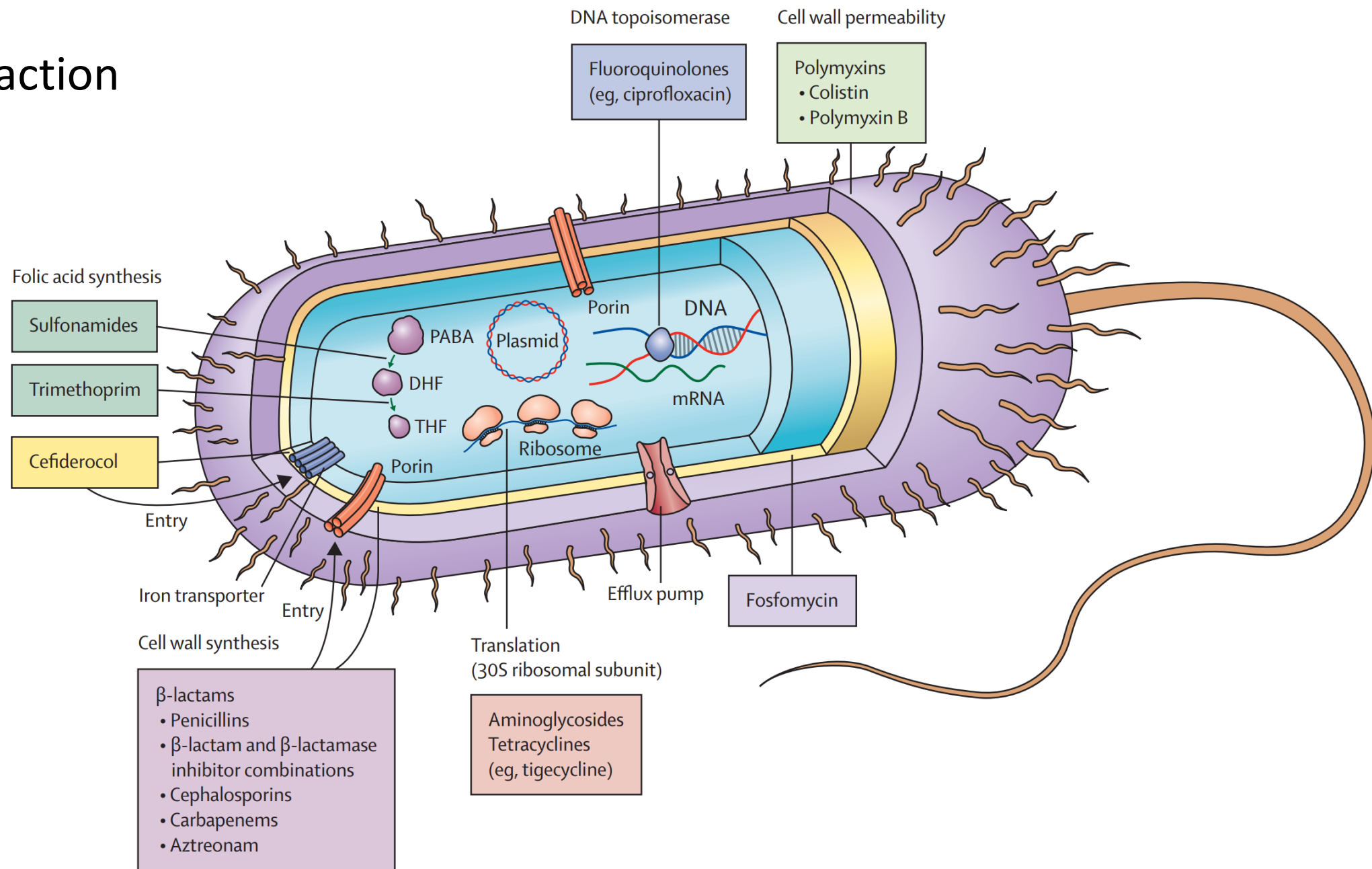


The Smart Choice of Novel Antibiotics in the Era of CRE in ICU

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陽明交通大學急重症醫學研究所

Mechanism of action for antibiotics



Lancet. 2025 Jan 18;
405(10474):257-272.

