2019台灣胸腔暨重症加護醫學會年會

THE ROLE OF EPIGENETICS IN HUMAN IMMUNE RESPONSES AGAINST MYCOBACTERIUM TUBERCULOSIS INFECTION 表觀遺傳變異在人類抵抗結核菌的免疫反應中 所扮演的角色

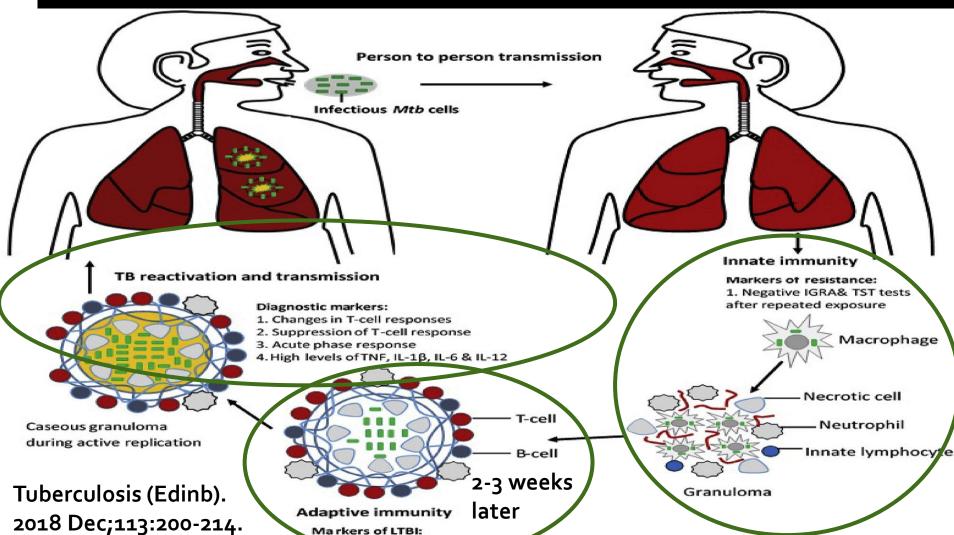
陳永哲醫師 高雄長庚醫院 呼吸胸腔科 Yung-Che Chen, M.D., PhD Division of Pulmonary and Critical Care Medicine, Kaohsiung Chang Gung Memorial Hospital 日期: 2019年12月07日(星期六)上午09:20~10:00 地點:高雄展覽館 3F 301 B

I have No commercial interest.

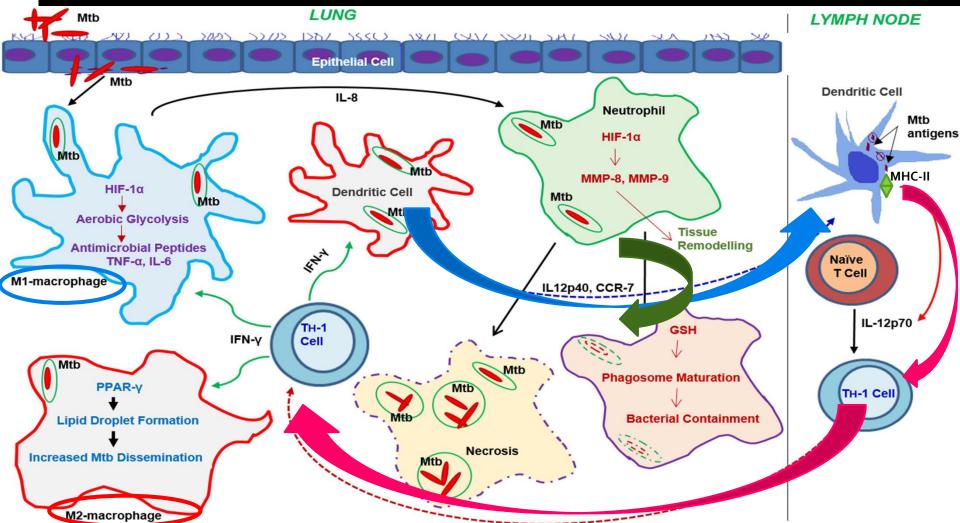
Contents

- Epigenetics-mediated immune responses in patients with active TB disease and macrophage infected with *M.TB* in vitro
 - Aberrant DNA methylation: global/gene-specific
 - Histone modification patterns and histone modifying enzymes
 - Non-coding RNA: microRNA
- Clinical application
 - Epigenetic predictors for BCG responders
 - Host-directed immunotherapy
 - Future perspectives

Mycobacterium tuberculosis (MTB) elicits the host innate immune response by recruiting macrophages/neutrophil, followed by adaptive immune response, mainly comprised of T-cells.



Dendritic cell (DCs) infected with Mtb migrate to the draining lymph nodes, driving Th-1 cell differentiation. The activated Th-1 cells migrate back to the lungs, producing IFN- γ and TNF- α , which activate macrophages leading to bacterial clearance. Front. Mol. Biosci.2019; 6:105.



Which factors play a major role in determining TB susceptibility or resistance?

- Mycobacterium TB:
 - rpf genes (rpfA-E), cAMP receptor proteins, Clp protease gene regulator (Rv2745c;clgR), DosR regulon,
 - Drug-Resistant genes: rpoB, katG, inhA, rrs, tlyA, gyrA or gyrB, atpE gene, Rv3547
- Comorbidities: HIV-TB co-infection, Diabetes, Malnutrition, Smoking and alcohol
- Environmental factors: drugs, toxin, diet, exercise, emotion, stress
- Host Genetic and Epigenetic factors: immune competence

Tuberculosis (Edinb). 2018 Dec;113:200-214.

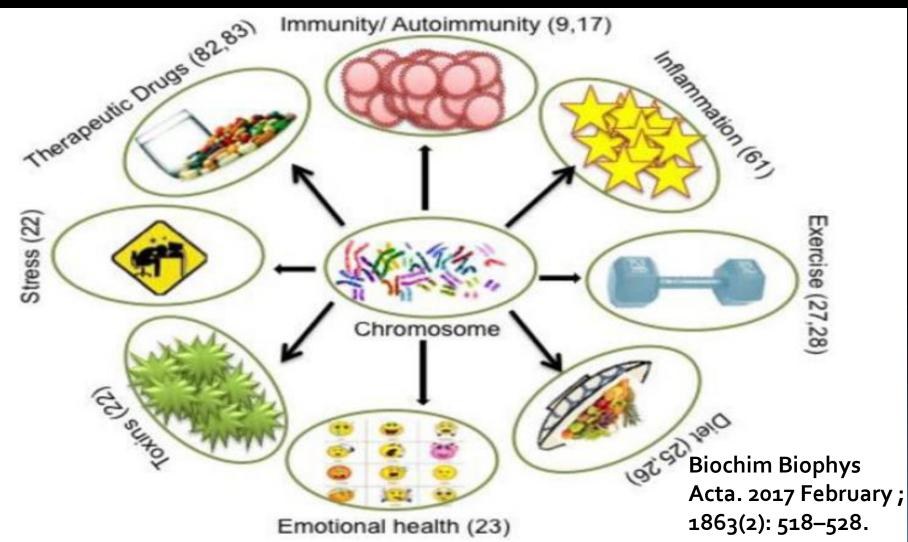
As in several other human diseases, the role of common genetic variants in pulmonary TB seems to be scarce. Lancet Infect Dis 2018;18: e64-75

- Most patients with latent TB infections (90–95%) never develop clinical disease.
- In household studies, 30–50% of contacts with heavy short term exposure do not become infected.
- Twins study under comparable environmental exposure and social conditions
 - Higher concordance of TB in monozygotic than in dizygotic
 - Heritability of TST response at 71% and IGRA response at 39%
- Independent gene association studies: an absence of consistency and replication
- GWAS: Common variants might have a little effect on individual predisposition to adult pulmonary TB, at least when considered as a single homogeneous phenotype.

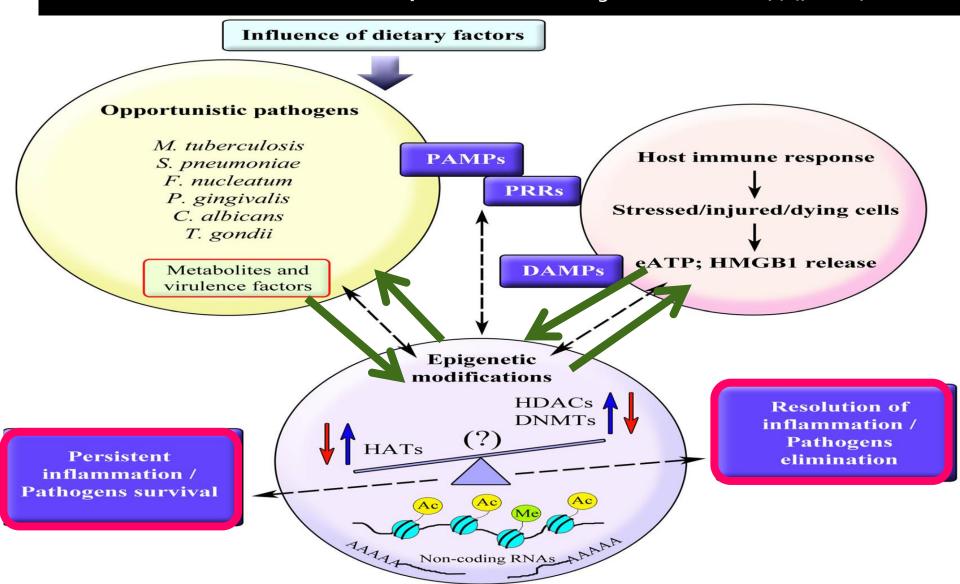
A real challenge is to associate candidate genetic variants with a biologically plausible mechanism that explains the epidemiological data for TB in which only 10% of the infected individuals will develop active TB.

Meta-Analysis	Odds Ratio	Number of studies	Risk of active TB	Race - dependent
mannose-binding lectin gene (MBL2)	0.42-2.7	22	Decreased or increased	Africans, Americans, Europeans
vitamin D receptor (VDR)	0.87-0.92	54	Decreased	South Asians, Caucasians
Interleukin-10/17/18 (IL-10/17/18)	0.53-1.37/0.64- 1.36/1.17-1.43	28/11/8	Decreased or increased	Asians, Caucasians.
Toll-like receptor (TLR) 1/2/6	0.61-5.82	16	Increased or decreased	Asians, Europeans, Africans, American Hispanics.
TNF-α		31	Increased	East Asians
IFN-γ/IFNGR1	1.51-1.56/1.24	19/6	Increased	Africans
HLA-Class : DRB1/DOB1/DOA1	0.5-2.27	11	Decreased or	Caucasians

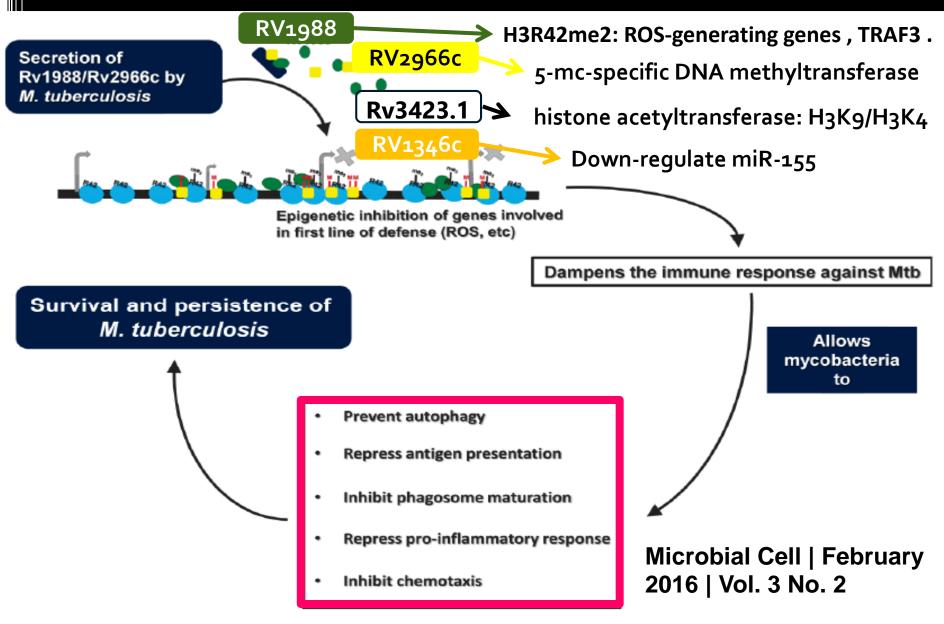
Environmental factors and epigenetic modulation in humans: Various sources presents in the environment regulates epigenetic parameters on humans.

