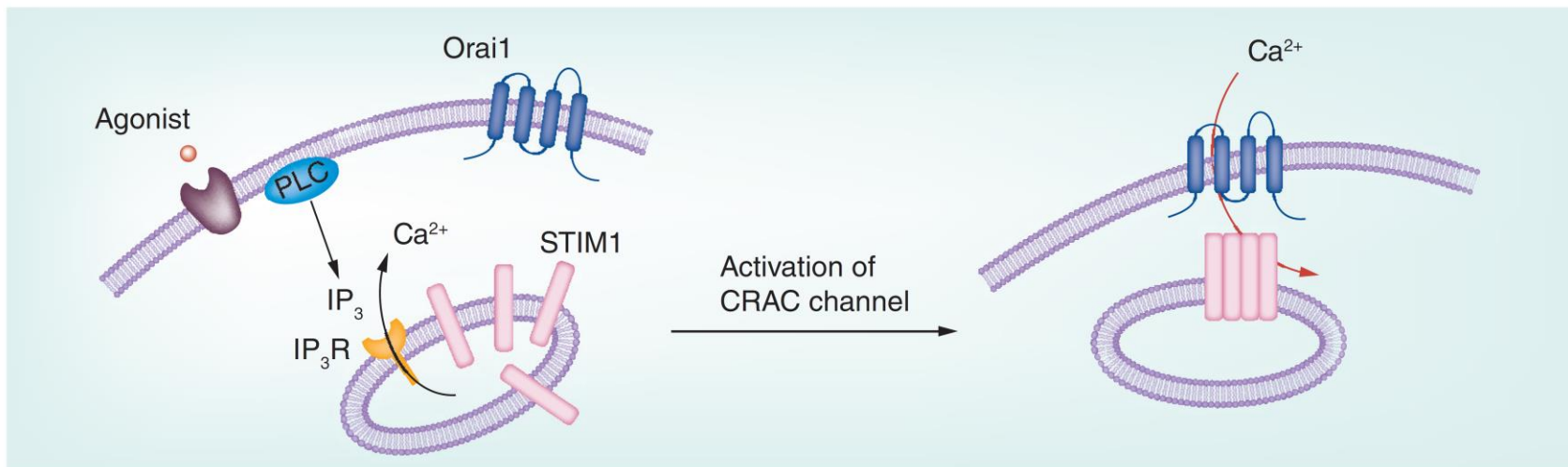
Calcium release-activated Calcium (CRAC) Channel

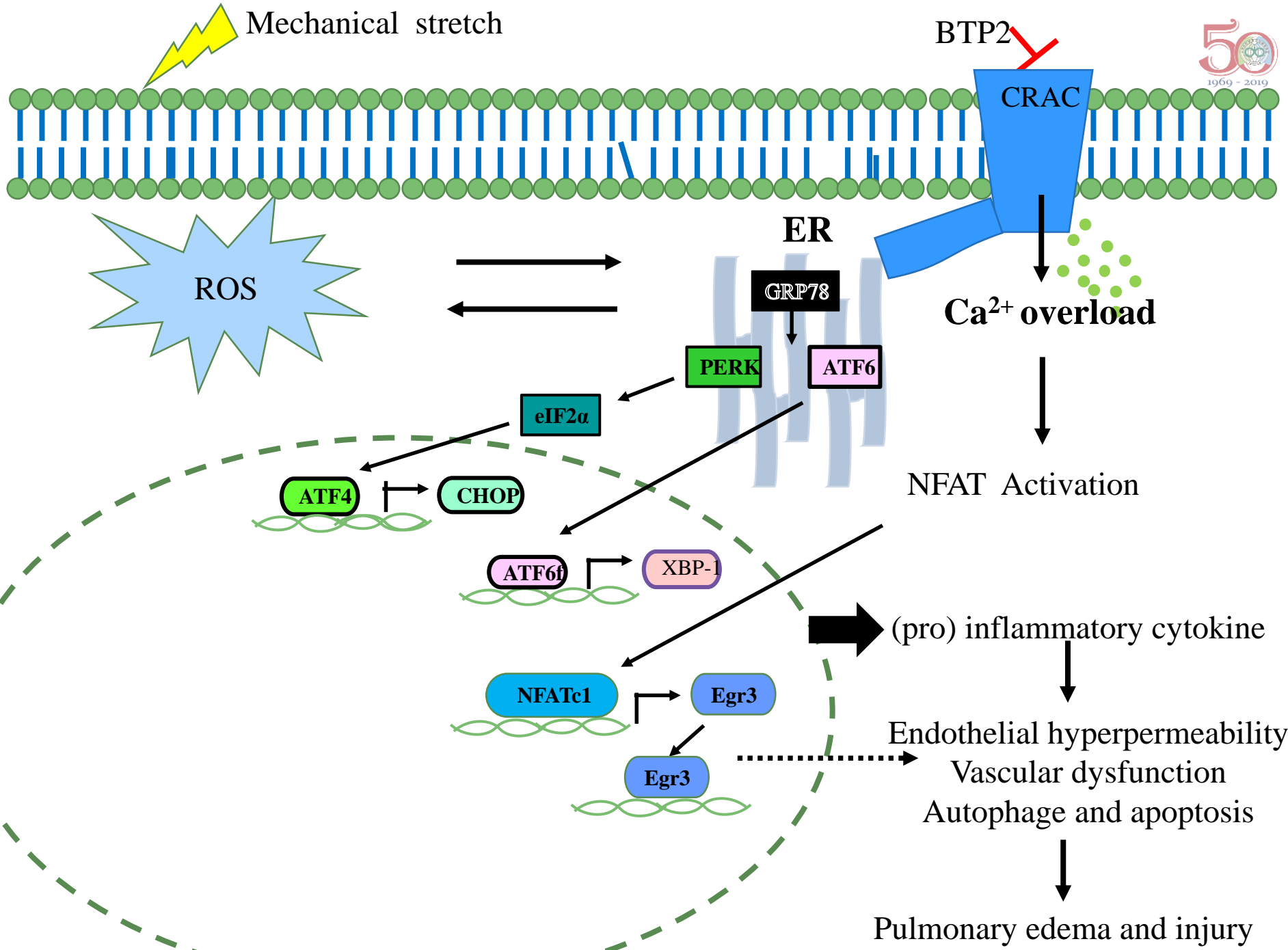
- **Store-operated calcium (Ca²⁺) entry (SOCE)** : molecules located on the endo/sarcoplasmic reticulum (ER/SR) respond to decreased luminal Ca²⁺ levels by signaling Ca²⁺ release activated Ca²⁺ channels (CRAC) channels to open on the plasma membrane (PM).
- ER: **stromal interaction molecules (STIMs)**: functioning as the ER/SR luminal Ca²⁺ sensors

Plasma membrane: CRAC channels: Orai proteins



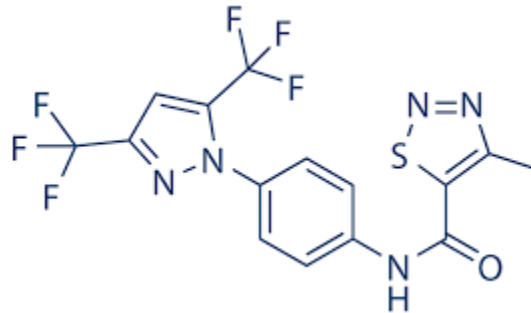
Activation of release-activated Ca^{2+} channel