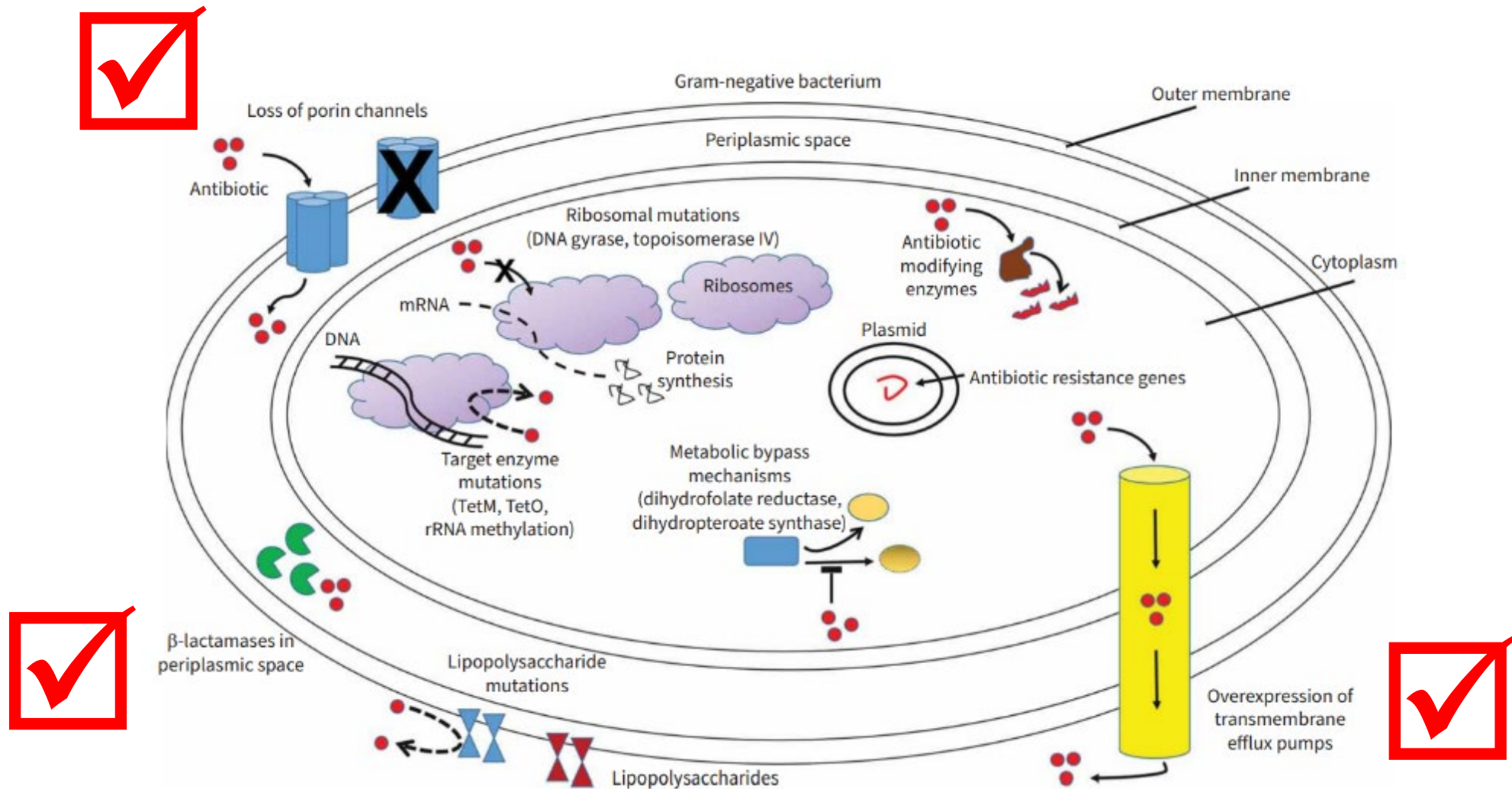
WHO priority pathogens

Priority	Pathogen	Resistance
Critical	<i>Acinetobacter baumannii</i>	Carbapenem
	<i>Pseudomonas aeruginosa</i>	Carbapenem
	<i>Enterobacteriaceae</i>	Carbapenem Third generation cephalosporin
High	<i>Enterococcus faecium</i>	Vancomycin
	<i>Staphylococcus aureus</i>	Methicillin Vancomycin
	<i>Helicobacter pylori</i>	Clarithromycin
	<i>Campylobacter</i>	Fluoroquinolone
	<i>Salmonella</i> spp.	Fluoroquinolone
	<i>Neisseria gonorrhoeae</i>	Fluoroquinolone Third generation cephalosporin
	<i>Streptococcus pneumoniae</i>	Penicillin
Medium	<i>Haemophilus influenzae</i>	Ampicillin
	<i>Shigella</i> spp.	Fluoroquinolone