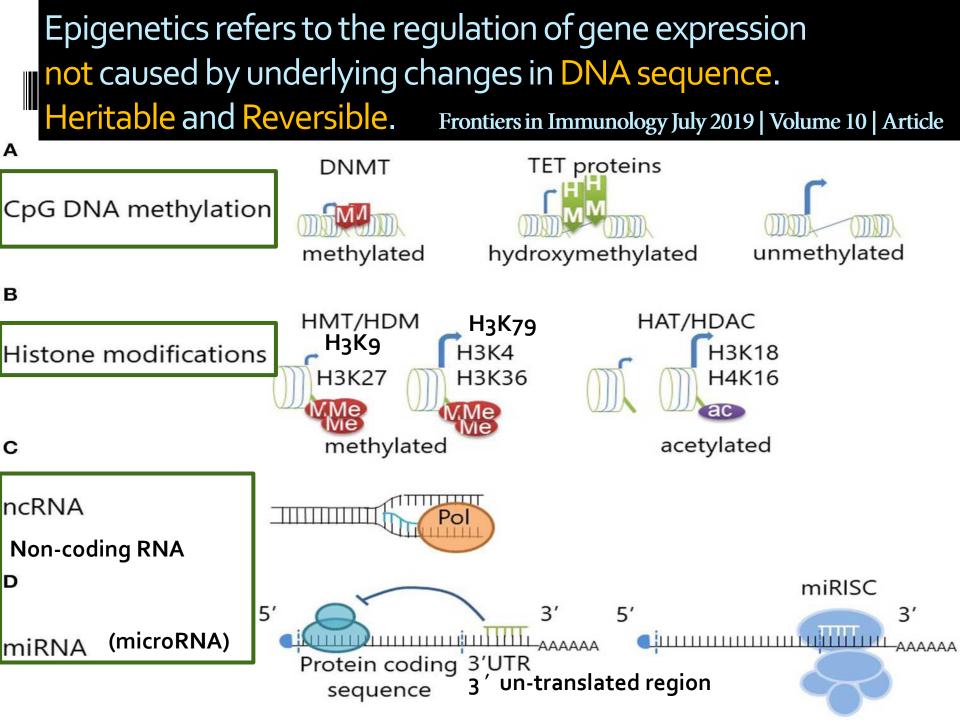


Bi-directional relationship between epigenetic modifications induced by colonizing pathogens / the host immune response. Pathogens and Disease, 74, 2016, ftwo82



M. TB uses Rv1988/Rv2966c/Rv3423.1/RV1346c to hijack the host transcriptional machinery.

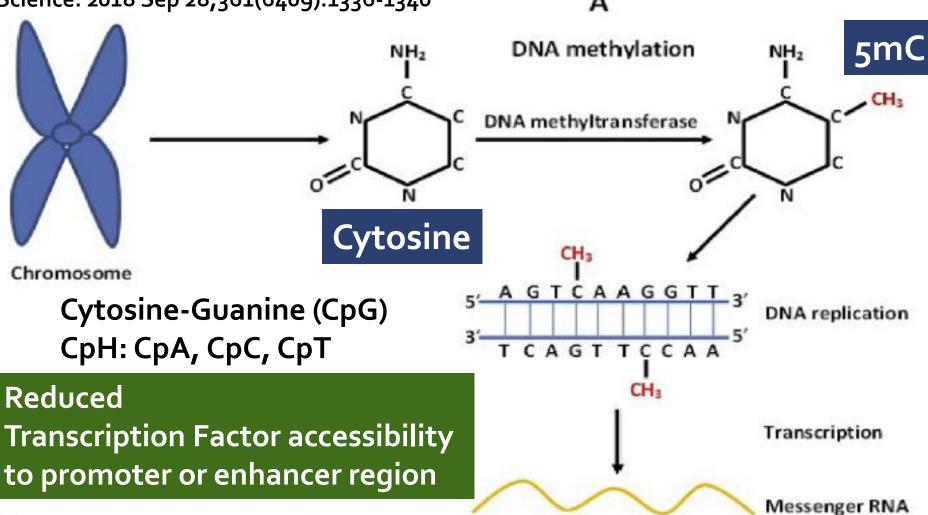


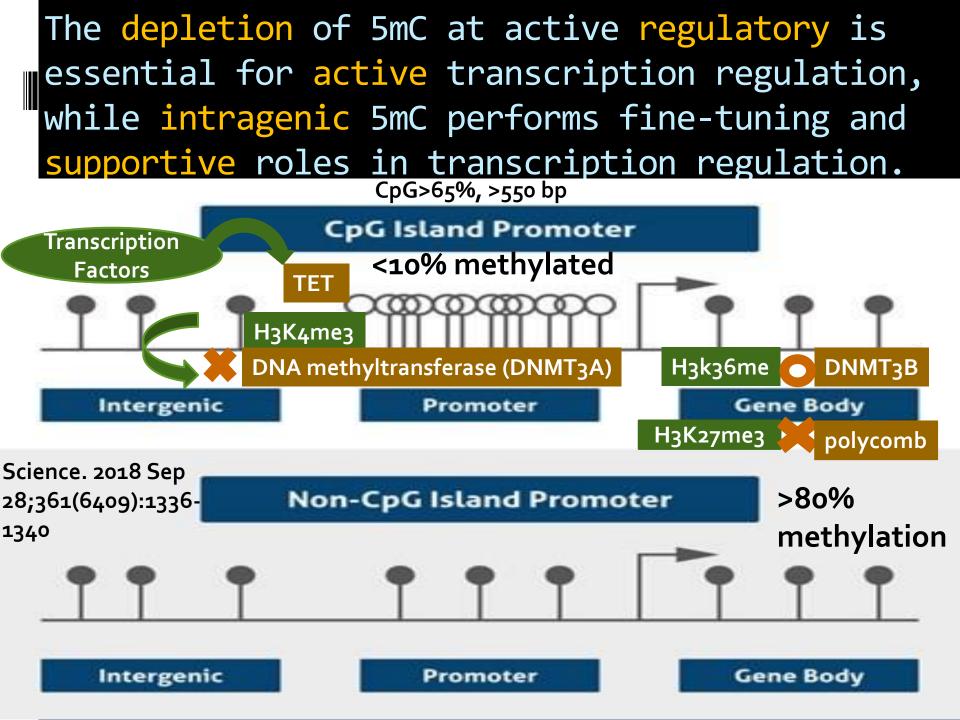


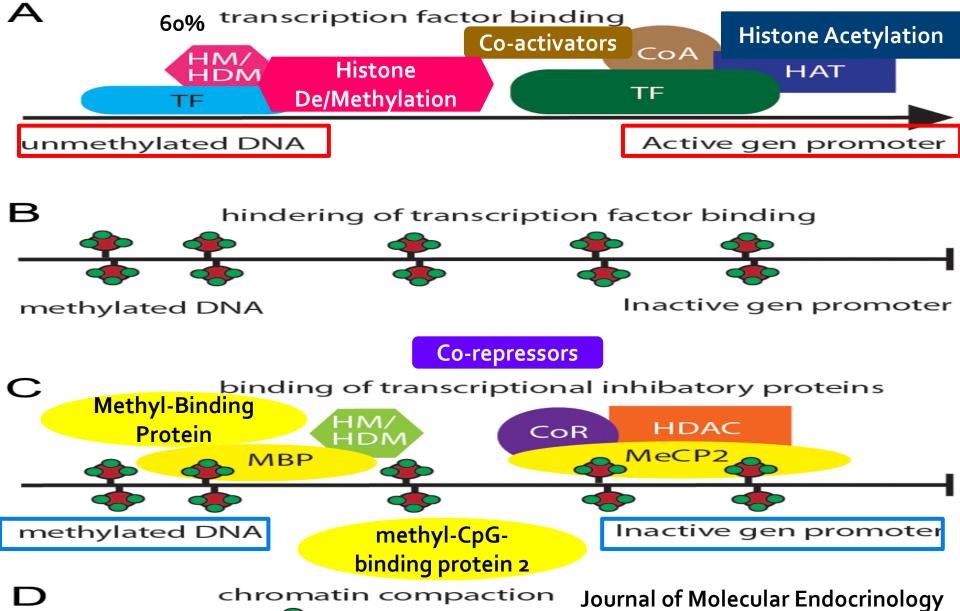
Aberrant DNA methylation of the immune-related genes in active TB disease or in response to *M.tb* infection

Methylation of DNA promoter or enhancer regions generally results in transcriptional silencing or repression.

Science. 2018 Sep 28;361(6409):1336-1340





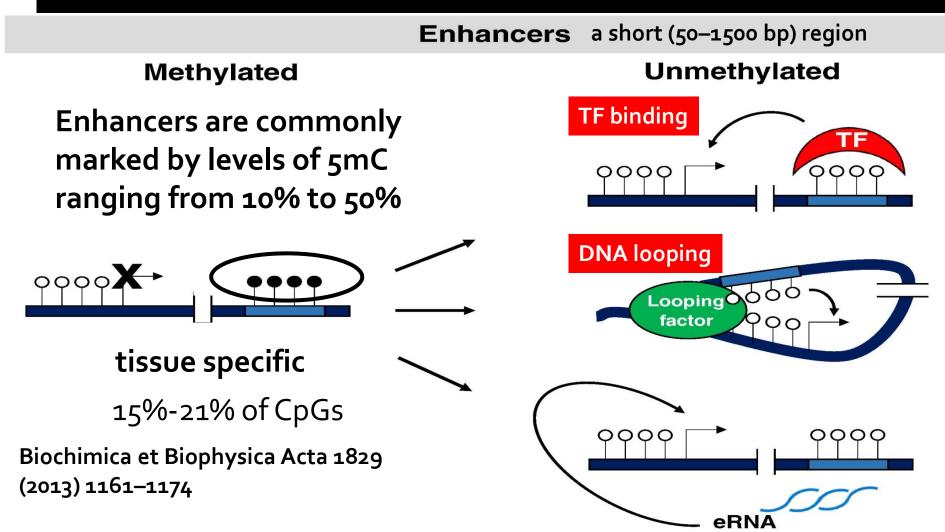


methylated DNA

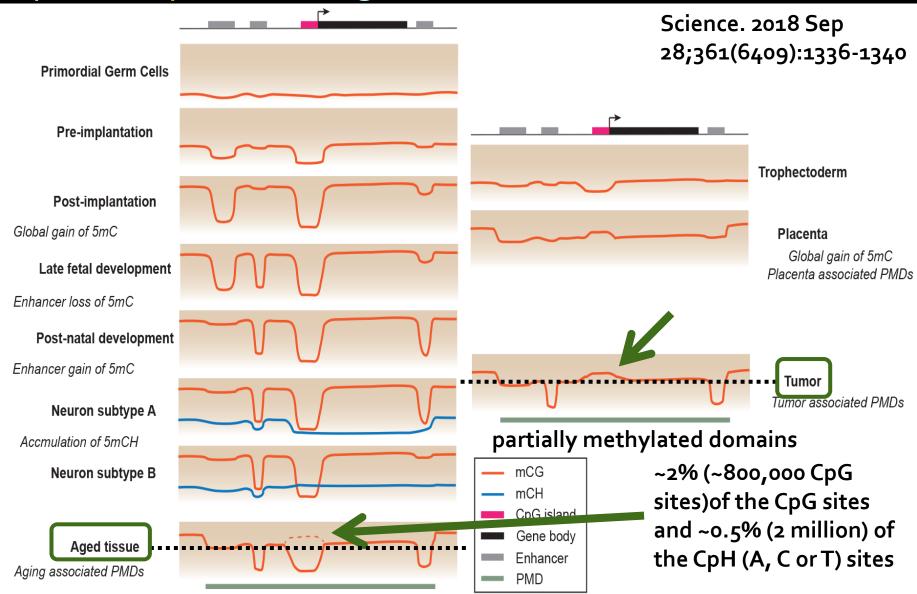
2018; 17-0189:R39

Inactive gen promoter

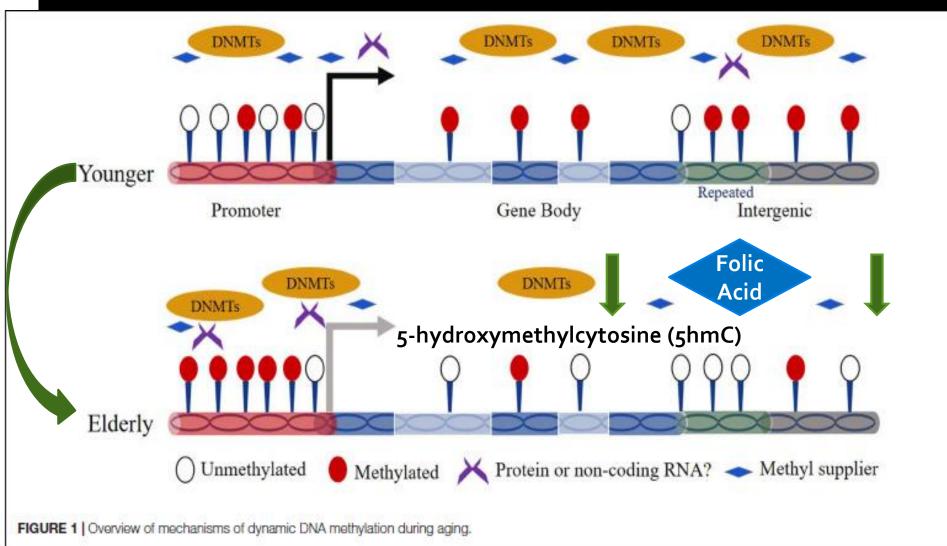
Dynamically methylated CpGs cluster into over 1 million tissue specific differentially methylated regions (DMRs), which are distal to TSS and overlap with enhancers.