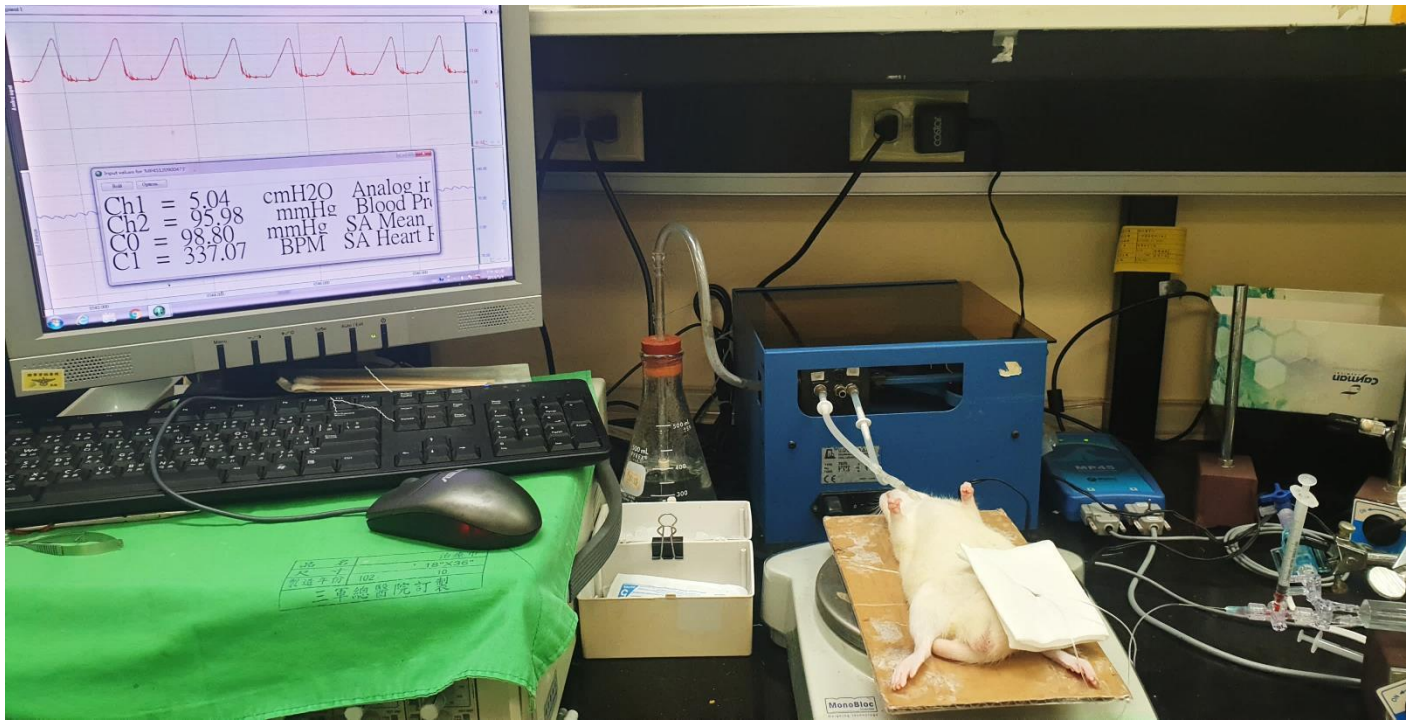




Methods

- **Methods:** Male SD rats were exposed to ventilator with normal or high tidal-volume ventilation with intraperitoneal injection of BTP2.
- **BTP2: Calcium release-activated Calcium Channel (CRAC channel) inhibitor**





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PEEP = 4 cmH₂O
 Respiratory rates(RR) = 66-68/min
 BTP2(0.5 mg/kg 、 1mg/kg 、 2mg/kg)

BTP2 i.p.

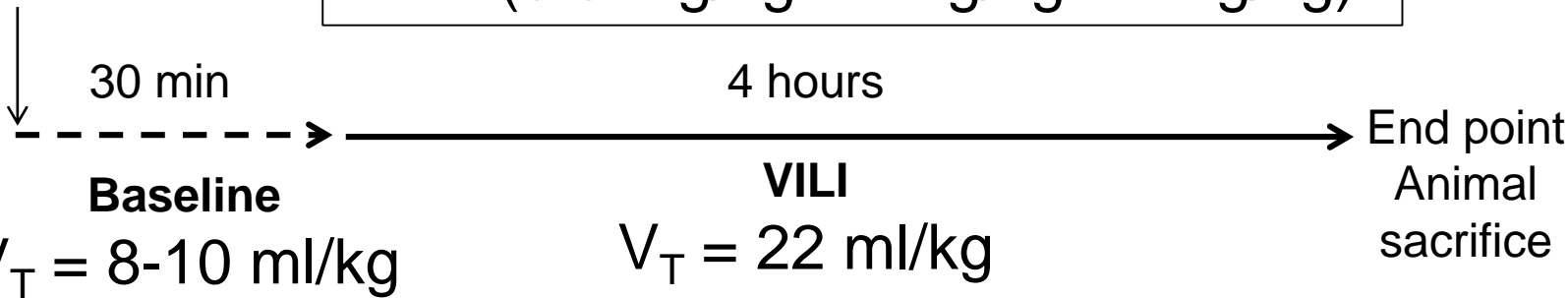
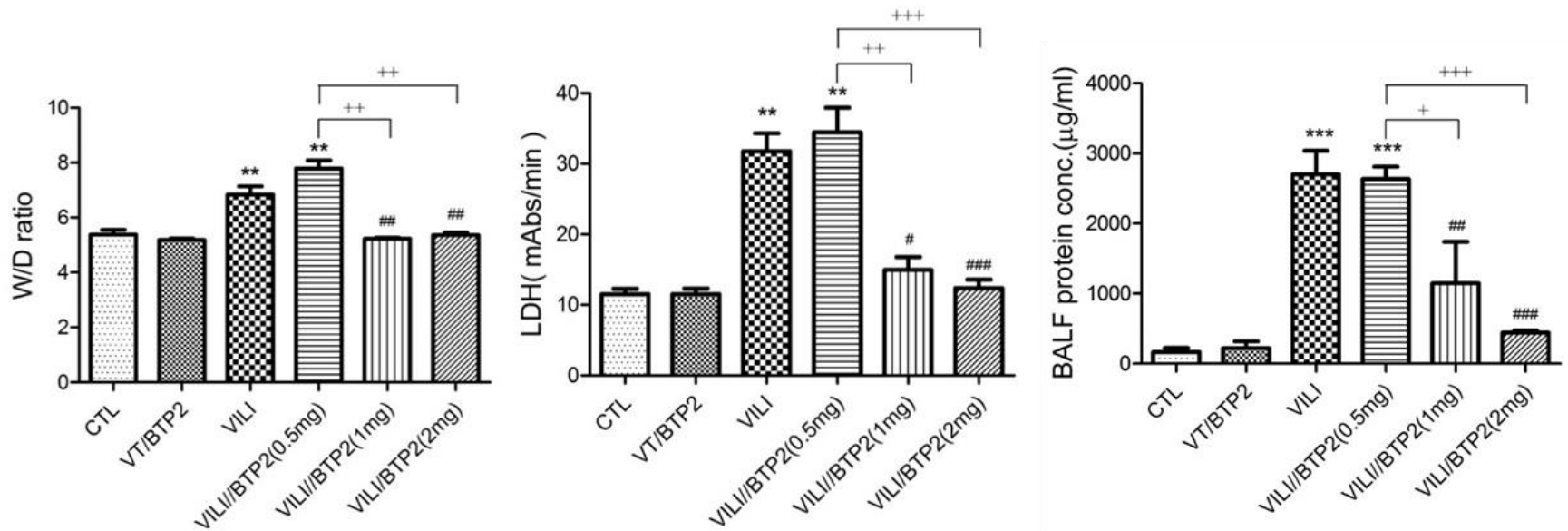


FIGURE 1 Effect of BTP2 on lung edema.