Carbapenem resistance rates by region

	Asia-Pacific	Europe	North America
<i>Acinetobacter baumannii</i>	25-90.5	2.5-100	11.4-50.4
<i>Pseudomonas aeruginosa</i>	10.3-50	0-56.4	10.3-58.5
<i>Klebsiella pneumoniae</i>	1.6-25	0.2-58.6	3.1-12.9
<i>Escherichia coli</i>	0-7.1	0-7	0.2-4.3

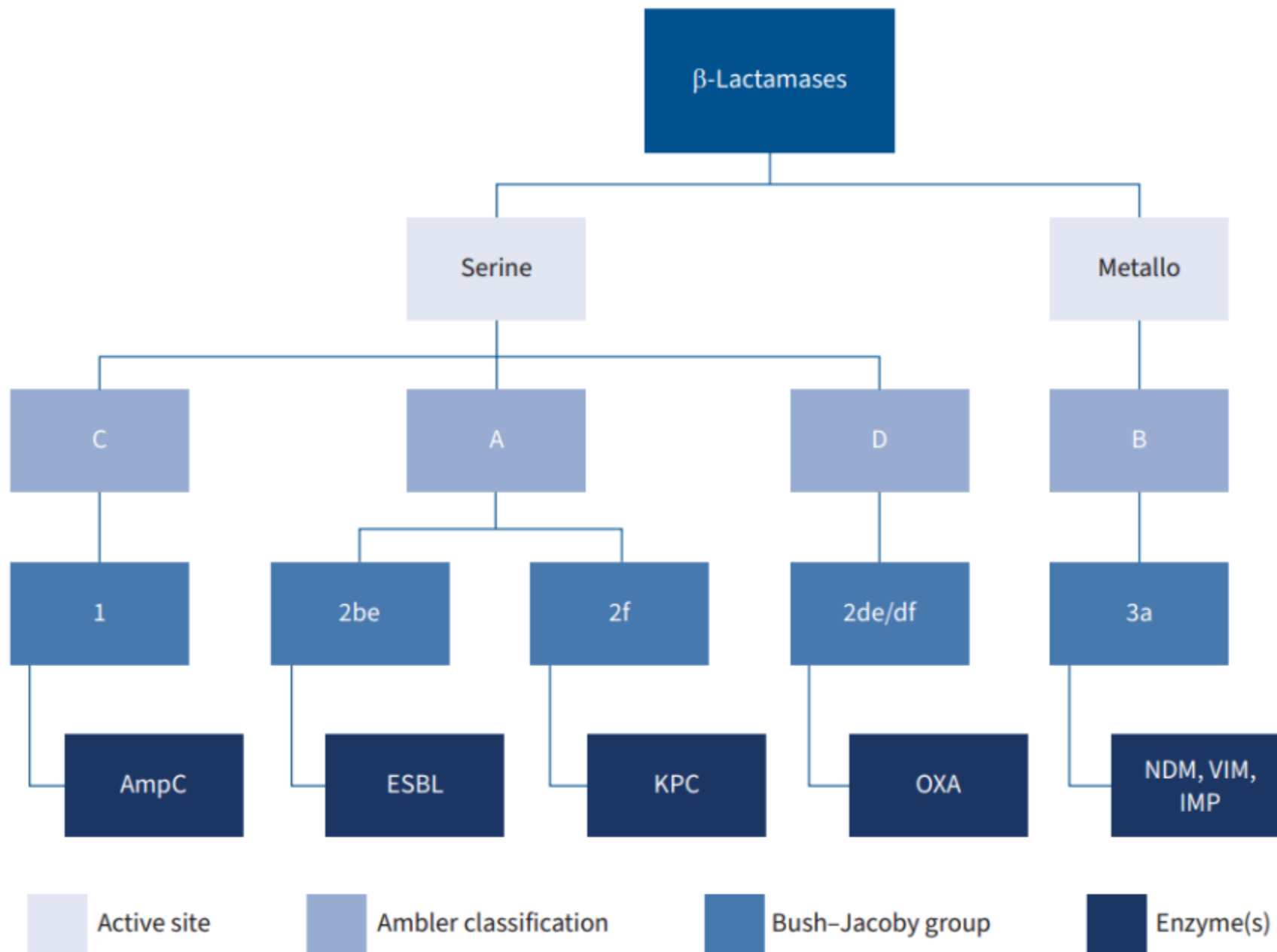
Mechanisms of antibiotics resistance in GNB