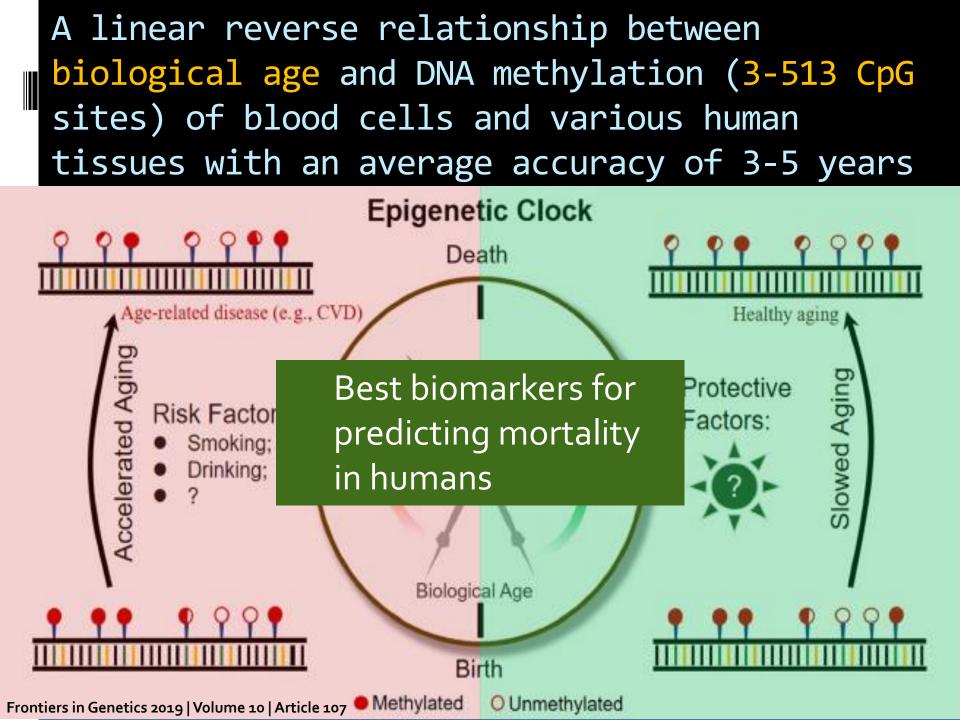


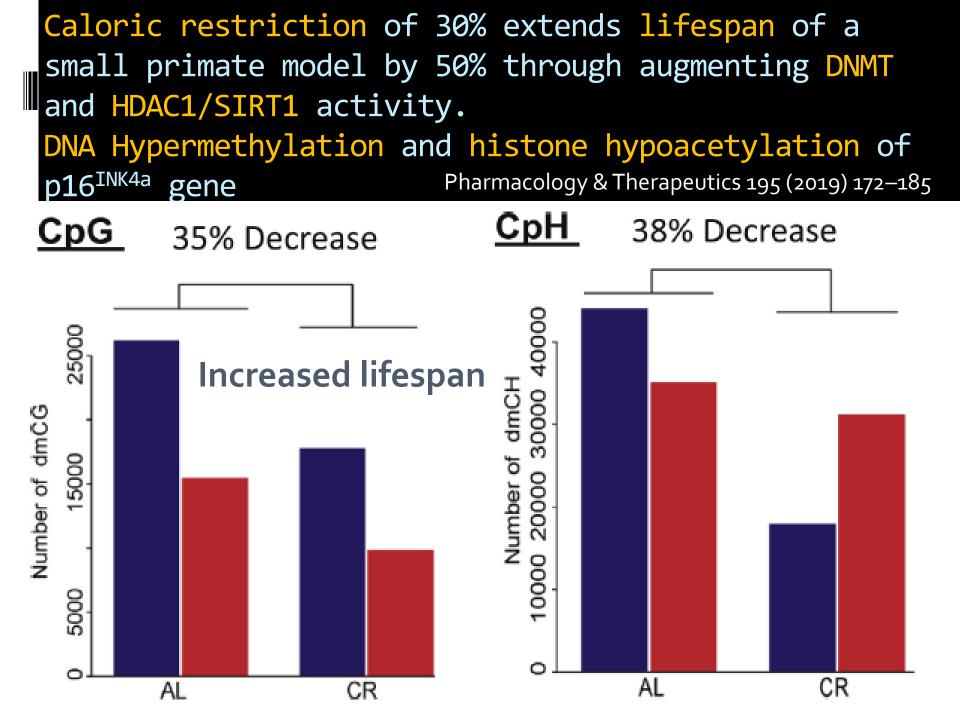
Both Aged tissues and Tumor methylomes have global hypomethylation and localized hypermethylation over specific promoter regions.



The global reduced methylation content can be caused by down-regulated expression of DNMTs or insufficient supply of folic acid in elderly subjects. Frontiers in Genetics 2019 | Volume 10 | Article 107







DNA methylation state during innate and adaptive immune cell development, differentiation, and function.

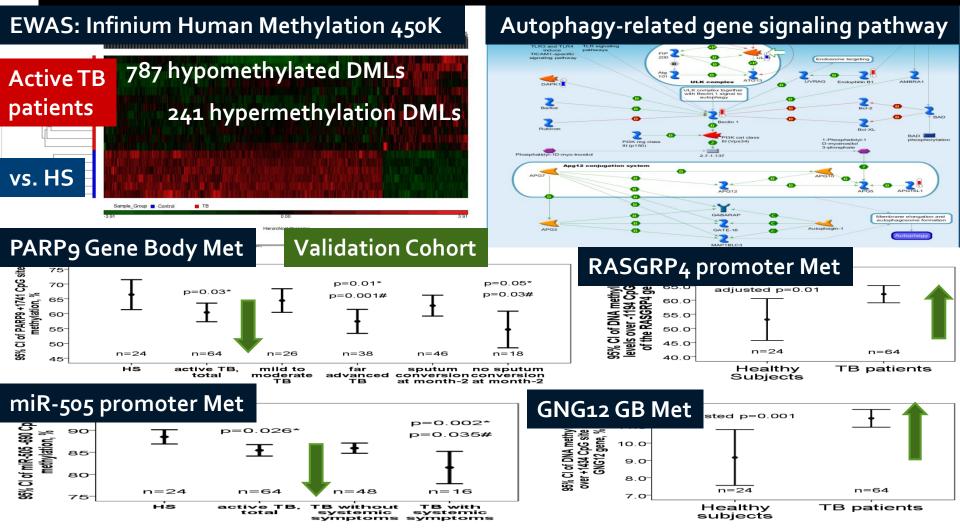
Translational Research 2019; 204:118

Immune cell	DNA methylation-mediated changes
macrophage	myeloid over lymphoid differentiation, monocyte- to-macrophage differentiation, polarization
CD4+T lymphocytes	differentiation into distinct phenotypes
CD8+T lymphocytes	cytolytic function , de-differentiates into memory
B cell	Differentiation, activation and plasma cell differentiation.
Regulatory T lymphocytes	Differentiation, suppressive capability, Th2-biased subset

Association between gene-specific aberrant DNA methylation and active TB disease

Gene Name	Region	DNA MET change	Gene or protein change	Model	Clinical Outcome
Vitamin D receptor	Gene body (Exon, 3'UTR)	Hyper-Me (9 CpG sites)	Decreased GE of AKT, GSK38,FOXO1	Active TB patients vs. HS	
Toll-like receptor 2	promoter	Hyper-Me (5 CpG sites)	Depressed TLR2	Active TB patients vs. HS	Drug- resistant TB
IL 18 receptor 1	promoter	Hyper-Me (1 methyl- SNP)	Depressed IL18R1	Active TB patients vs. HS	
CYP27A1	promoter	Нуро-Ме	Decreased 1,25-dihydroxy vitamin D	Active TB patients vs. HS	
	ol. 2011 ; 72(3 4;69(6):546-5		PLoS One. 2014 ;9(10):e110734 Thorac Dis 2017;9(11):4353-4357		

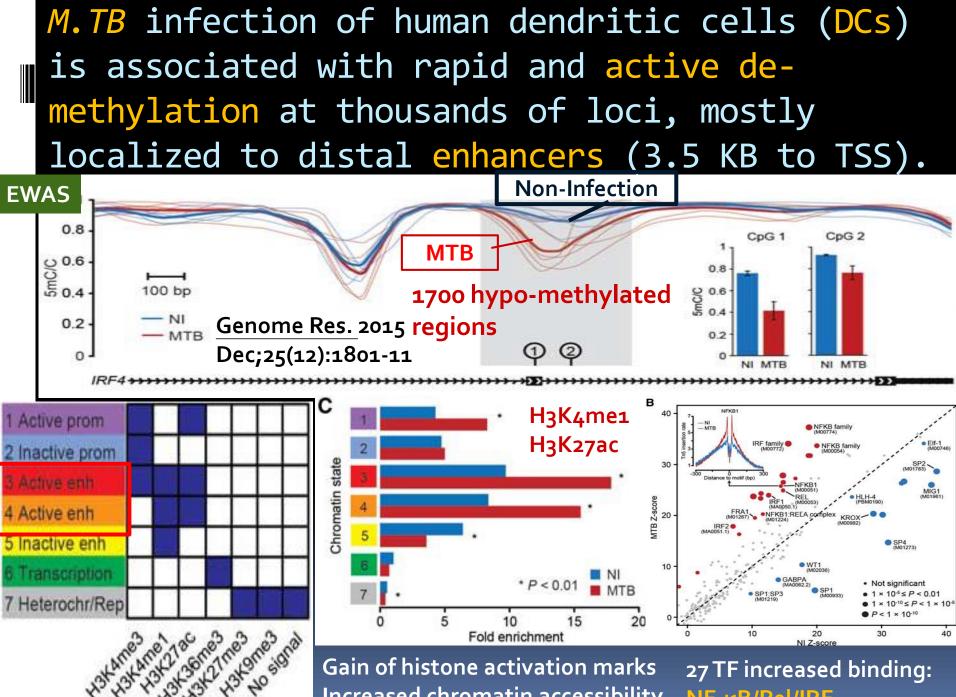
Whole genome DNA methylation analysis of active pulmonary TB disease identifies novel epigenotypes: *PARP9/miR505/RASGRP4/GNG12* Met and clinical phenotypes (under review at "J. Infection")



Gene-specific aberrant DNA methylation in response to *M.TB* infection in vitro or in vivo