(A) Lung wet/dry ratio
(B) LDH level
(C) Protein concentrations in bronchoalveolar lavage fluid (BALF)

Fig. 2

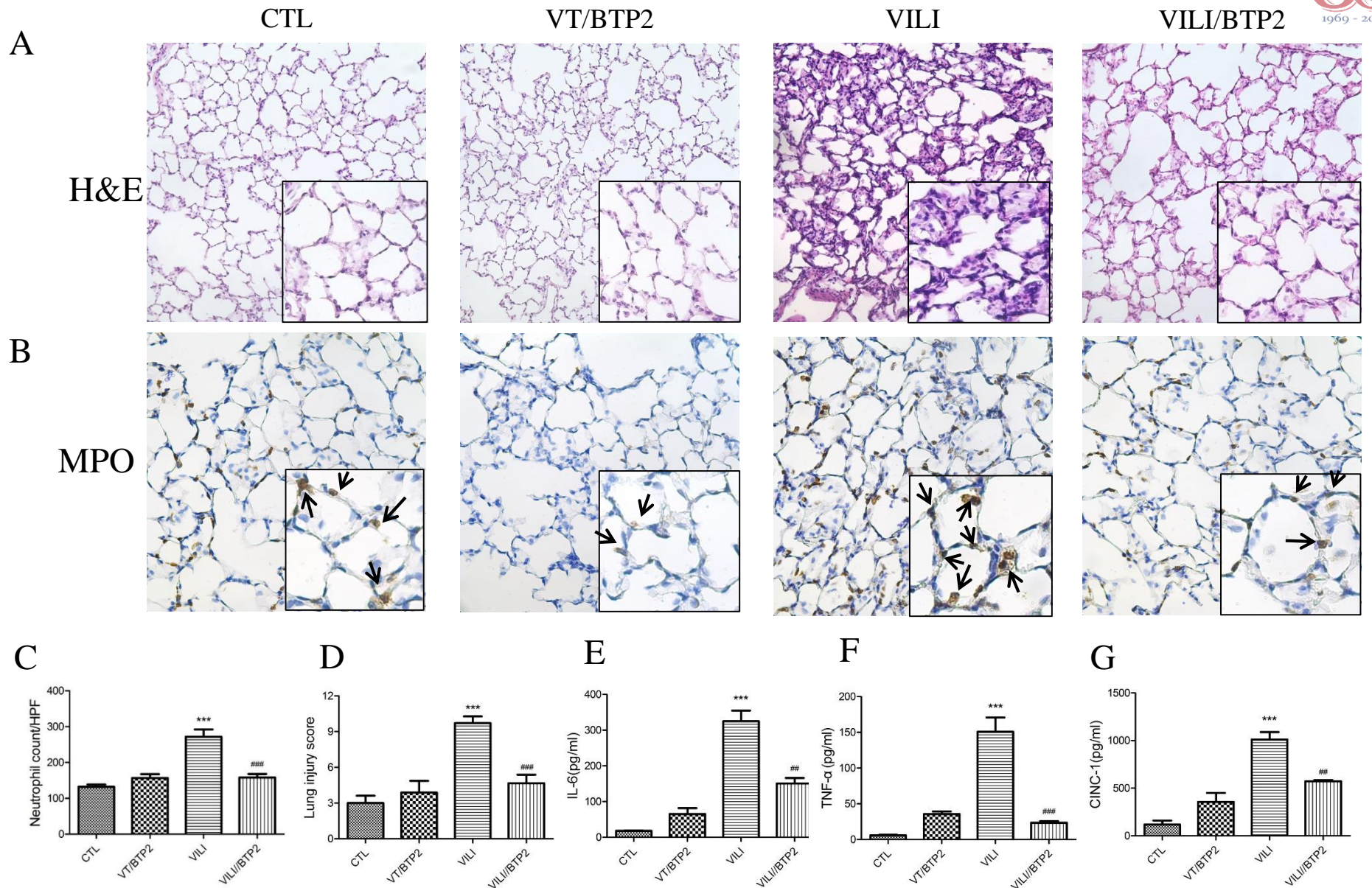


FIGURE 2 Assessment of BTP2 treatment on lung inflammatory.

Fig. 3

Lung injury: Autophagy

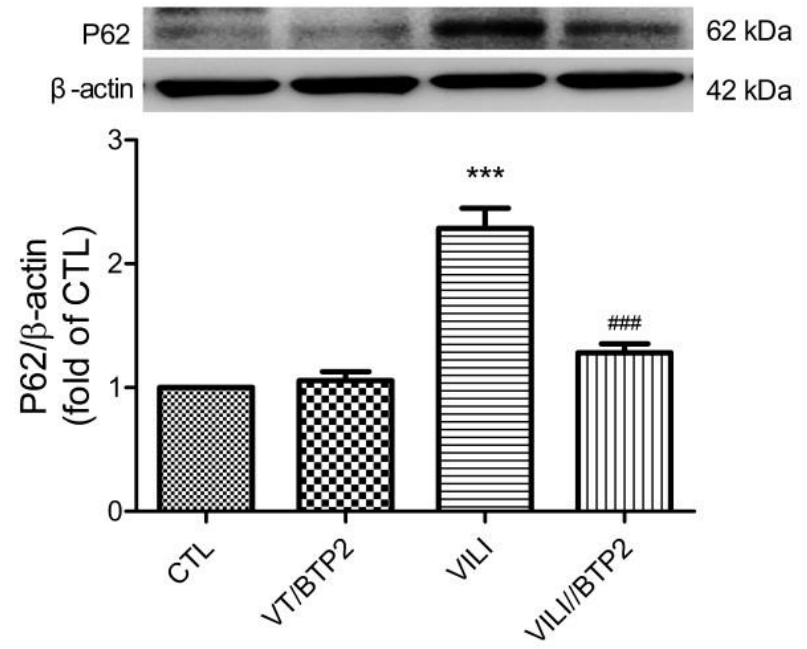
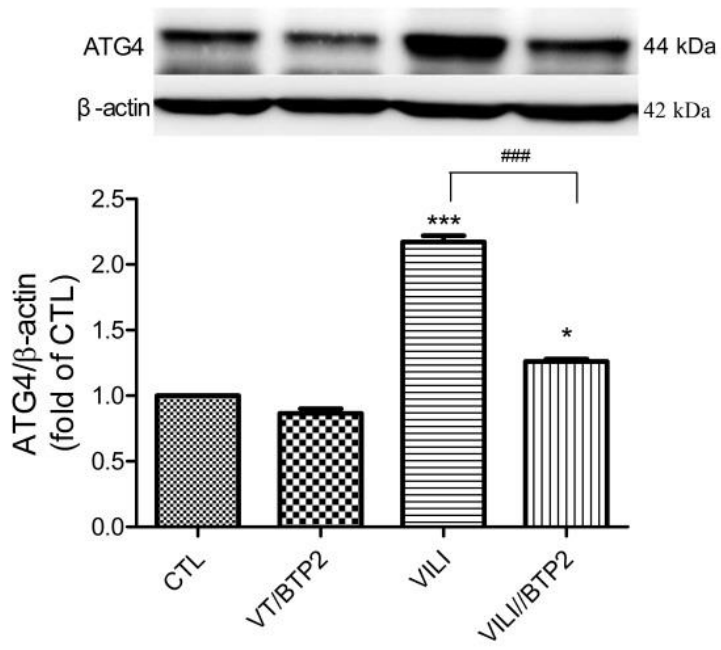