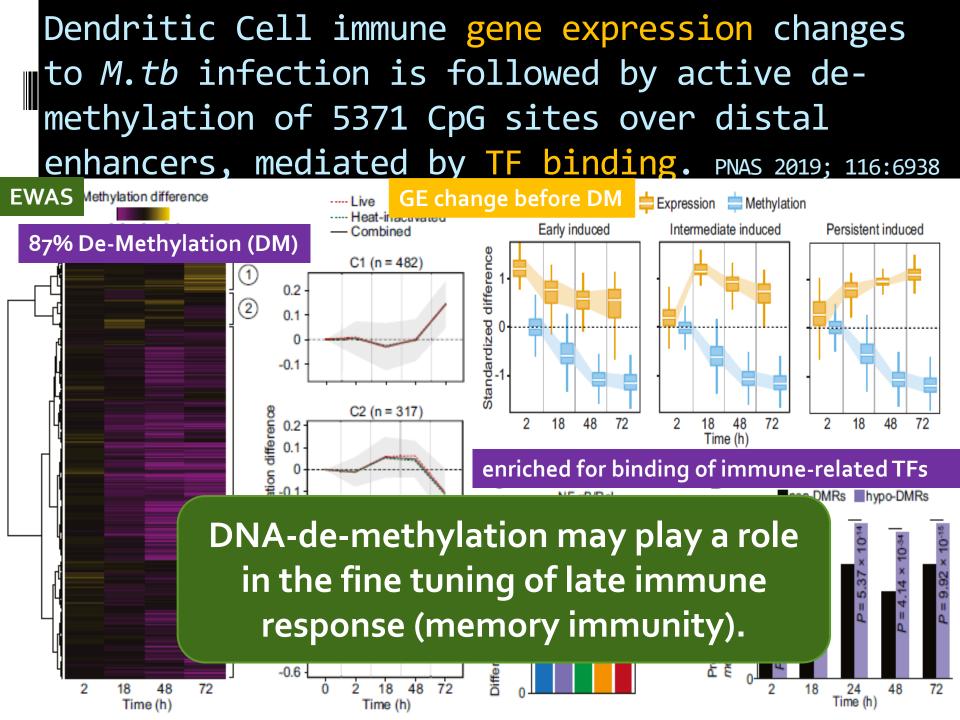
Gene Name	Region	DNA MET	GE	Model	Functional Outcome
IL6R	promoter	Hyper-Me		Beijing/Wild MTB-infected THP-1	
NLRP3	promoter	De- methylation	NLRP3 activation	Mtb H37Rv- infected THP-1	Increased inflammatory cytokines
CD82/K Al1	promoter	De- methylation	RUNX1- binding induced CD82 activation	MTB-infected THP-1 /BMDMs/Mice	Decreased inflammatory cytokines/ phagosome maturation, enhanced MTB survival

Biomed Res Int. 2016;2016:4323281. Tuberculosis (Edinb). 2016;98:139-48. Experimental & Molecular Medicine (2018) 50:62



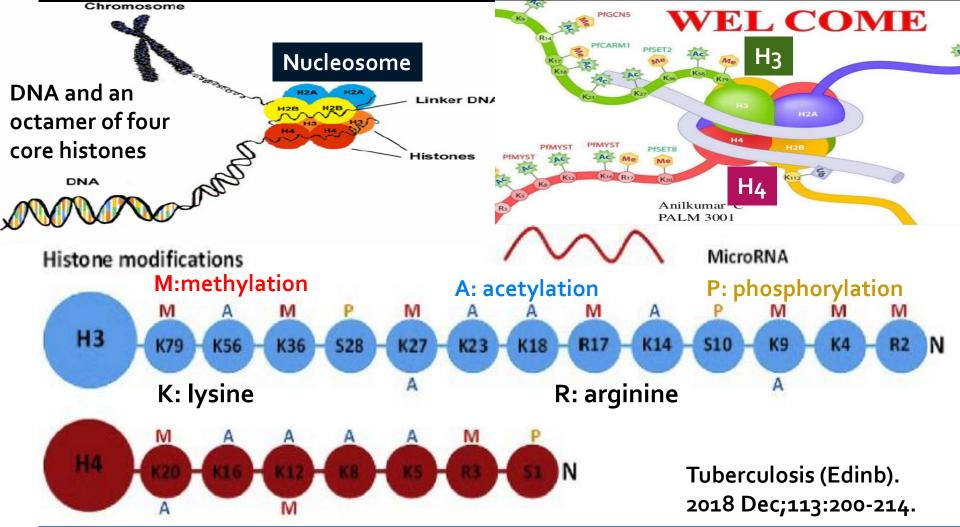
Increased chromatin accessibility

27 TF increased binding: NF-kB/Rel/IRF

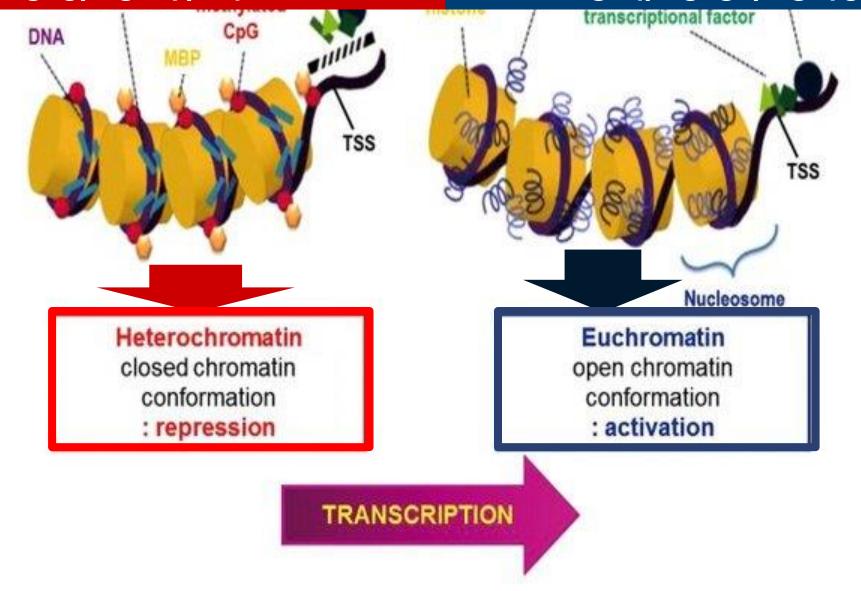


Aberrant histone modifications and histone modifying enzymes in active TB disease or in response to *M.TB* infection

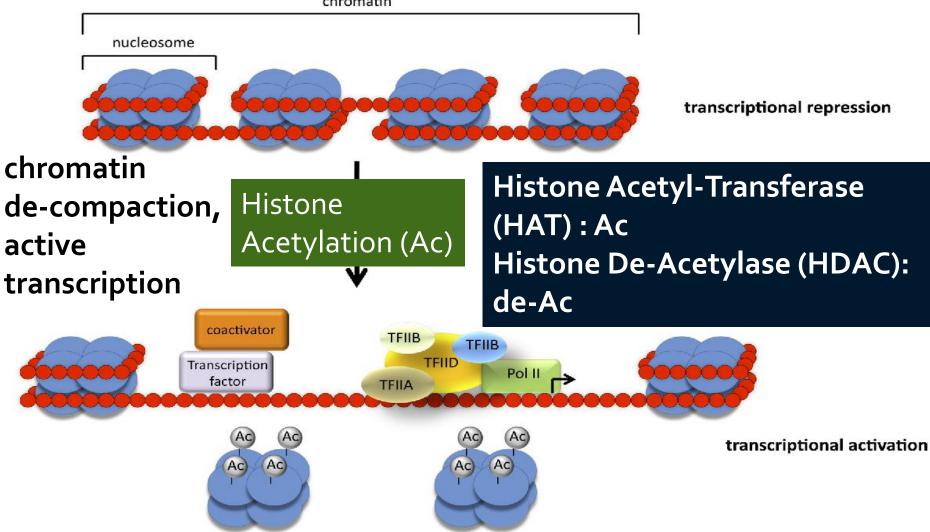
Amino-terminal tails of the core histones (H3, H4) can be posttranslational covalent modified by addition of methyl (red), acetyl (blue) or phosphoryl moiety (orange).



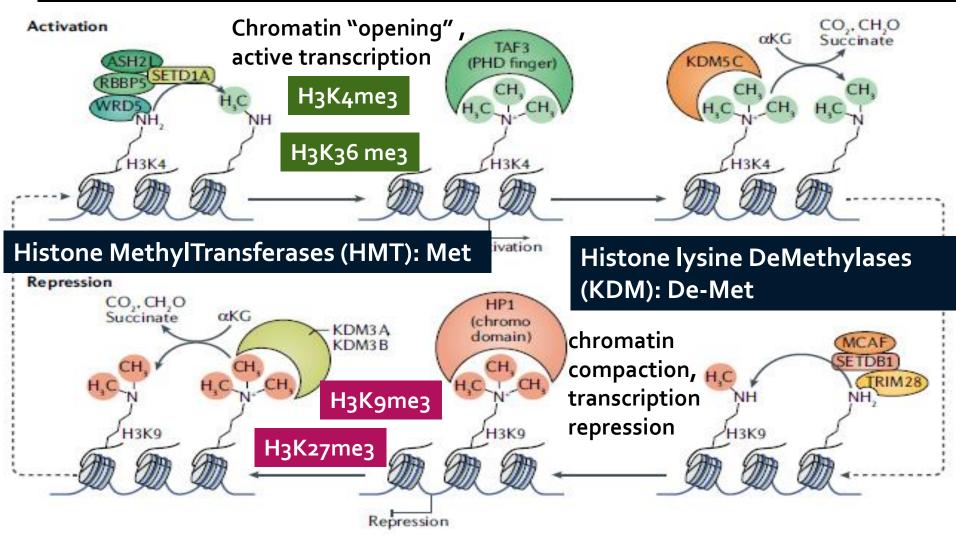
Low Histone Acetylation High levels of Methylation over H3K9, H3K27, H4K20 High Histone Acetylation High levels of Methylation over H₃K₄, H₃K₃6, H₃K₇₉,



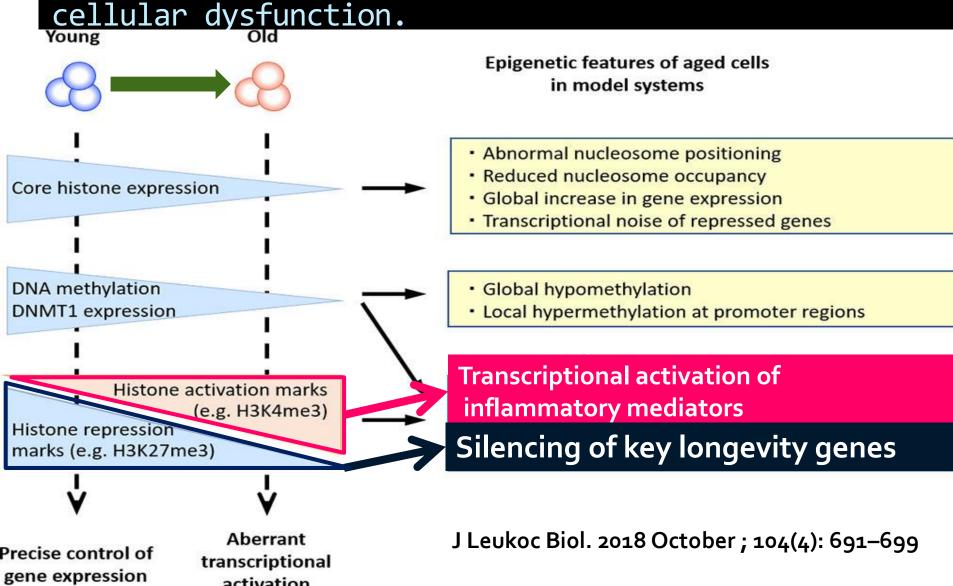
Histone acetylation exposes promoter and coding regions to transcriptional regulators, including RNA polymerase (Pol II) and the various isoforms of the basal transcription factors (TFIIs), which results in transcriptional activation.



Gene-activating H₃ Lys₄ tri-methyl (H₃K₄me₃)/H₃K₃6me₃/H₃K₇₉ mark at the promoters of various genes. Gene-repressive states established by the deposition at the promoters of H₃K₉me₃/H₃K₂₇me₃. Nature Reviews | MolECulaR CEII BioloGy 20 | 2019 | 625



Changes in chromatin structure due to altered histone expression, histone modifications and DNA methylation occur with aging and contribute to cellular dysfunction



Association between histone modification/modifying enzymes and active TB

		y	0 ,		
Histone	Attribu te	Change	Model	Mechanism	Clinical Outcome
H3K14 Ac	Global	Hypo- acetylation (Ac)	PBMCs from Active TB patients vs. HS	TNF-α /IL12B promoter- specific H3K14 hypo-Ac	lower one-year survival
H3K27 Me	Global	Hyper- methylatio n (me2/me3)	PBMCs from Active TB patients vs. HS		bacterial burden, symptom
HDAC1	Non- specific de-Ac	Increased	PBMCs from Active TB patients vs. HS		Reversed with anti-TB Treatment
KDM6B	H3K27me 3 De-Me	Decreased	PBMCs from Active TB patients vs. HS		Reversed with anti-TB Treatment

Am J Transl Res. 2017 Apr 15;9(4):1943-1955.

Altered expressions of Histone modifying enzymes in response to *M.tb* infection in vitro

Histone modifying enzymes	Attribut e	Regul ation	Model	Mechanism	Functional Outcome
HDAC1	Non- specific de- Ac	Up	MTB H37Rv infection	Decreased H3Ac over the IL-12B promoter	Increased survival of intracellular MTB
HDAC 6	Non- specific de- Ac	Up	Mtb H37Ra infection in mice/THP-1 cells	Increased IL-10 expression	Increased MTB growth
JMJD3 (KDM6B)	H3K27me3 (repressive) De-Me	Up	MTB H ₃₇ Rv infected macrophage of mice	Augmenting NOTCH1-PI3K- mTOR-NF-κB signaling	Foamy Macrophage, M2 polarization
SET8	H4K20 monometh ylase	Up	Mtb-infected macrophages	Induce NQO1- TRXR1	M2 polarization

Tuberculosis (Edinb). 2018 Jan;108:118-123. PLoS Pathog. 2016 Aug 17;12(8):e1005814.

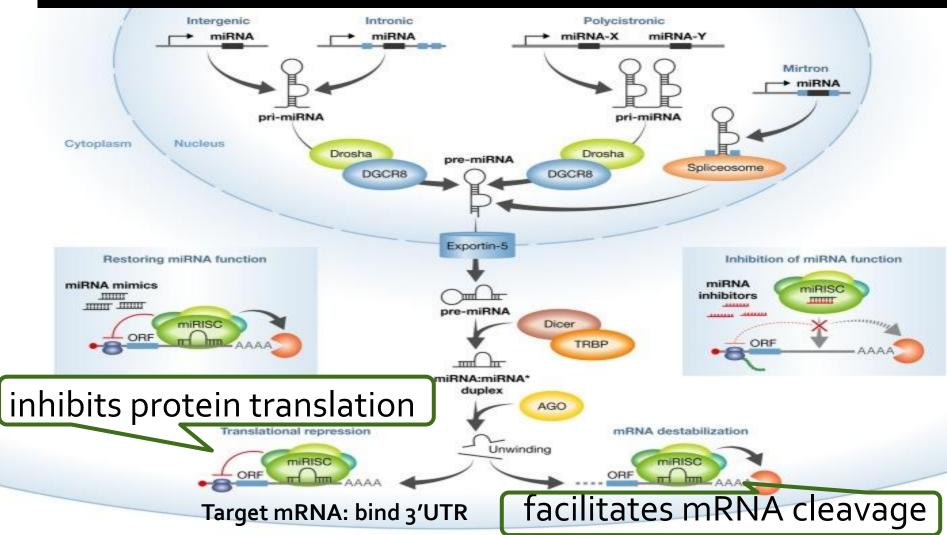
J Infect Dis. 2017 Aug 15;216(4):477-488.

Altered Histone modification patterns in response to M.tb infection in vitro

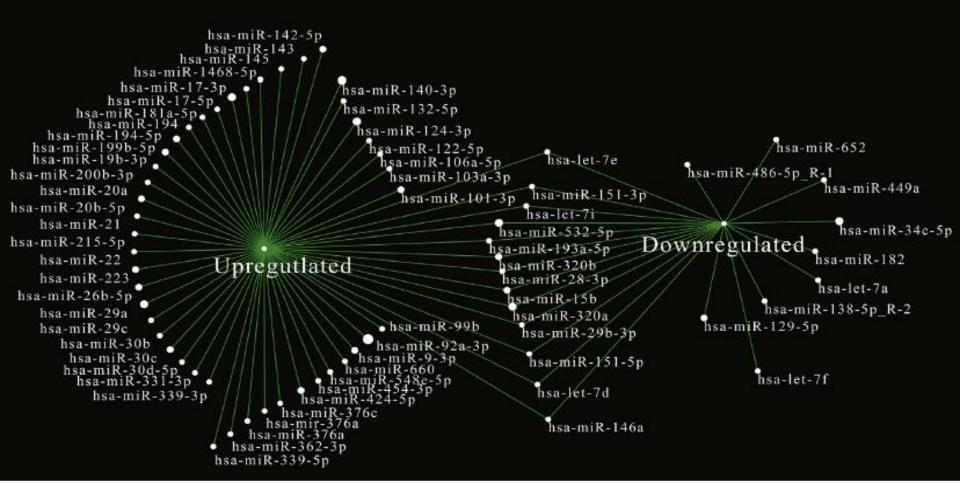
Histone modificati on	Change	Model	Mechanism	Functional Outcome		
H3 and H4	Hyper-Ac	Mtb-infected macrophages /epithelial cells	Increased RNA Pol II binding to MMP-1 promoter	increased MMP-1 secretion		
H3K9Me (repressive)	Hyper- Me	Mtb-infected macrophages	down-regulated the expression of CIITA/MHC-II	inhibits antigen presentation		
H3K4Me (active mark)	Hypo- Me/Ac	ESAT-6 – stimulated macrophage	Inhibit class II transactivator (CIITA)	Inhibit MHC II		
H3R42Me (repressive)	Hyper- Me	H37Rv Mtb- infected THP1	Rv1988 repress NOX1, NOX4, NOS2, TNFAIP2, and lincRNA, ENSG00000250584	Increased MTB survival		
Adv Protain Cham Struct Rial 2017:106:71 Erant Immunal 2017 May 24-8:602						

Adv Protein Chem Struct Biol. 2017;106:71 J Biol Chem. 2017 ;292(17):6855-6868. Front Immunol. 2017 May 24;8:602.

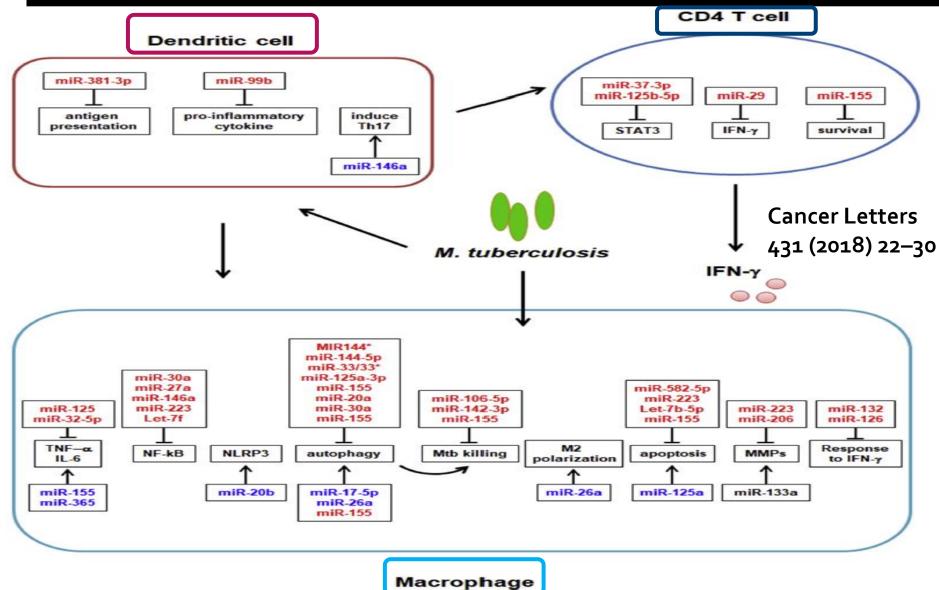
Altered microRNA (miR) expressions in active TB disease or in response to *M.TB* infection MicroRNAs (miRNAs) are small non-coding ssRNAs, ~22 nucleotides in length, are produced by two RNase III proteins (Drosha/Dicer), regulate up to 60% of the protein-encoding genome.



53 miRNA differentially expressed in TB p'ts vs. HS. Only miR-21 showed an overlap in up / down regulation. Only 8 of these miRNA were identified in 2 or more studies: miR-20b, 21–5p, 22–3p, 26a, 29a-5p, 29c-3p, 378a-3p, 155. Tuberculosis 118 (2019) 101860



miRNAs either promotes or inhibits important pathways and cellular responses in macrophages, dendritic cells and CD4+ T cells against *M.tb*.



List of immune-suppressive miRNAs in TB

MirRNA	Mechanism of action	Final effect		
miR-21	Overexpression of IL-10 mRNA and Down-regulation of IL-12 mRNA	Suppression of immune- response against TB		
miR-29	Degradation of IFN-Y	Intercellular growth of tubercle bacillus within macrophages		
miR-99b	Decline expression of TNF- α	Suppression of immune- response against TB		
miR-125b	Blockade TNF-α mRNA	Suppression of immune- response against TB		
miR-27b	Suppression of NF-kB signaling pathway	Suppression of immune- response against TB and intercellular growth of MTB		
miR-1178	Attenuation of TLR-4 expression and inhibition of pro-inflammatory cytokines	Suppression of immune- response against TB		
Adv Biomed Res. 2019; 8: 3				

List of immune-effective miRNAs in TB

MirRNA	Mechanism of action	Final effect		
miR-155	Stability of TNF-α mRNA and activation of MAPKs signaling pathway	Efficient immune-response against MTB, provoke phagocytosis and elimination of MTB		
miR-424	Dysregulation of NFI-A78	Macrophage maturation and differentiation		
miR-223	Targeted Mef2c	Granulocytes production and stimulation pro-inflammatory response		
miR27a	Blocking IRAK4 signaling pathway	Increase of pro-inflammatory cytokines such as IL- β , IL-6 and IFN- γ		
miR-20b	targeting the NLRP3/caspase- 1/IL-1β pathways	Induce inflammation process		
miR-582-5p	Decline monocytes apoptosis via down-regulating FOXO1	Promotion of anti- tuberculosis immune response		

Adv Biomed Res. 2019; 8: 3

BCG-induced trained innate immunity through epigenetic mechanisms

How dose the BCG vaccine induce specific and non-specific immunity?

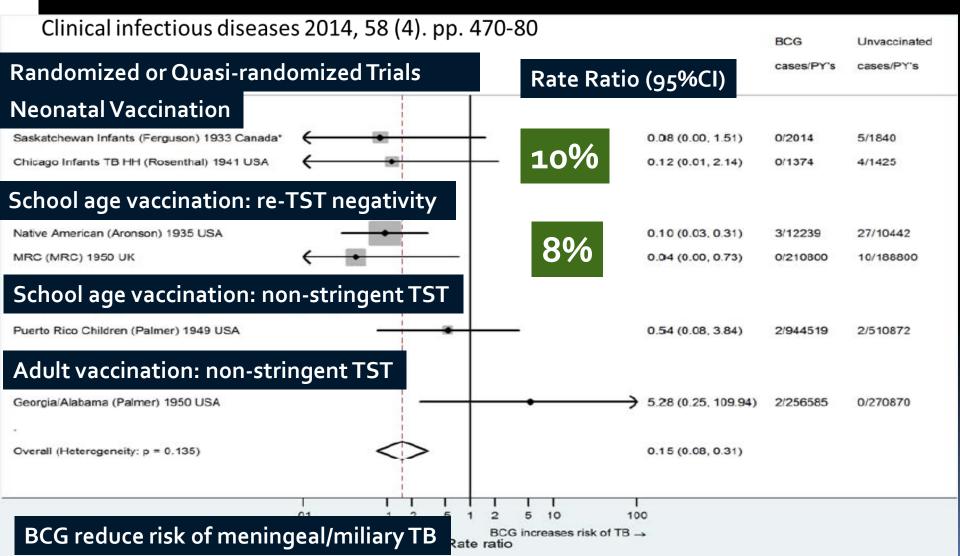
What factors influence the immune responses induced by BCG? Front Immunol. 2019 Jun 11;10:1317

- The BCG vaccine has been used since 1921 to prevent TB and is considered to be the world's most widely used vaccine.
- Specific effect of BCG : about 60% efficacy
 - good protection against disseminated and pulmonary TB disease in young children
 - variable efficacy against pulmonary TB in adults when given later
 - lasting for up to 15 years in the United Kingdom, 30–40 years in Norway, and even as long as 50–60 years in Alaska.
- Non-specific effects of BCG : about 25% efficacy
 - beneficial effects on all-cause mortality in infants (low birth weight) in West Africa.
 - There was no difference between outcomes in normal birth weight infants or premature infants in this European setting.