Fig. 3

Lung injury: Apoptosis

CTL

VT/BTP2

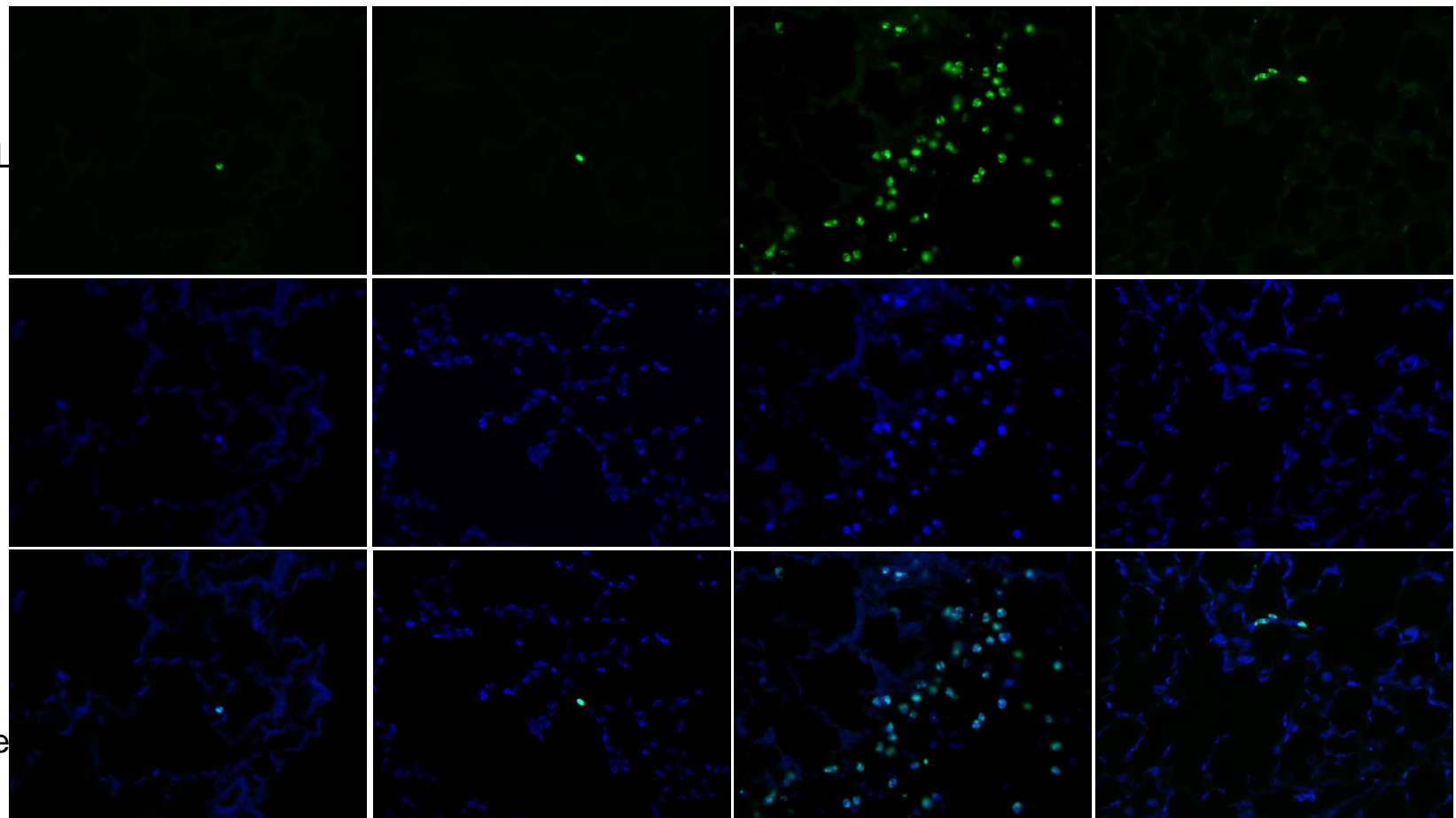
VILI

VILI/BTP2

TUNEL

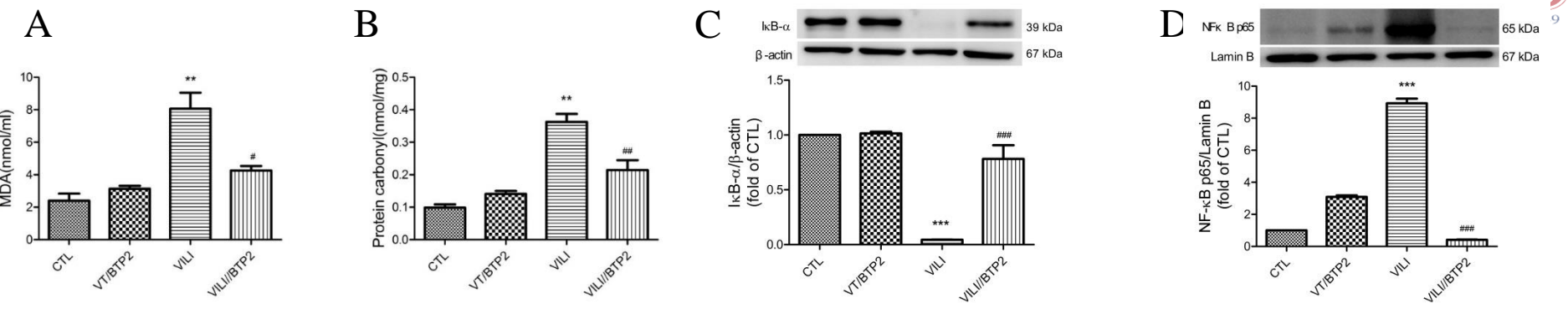
DAPI

Merge



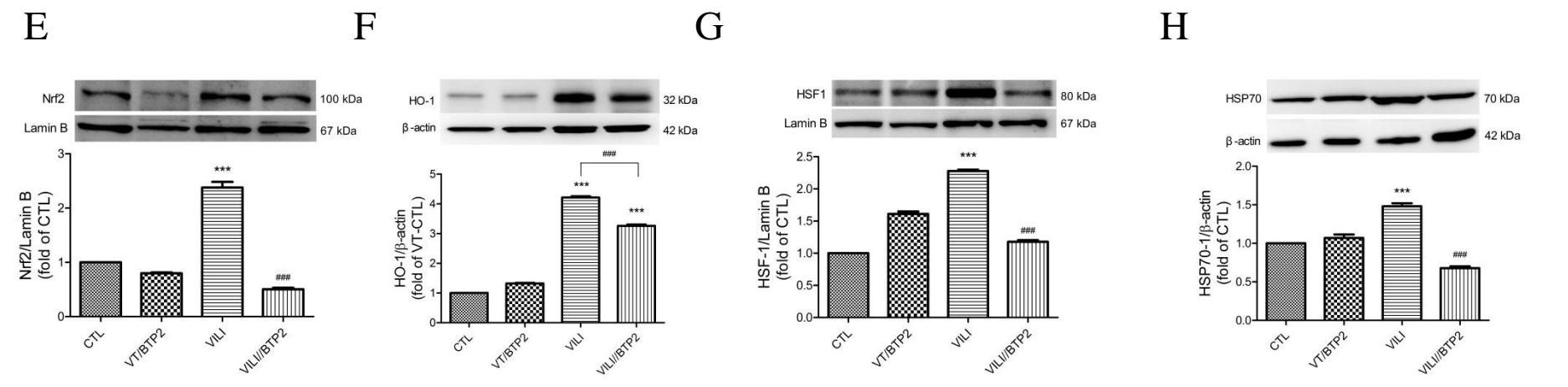
TUNEL assay