BCG Protection against active pulmonary TB appeared greatest in children stringently TB tested, (rate ratio [RR], 0.26; 95% confidence interval [CI], .18-.37), or infants (RR, 0.41; 95% CI, .29-.58). Clinical infectious diseases 2014, 58 (4). pp. 470-80

BCG Unvaccinated **Randomized or Quasi-randomized Trials** Rate Ratio (95%CI) cases/PY's cases/PY's **Neonatal Vaccination** Saskatchewan Infants (Ferguson) 1933 Canada 0.20 (0.06, 0.69) 3/2162 14/2000 Chicago Infants CCH (Rosenthal) 1937 USA 0.34 (0.19, 0.60) 16/15618 48/15822 Turtle and Recebud Infants (Aronson) 1938 USA 0.40 (0.13, 1.25) 4/968 11/1060 Chicago Infants TB HH (Rosenthal) 1941 USA 0.28 (0.08, 1.02) 3/1374 11/1424.5 0.62 (0.35, 1.07) 22/962.5 Bombay Infants (Mehta) 1976* India 27/716.25 41% Fixed effect Within stratum (Heterogeneity: p= 0.387) 0.40 (0.28, 0.56) Random effects Within stratum (2 = .0065) 0.41 (0.29, 0.58) School age vaccination: re-TST negativity Native American (Aronson) 1935 USA 0.26 (0.20, 0.33) 79/66716 267/58219 Ida B. Wells Housing Project (Rosenthal) 1942 US 0.15 (0.01, 2.87) 0/7770 3/8050 Georgia School (Shaw) 1947 USA 1.25 (0.28, 5.58) 4/29830 3/27966 MRC (MRC) 1950 UK 0.22 (0.16, 0.31) 40/210800 163/188800 Fixed effect Within stratum (Heterogeneity: p= 0.160) 0.25 (0.21, 0.31) Random effects Within stratum 42 = .048) 0.26 (0.18, 0.37) 26% 5 100 BCG reduce risk of pulmonary TB k of TB BCG increases risk of TB Rate ratio

BCG Protection against meningeal and miliary TB was also high in infants (RR, 0.1; 95% CI, .01-.77) and children stringently tuberculin tested (RR, 0.08; 95% CI, .03-.25).



Primary analysis with 14 studies (n=3855) estimated an overall risk ratio of 0.81 (95% confidence interval 0.71 to 0.92), indicating BCG 19% protective efficacy against latent TB infection among vaccinated children. BMJ. 2014;

	No	with TB		
Study (trawa score)	Vaccinated	Not vaccinated	Risk ratio (95% Cl)	Risk Ratio
Elispot				
Ewer 2003 (4)	131/467	16/6B		4 1.19 (0.76 to 1.87)
Soysal 2005 (7)	306/770	110/209		28 0.76 (0.65 to 0.88)
Eisenhut 2009 (5)	16/56	67/143		6 0.61 (0.39 to 0.96)
HIII 2011 (3)	102/330	106/313		17 0.91 (0.73 to 1.14)
M-H subtotal: Pr0.10, I ² =52	194		- - -	55 0.82 (0.73 to 0.93)
D+L subtotal			-	0.83 (0.68 to 1.02)
QuantiFERON	37/115	12/44		3 1.18 (0.68 to 2.05)
Domingues 2008 (4)	29/82	13/35		3 0.95 (0.57 to 1.60)
Bianchi 2009 (4)	1/6	4/12		
Adetita 2010 (4)	98/148	60/88		12 0.97 (0.81 to 1.17)
Eriksen 2010 (6)	7/53	26/73	T	3 0.37 (0.17 to 0.79)
Tsolia 2010 (3)	20/51	43/7B		5 0.71 (0.48 to 1.06)
Attet-Gomez 2011 (5)	36/116	25/50		6 0.62 (0.42 to 0.91)
Rutherford 2012 (3)	114/263	42/67		11 0.69 (0.55 to 0.87)
Neira-Munoz 2008 (4)	0/2	18/21	-	1 0.03 (0.00 to 194.41)
Okada 2008 (7)	30/173	3/22		1 1.27 (0.42 to 3.82)
M-H subtotal: Pr0.07, 12=43	1%		+	45 0.78 (0.69 to 0.89)
D+L subtotal				0.78 (0.64 to 0.06)
M-H overall: P+0.06, 1 ³ =409	5		+	100 0.81 (0. VE
0+Loverall				1 (0.
		BCO	protect TB infection	0.81 19%
			BCG non-prat	ective

Subgroup analysis of six studies (n=1745) that reported the number of people who progressed to active tuberculosis disease during screen.

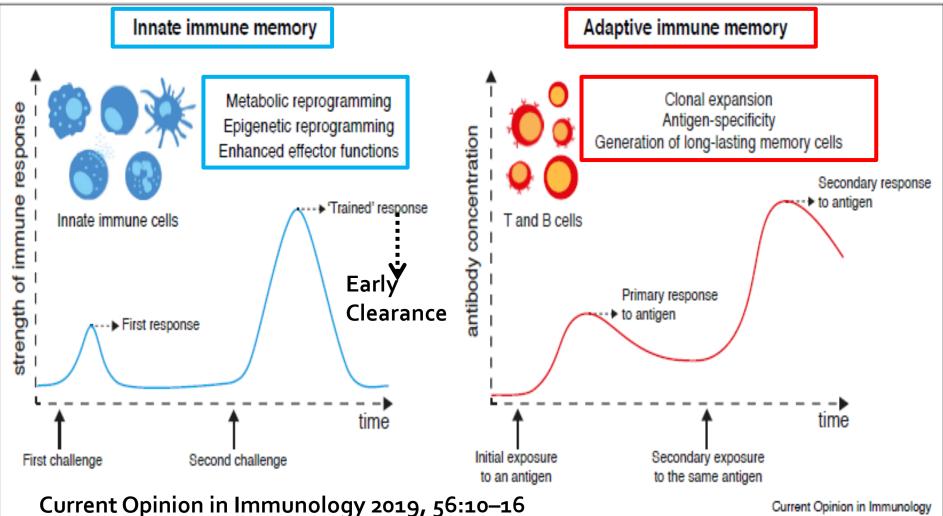
	No	with TB			
Latent TB infection	Vaccinated	Not vaccinated	Risk ratio (95% CI)	Risk Ratio	
Soysal 2005	306/770	110/209		60 0.76 (0.65 to 0.88)	
Domingues 2008	29/82	13/35		6 0.95 (0.57 to 1.60)	
Okada 2008	30/173	3/22		2 1.27 (0.42 to 3.82)	
Elsenhut 2009	16/56	67/143		13 0.61 (0.39 to 0.96)	
Eriksen 2010	7/53	26/73		8 0.37 (0.17 to 0.79)	
Tsolia 2010	20/51	43/78		0.71 (0.48 to 1.06)	
M-H subtotal: P=0.31, I ² =16%	408/1185	262/560		0.73 0.72 VE 27%	
0+L subtotal			•	0.73	
Active TO			and the second sec		
Active TB	3/770	10/209	B	25 0.08 (0.02 to 0.29)	
B	1/82	5/35		11 0.09 (0.01 to 0.70)	
Okada 2008	13/173	3/22		8 0.55 (0.17 to 1.78)	
Elsenhut 2009	3/56	15/143		13 0.51 (0.15 to 1.70)	
Eriksen 2010	1/53	7/73		9 0.20 (0.02 to 1.55)	
Tsolia 2010	B/S1	26/78		0.20 0.47 VE 7106	
M-H subtotal: P=0.12, 1 ² =42%	29/1185	66/560	-	0.29 U.av VE 71%	
0+L subtotal				0.29 (0.15 to 0.58)	
Latent Infectio	n to	10/110		28 0.11 (0.03 to 0.38)	
		5/13		13 0.09 (0.01 to 0.69)	
active TB disea	se	3/3		12 0.50 (0.29 to 0.86)	
		15/67		- 11 0.84 (0.28 to 2.55)	
Eriksen 2010	1/7	7/26		6 0.53 (0.08 to 3.63)	
Tsolla 2010	B/20	26/43		31 0.66 (
M-H subtotal: Px0.06, 1 ² =53% 0+L subtotal	29/408	66/262	-		
BMJ. 2014; 349: g46	43.	0.1	BCG protective	0.42 58%	

Non-Specific Effect: SIX RCTs showed that BCG reduced mortality from diseases other than TB by 25% (95% CI 6% to 41%).

 Table 2
 Controlled trials of the effect of BCG on mortality from causes other than tuberculosis among children in the USA and the UK (reported in papers published between 1948 and 1961)

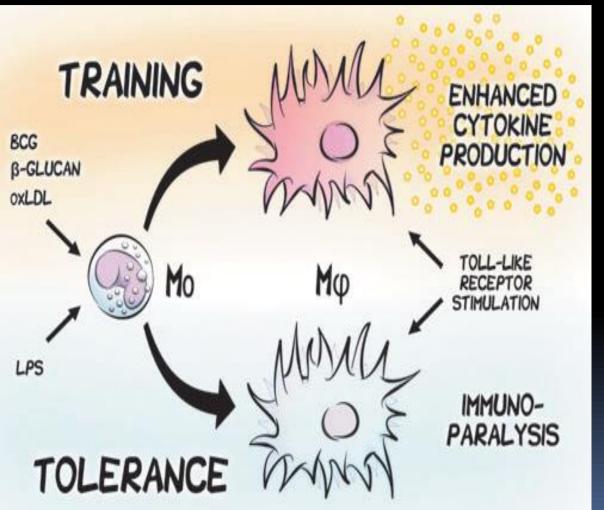
Study	Age followed	Allocation	BCG	No BCG	Reduction in mortality (95% CI)*
USA ⁴⁵	0–13 years	Alternate	N=231: 3/1261 (2.4‰)	N=220: 3/1320 (2.3‰)	-4% (-682% to 86%)†
USA ⁴⁶	0–15 years	Random	N=306: 49/2013.5 (24.3‰)	N=303: 51/1839.2 (27.7‰)	12% (–33% to 42%)†
USA ⁴⁷	0–20 years	Alternate	N=1551: 49/16406 (3.0‰)	N=1457: 56/15207 (3.7‰)	19% (-21% to 46%)
UK ⁴⁸	14–21 years	Odd/even	N=6700: 7/6700 (1.0‰)‡ §	N=6500: 10/6500 (1.5‰)‡	32% (–98% to 78%)†
USA ⁴⁹	0–16 years	Alternate	N=566: 14/566 (24.7‰)‡	N=528: 25/528 (47.3‰)‡	48% (-4% to 75%)
UK ⁴⁸	14–21 years	Odd/even	N=14100: 8/14100 (0.6‰)‡	N=16/13200 (1.2‰)‡	53% (–12% to 83%)†
Total					25% (6% to 41%)¶
Twins ³⁵	0–17 months	Twins	1/5 (89)=0.20 (0.02–1.68)**	DTP: 22/3 (164)=7.3 (2-38)**	p<0.001

Innate immune cells are able to undergo longterm adaptation and acquire enhanced capability to respond to certain stimuli, termed innate immune memory or trained immunity.



Current Opinion in Immunology

Trained immunity: Monocyte (Mo) memories of past encounters with microbial/nonmicrobial products can elicit vastly different responses to future exposures on differentiation to macrophages (Mu).



Trained immunity:

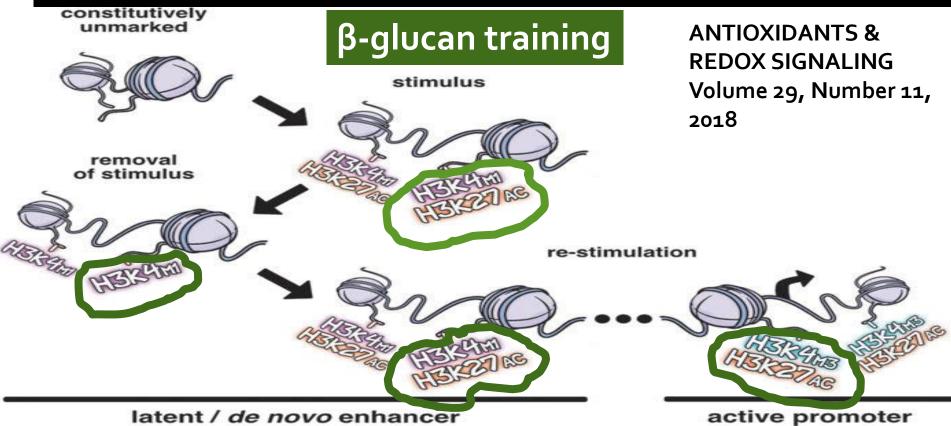
- induced by BCG, bglucan, or oxLDL,
- enhanced nonspecific response to subsequent infections
- enhancing the inflammatory and antimicrobial properties of innate immune cells.