Fig. 4



Oxidative Stress

NF-κB pathway



Stress Response Proteins
NRF2/HO-1 and HSF-1/HSP70

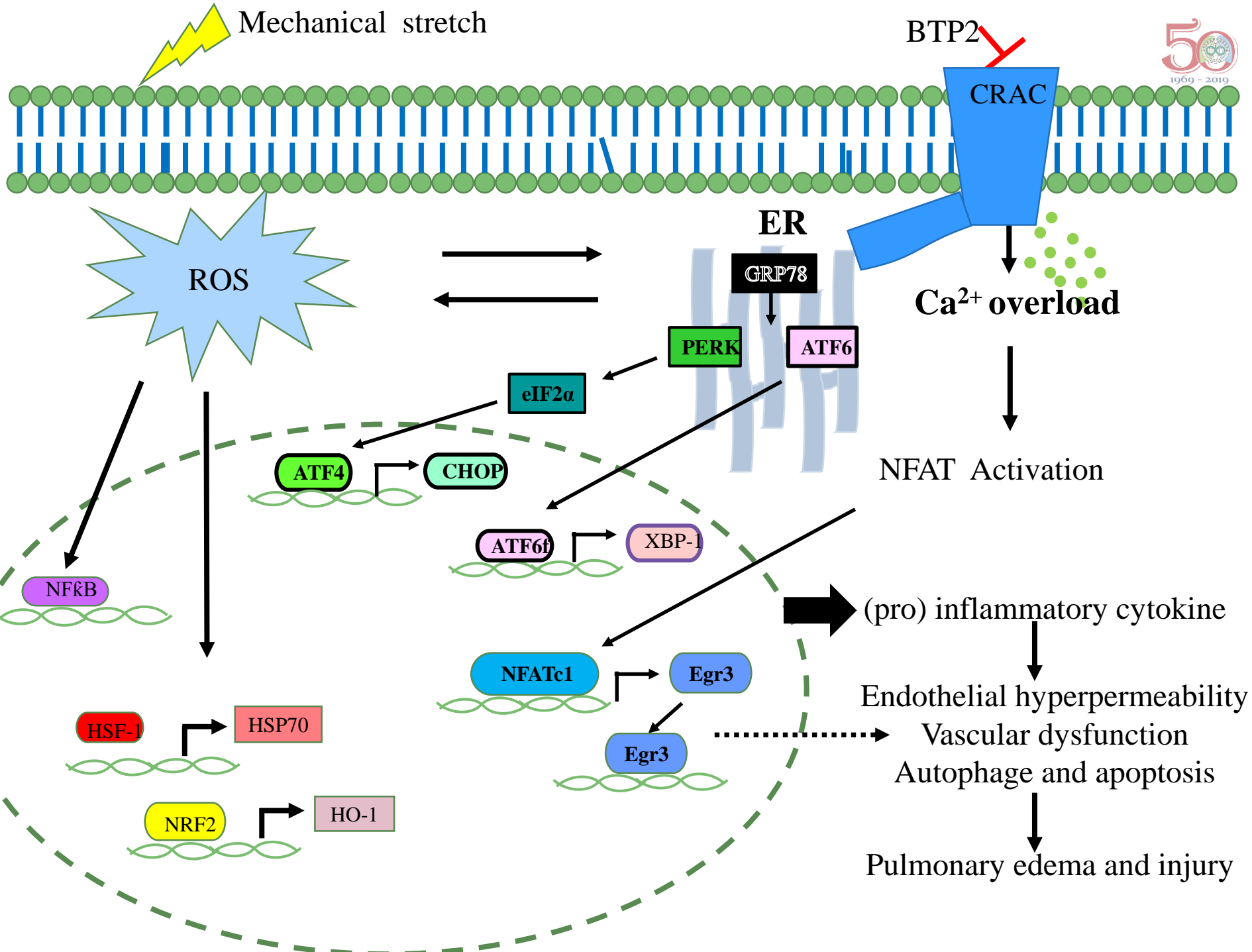
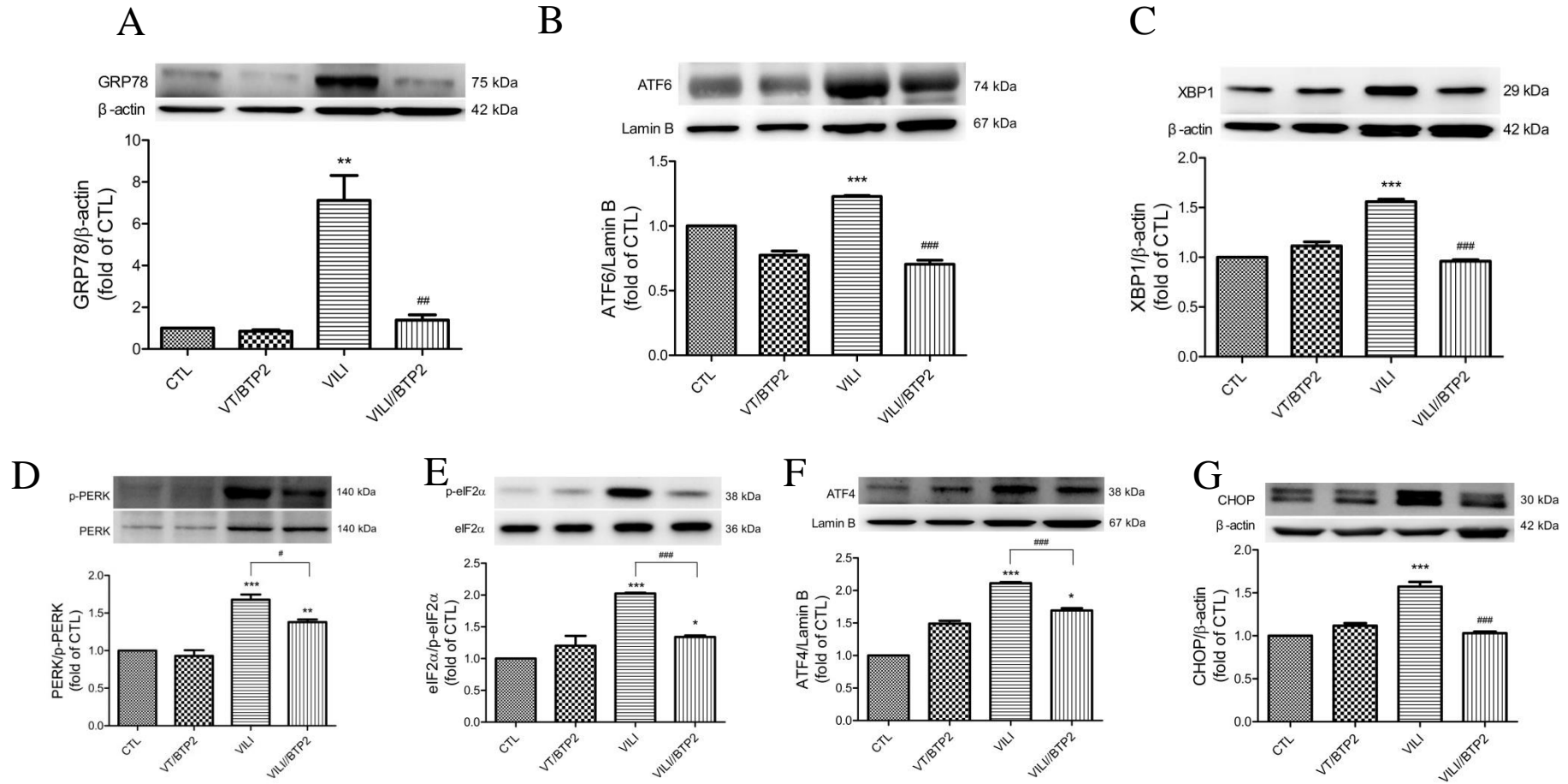
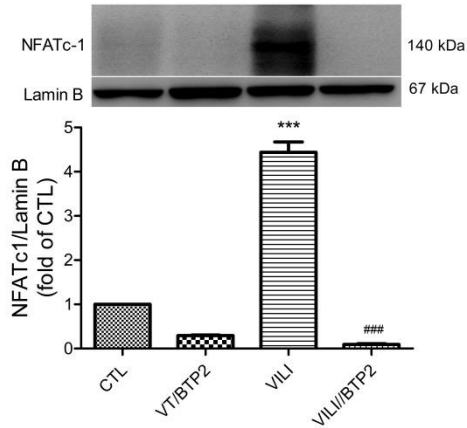


Fig. 5

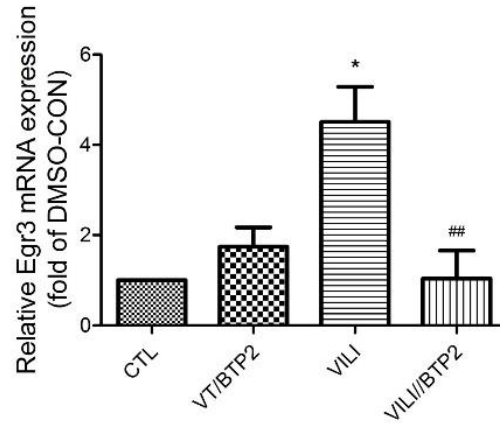


Lung injury: ER stress

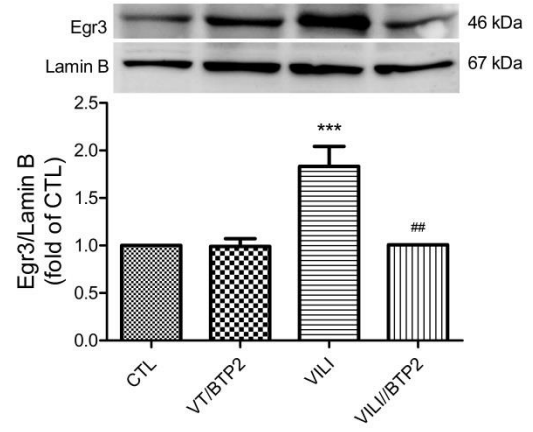
A



B

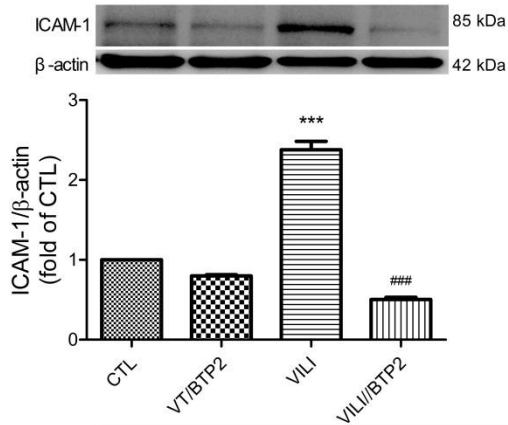


C

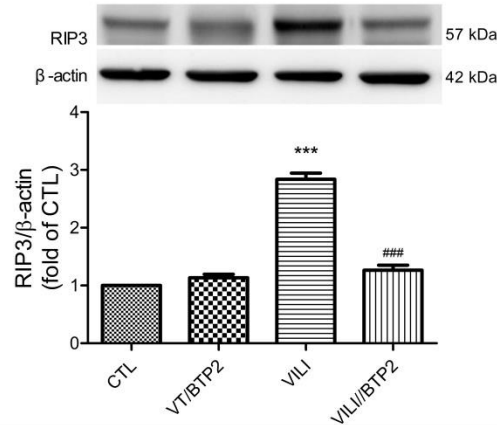


NFATc-1/Egr-3 pathway