Immune tolerance:

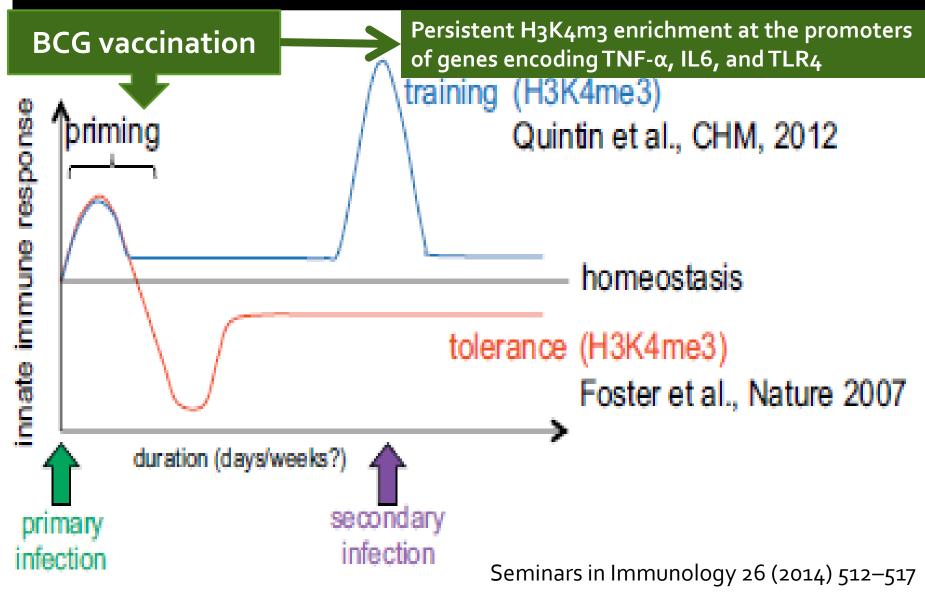
- primary stimulation with LPS induces a persistent refractory state,
- markedly reduced capacity to respond to re-stimulation.

ANTIOXIDANTS & REDOX SIGNALING Volume 29, Number 11, 2018

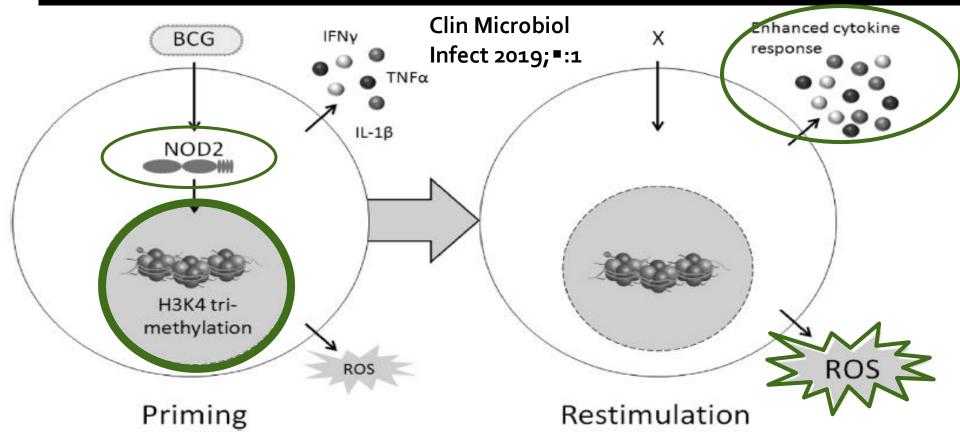
Latent enhancers prime a transcriptional memory in macrophages via Epigenetic change. Constitutively unmarked distal regulatory elements acquire epigenetic features of enhancers (open chromatin: H₃K₄m1 and H₃K_{27ac}) in response to specific stimuli.



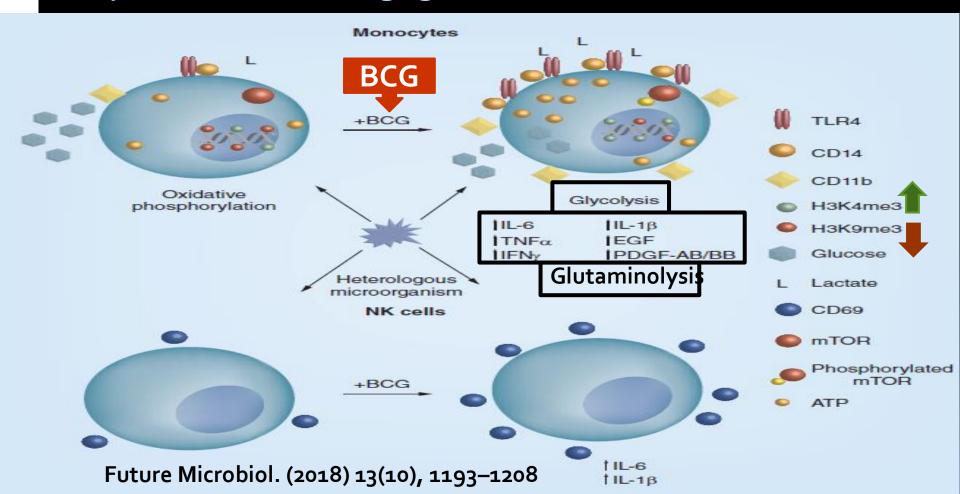
BCG vaccination induces trained immunity and this effect is beneficial both for protection against TB, as well as unrelated non-mycobacterial infections.



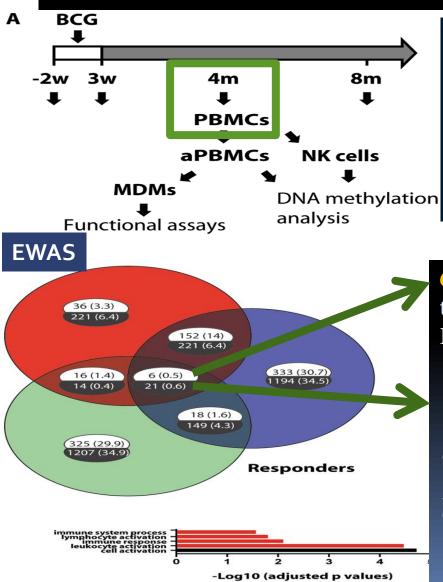
After BCG is taken up by the monocyte, it is recognized by the NOD2 receptor, which upon activation induces epigenetic and metabolic reprogramming of the cell (H3K4me3), leading to an enhanced, non-specific response to a subsequent infection through an increased production of cytokines and reactive oxygen species



BCG training-induced increased frequency of permissive H3K4me3 and reduced presence of inhibitory H3K9me3 at the promoters of cytokine, receptor and metabolic pathway component encoding genes.



Anti-mycobacterial activity correlates with altered DNA methylation pattern in immune cells from BCG-vaccinated subjects.



4 responders vs. 4 non-responders: enhanced anti-mycobacterial activity in Mtb-infected macrophages and increased IL-1β production

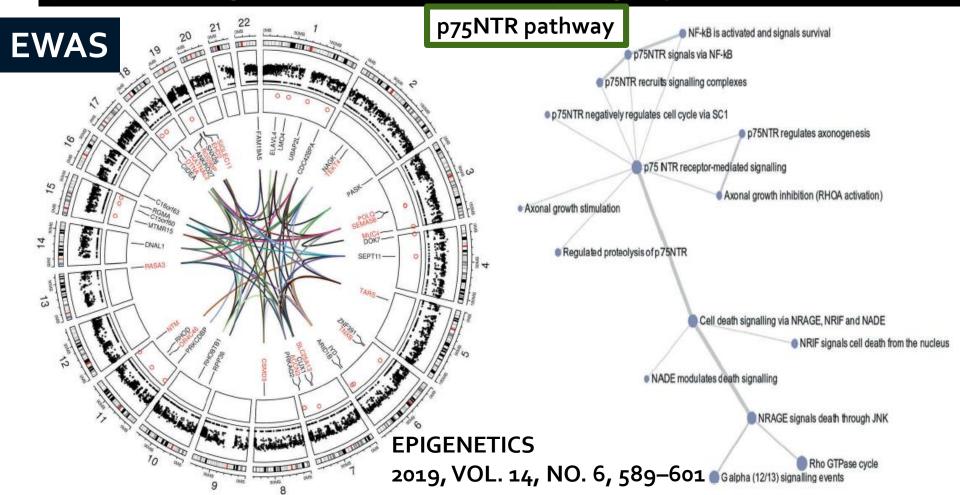
> ie 0.25 2 3w 4m 8m

6 Promoters displaying persistent hypo-Me at all time points (3 weeks, 4 months, 8 months): IFN-γ, RASAL1, GIMAP7, ADCY3, ATXN1, DIABLO

21 Promoters with persistent hyper-Me: TLR6, SRD5A2, SOX5, GNG7, SBNO2, SULT1C, NFKBIE, TRIM2, GPR84, SPATS2, CD59, ATXN1, NCOR2, ADARB1, LOC404266, SRGAP3, PIWIL2, SPG20, TSPAN4, CSGALNACT1

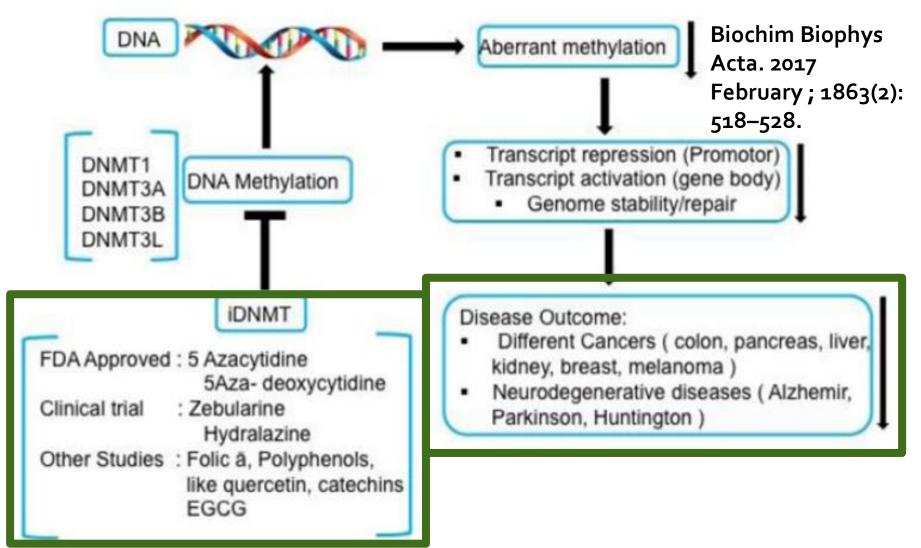
ScleNtlflc REPOrtS 2017 | 7: 12305

43 differentially methylated genes in PBMCs isolated from responders vs. non-responders at the time point before BCG vaccination-- enriched for actin-modulating pathways predicting differences in phagocytosis.

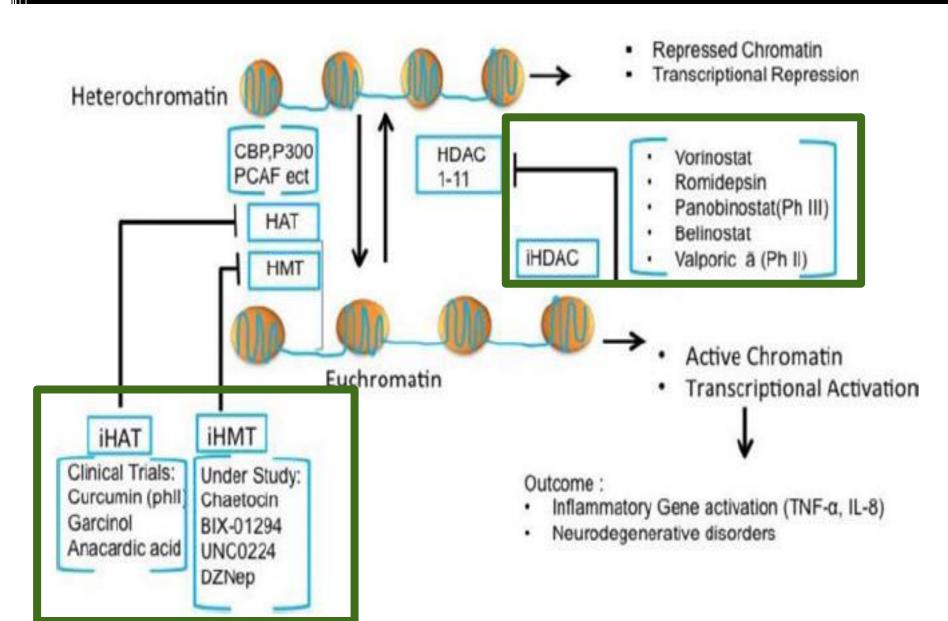


Future perspectives: Epigenetic targets for Host-Directed immunotherapy?