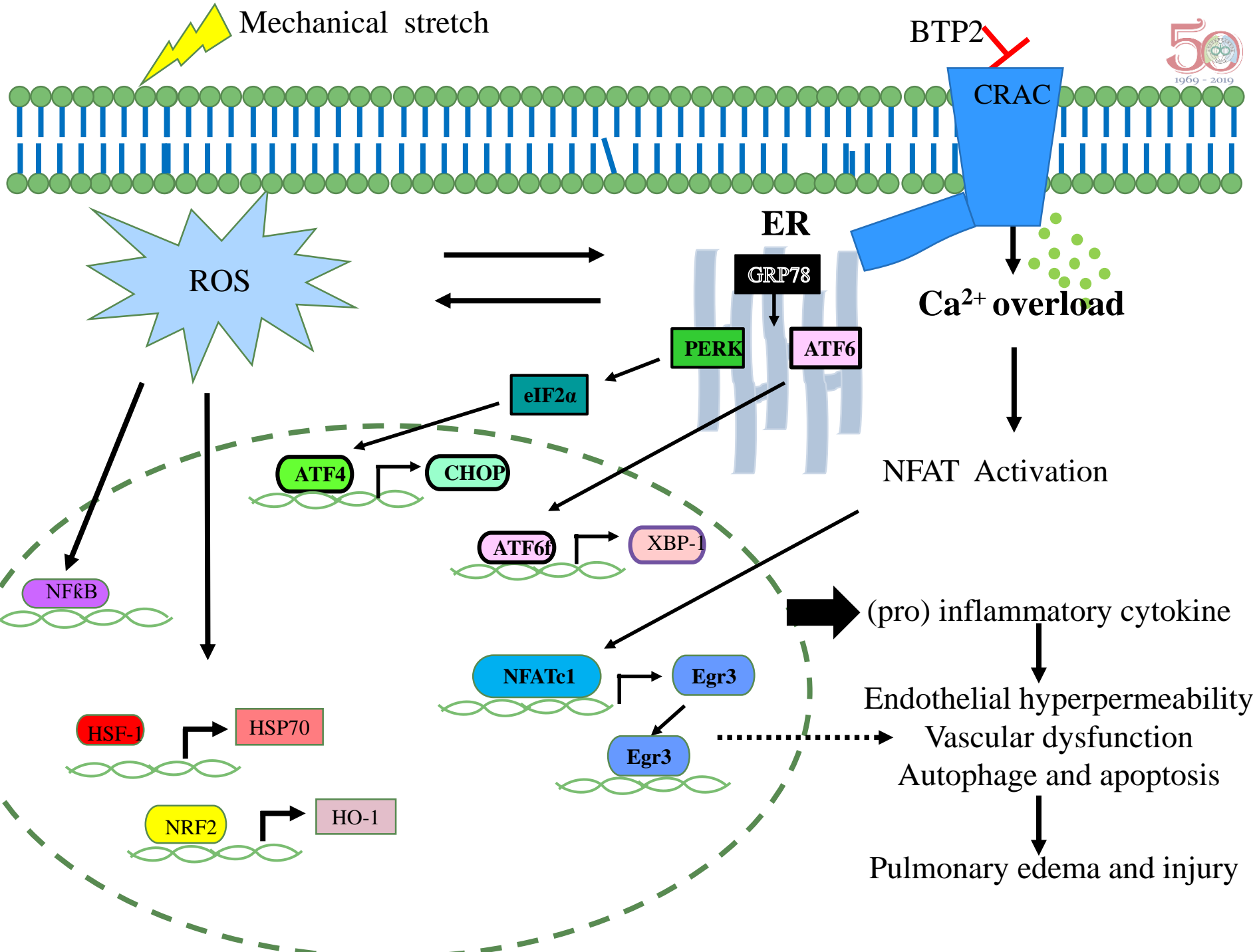
D



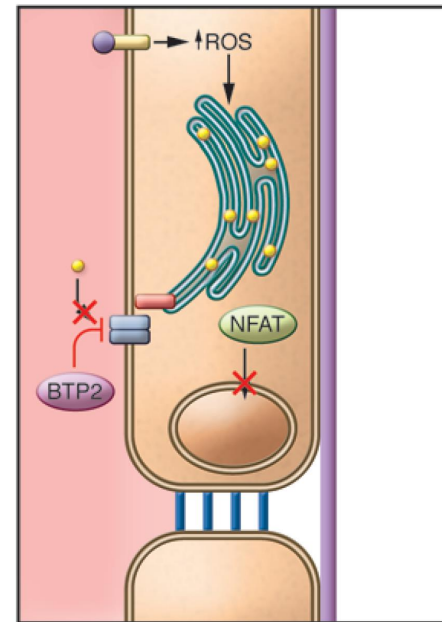
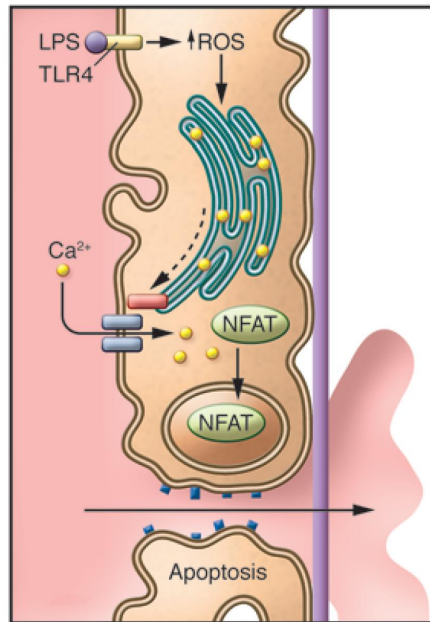
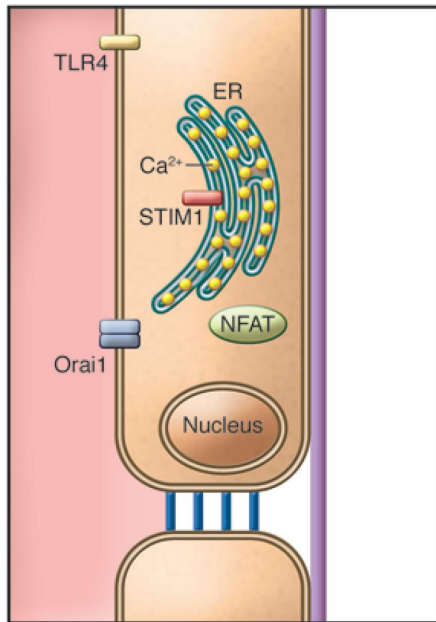
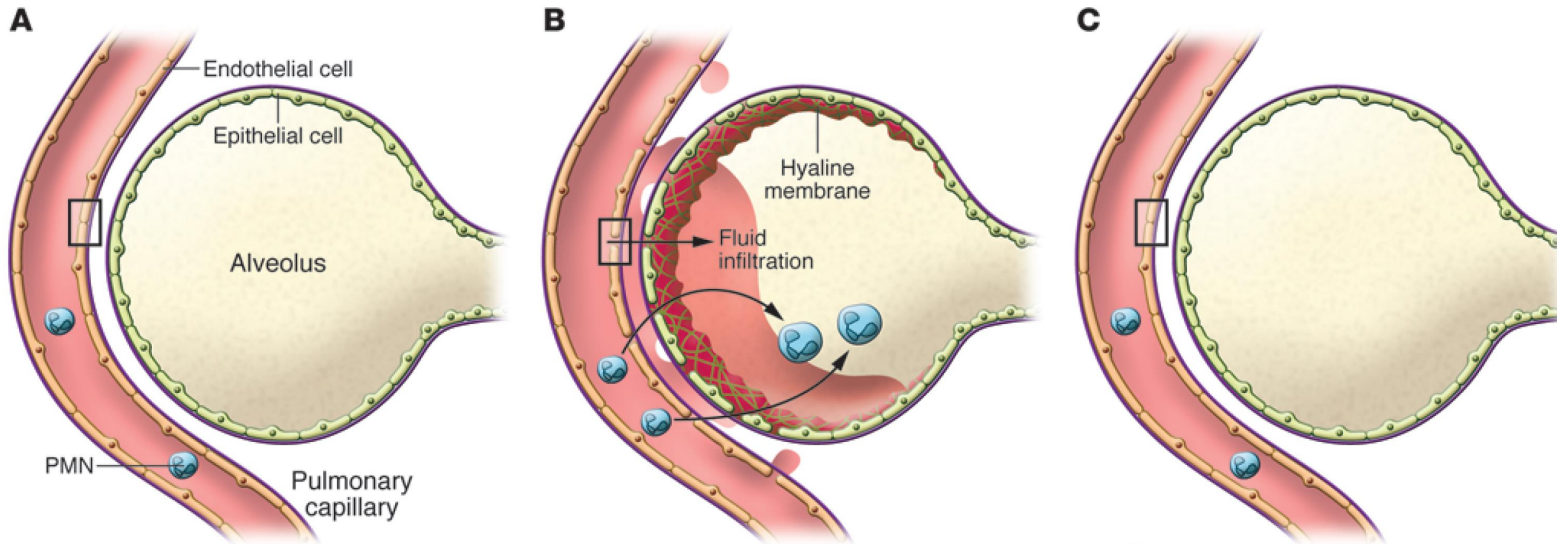
E