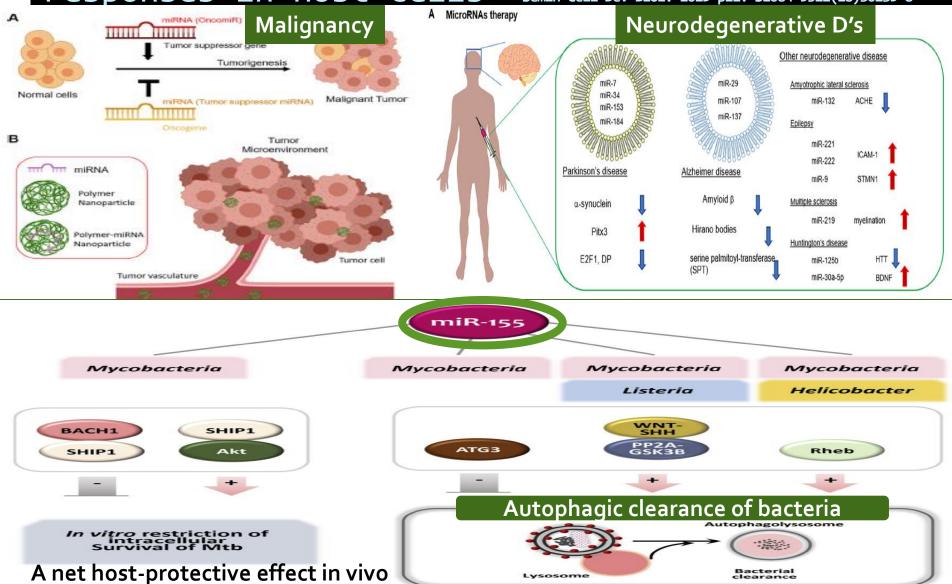
Several molecules have been screened for altering the DNA methylation status associated with the disease outcome and are currently in different phases of clinical trial.



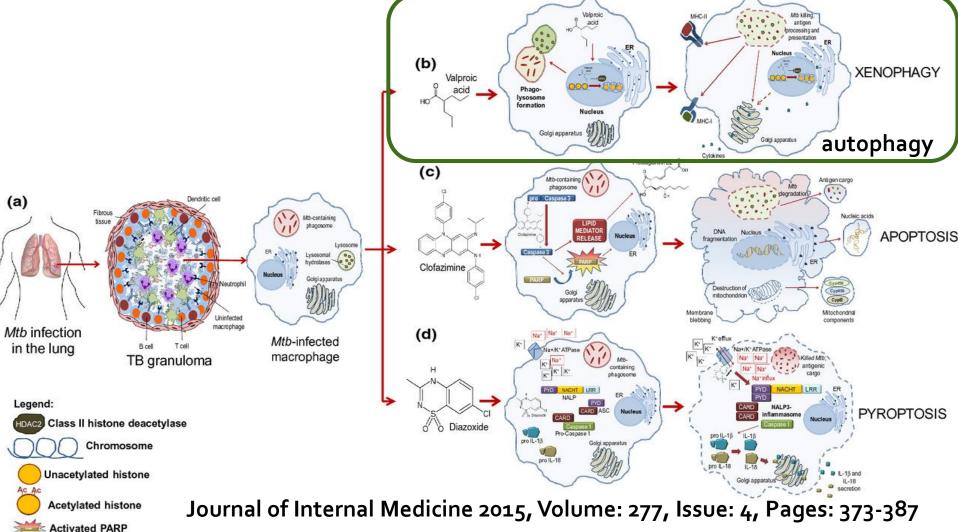
Clinical trials in aberrant histone modifications associated with disease outcome



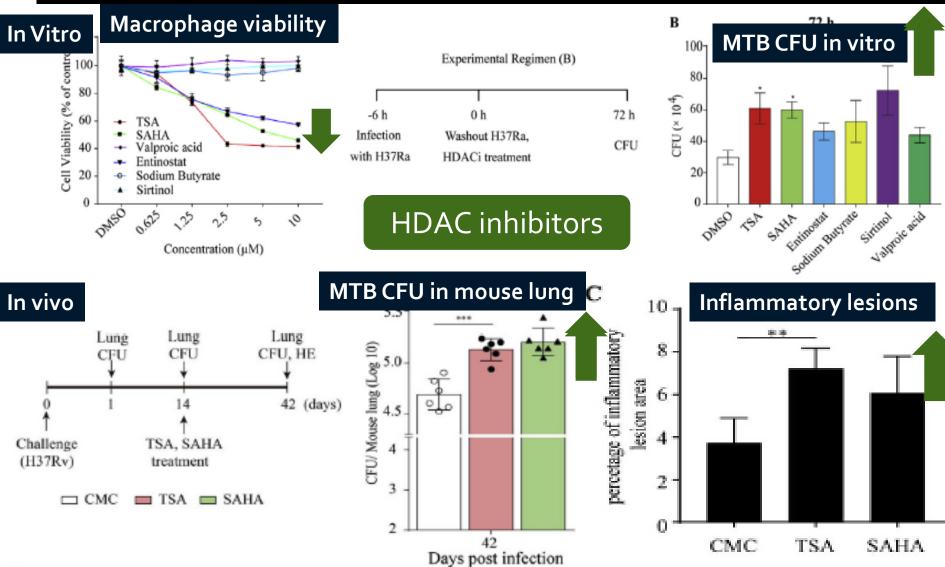
miRNAs may influence the outcome of bacterial infection by regulating autophagy/xenophagy responses in host cells Semin Cell Dev Biol. 2019 pii: 51084-9521(18)30239-8



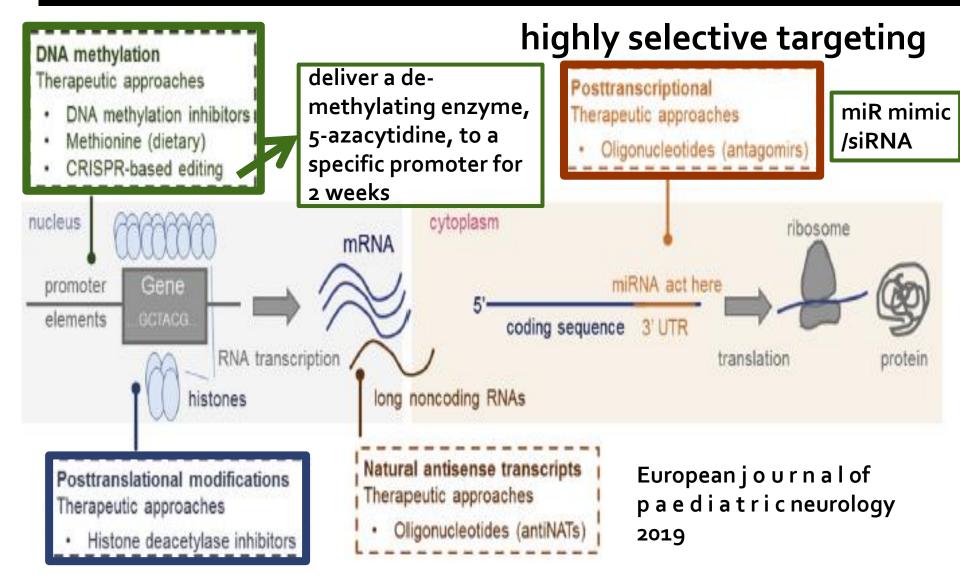
Host-directed therapies: Xenophagy may be initiated by valproic acid to allow histone acetylation followed by chromosome unwinding and subsequently enhanced gene expression.



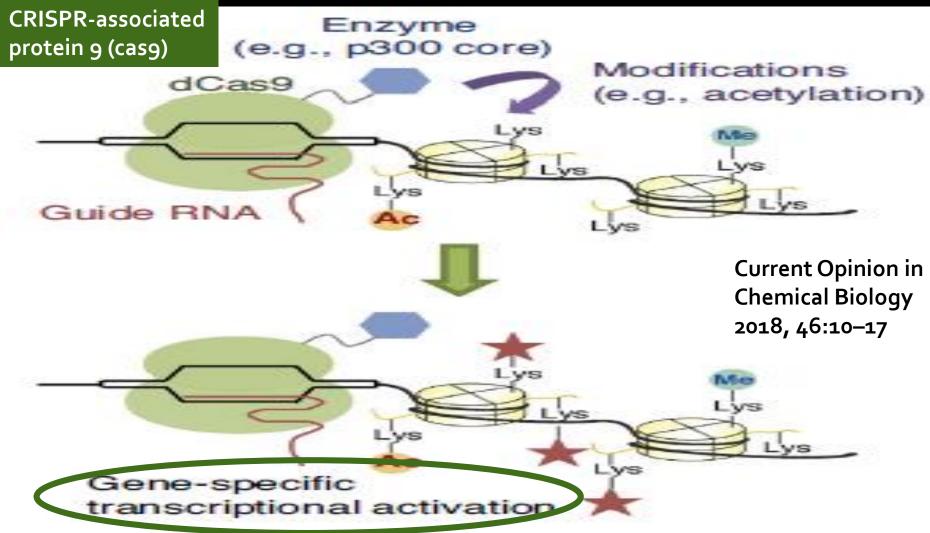
Non-specific Histone de-acetylase inhibitors impair the host immune response (autophagy & ROS) against MTB. Tuberculosis 118 (2019) 101861



Gene-Editing: clustered regularly interspaced short palindromic repeat (CRISPR) Sequence-specific RNAi: microRNA mimic/siRNA



The dCas9-p300 HAT core fusion protein was expressed in cells, which promoted lysine acetylation of histones at a specific genomic locus targeted by the guide RNA sequence.



Take Home Message (1)

- *M. TB* uses virulent factors Rv1988/Rv2966c/Rv3423.1/ RV1346c to hijack the host transcriptional machinery through epigenetic regulations.
- Association of active TB disease with
 - Aberrant DNA Met over VDR, TLR2, IL18R1, CYP27A1 genes
 - H3K14 hypo-Ac (TNF-α/IL12-β), HDAC1, KDM6B
 - At least 53 differentially expressed miRNAs
- M.TB infection in vitro leads to
 - de-Met of the NLRB3/CD82 gene promoters/1700 CpGs over distal enhancers
 - H3/H4 hyper-Ac, H3K9 hyper-Me, H3R42 hyper-Me, H3K4 hypo-Me/Ac, HDAC1/6, KDM6B, SET8
 - Immune-surppressive miRs: 21, 29, 99b, 125b, 27b, 1178
 - Immune-effective miRs: 155, 424, 223, 27a, 20b, 582

Take home message (2)

- Specific and Non-specific effects of BCG vaccine via trained innate immunity (innate immune memory)
 - H3K4 hyper-Me and H3K9 hypo-Me at the promoters of cytokine, receptor and metabolic pathway component encoding genes
 - BCG Responder: IFN-γ promoter DNA hypo-Me, TLR6 promoter DNA hyper-Me

Future perspectives for research:

- Epigenetic marks for Early clearance/Progression from LTBI to active TB D's; single cell methylome
- CRISPR-based gene editing to deliver de-methylation agent/histone modifying enzymes to specific genomic locus
- Highly selective antagomiR/miRNA mimic to enhance or surppress specific sequence of target genes

Acknowledgement

- Professor Meng-Chih Lin, Assistant Administrator of KCGMH, Taiwan
 - Advisor
- Division of Pulmonary & Critical Care Medicine, Kaohsiung CGMH, Taiwan
 - Dr. Tung-Ying Chao
 - Dr. Sum-Yee Leung
 - Dr. Chao-Chien Wu
 - Dr. Wen-Feng Fang
 - Dr. Yi-Hsi Wang
 - Dr. Huang-Chih Chang

- Graduate Institute of Clinical Medical Sciences, Chang Gung University College of Medicine, Taiwan
 - Professor Chang-Chun Hsiao
- Internal Medicine Core Facility, KCGMH, Taiwan
 - Ting-Ya Wang, Master
 - Yi-Xin Zheng
 - Yong-Yong Lin, Master
- Rheumatology, Kaohsiung Medical University Hospital,
 - Professor Chung-Jen Chen

Thank You for Your Attention