Endothelial cells injury markers

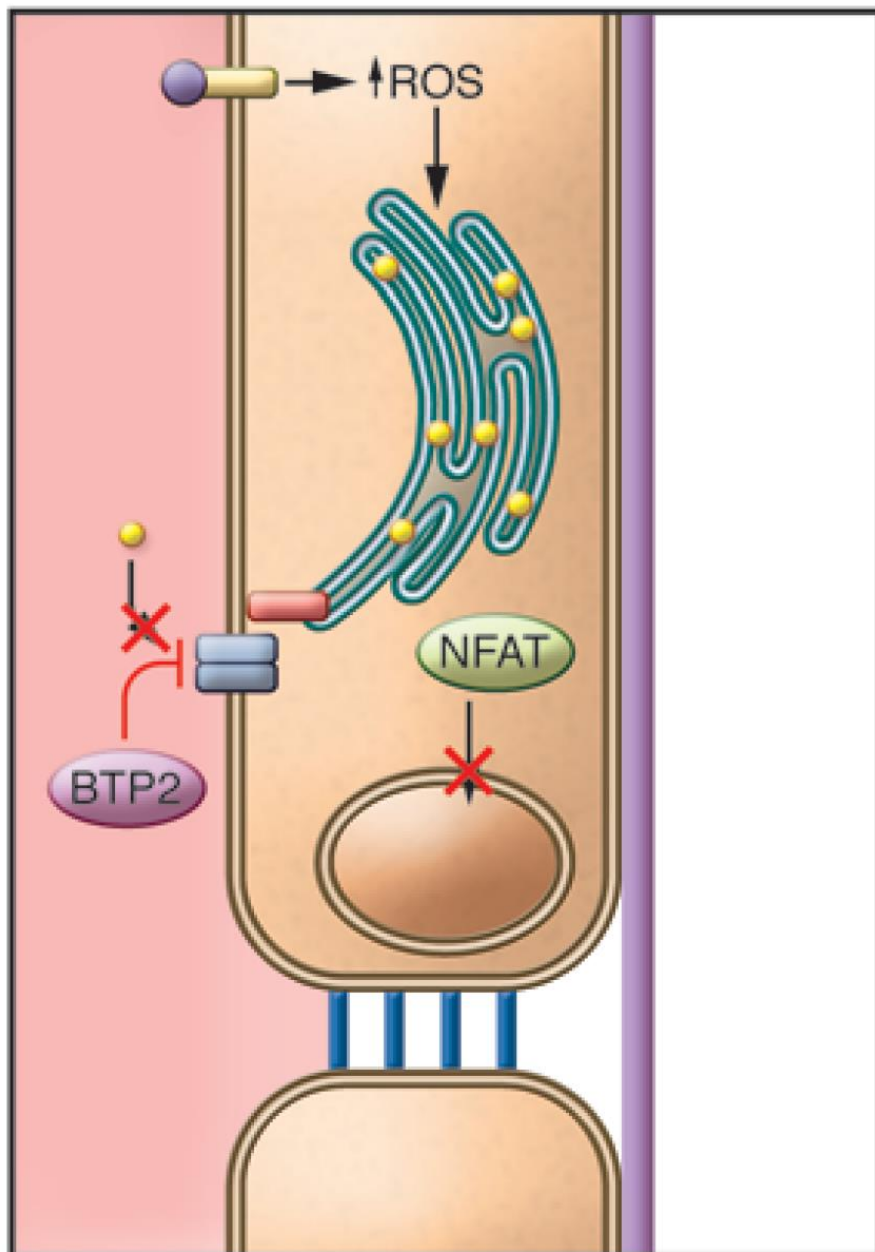


Species	CRAC channel inhibitory activity	Selectivity	Proposed mechanism of action
Lanthanides	Complete blockade at submicromolar concentration range [63]	Block other cationic ion channels, such as voltage-gated calcium channels and TRP channels [70,71]	Directly block ORAI1 [69]
SKF-96365	IC ₅₀ : 12 μM (I _{CRAC}) [72]	Suppresses voltage-gated calcium channels, nonselective cation channels and cyclic AMP-gated Cl ⁻ channels [72,73]	Has not yet been fully clarified
2-APB	Activates at low micromolar and inhibits at high micromolar [75,76]	Affect the activities of potassium channels, SERCA pumps, heat-gated recombinant TRPV1, TRPV2 and TRPV3 channels [90–92]	Might act on STIM1 multimerization, STIM1–ORAI1 interaction or the ORAI channel itself [85,86]
DPB162-AE	IC ₅₀ : 200 nM [93]	Relatively selective [93,95–97]	Probably acts directly on the coupling interface between SOAR and ORAI1 [93,94]
BTP2	Inhibited thapsigargin-induced Ca ²⁺ influx in Jurkat T cells with an IC ₅₀ of 100 nM [101]	Activates TRPM4 channels and inhibits the activities of TRPC3 and TRPC5 channels [105,106]	Has not yet been fully clarified
GSK-7975A	IC ₅₀ : 4 μM (I _{CRAC} in HEK293 cells) [111]	Potently blocks TRPV6 channels [111,113]	May act by altering the ORAI pore geometry [111]
Synta 66	IC ₅₀ : 1.4 μM (I _{CRAC} in RBL cells) [114,115]	Relatively selective [115]	Has not yet been fully clarified
ML-9	Reversibly inhibit SOCE with an IC ₅₀ of approximately 10 μM [116,117]	Inhibits MLCK [116]	Might target STIM1 [117]
DES	IC ₅₀ : 0.6 μM (I _{CRAC} in RBL cells) [118,119]	Activates estrogen receptors [118]	Might act on the extracellular regions on CRAC channel [118,119]
CAI	IC ₅₀ : ~0.5 μM (I _{CRAC} in HEK293 cells) [120,121]	Not very selective	Reduces the production of IP3 and depolarizes mitochondria [123–125]
RO2959	IC ₅₀ : 400 nM (I _{CRAC} in RBL-2H3 cells) [126]	Relatively selective [126]	Has not yet been fully clarified
Linoleic acid	Inhibit antigen- or thapsigargin-mediated SOCE in mast cells by acute addition at micromolar concentrations [127]	Has not yet been examined	Inhibits SOCE by affecting STIM1 oligomerization and subsequent STIM1/ORAI1 coupling [127]
1-phenyl-3-(1-phenylethyl) urea	Inhibits Ca ²⁺ influx with IC ₅₀ of ~3 μM in HEK293 cells [128]	Has not yet been examined	Targets ORAI1 [128]





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