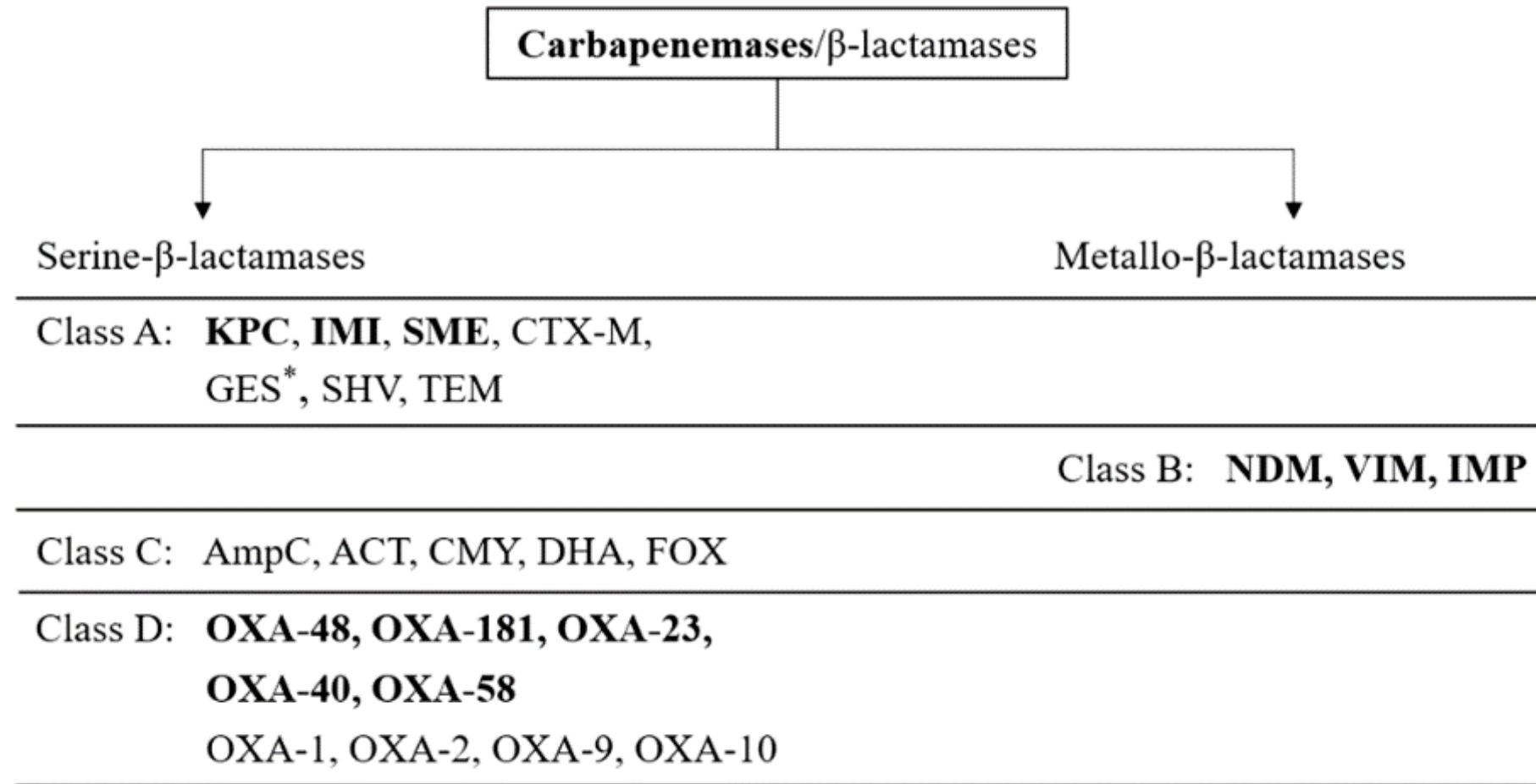
Mechanisms of GNB resistance

Categories		Examples
β-lactams^{19,20} (eg, piperacillin-tazobactam, cefepime, meropenem, and ceftazidime-avibactam)		
Antibiotic modification	β-lactamase enzymes	Extended-spectrum β-lactamases (SHV, TEM, and CTX-M); carbapenemases (KPC, OXA-48, NDM, IMP, and VIM)
Decreased entry	Porin mutations	<i>Escherichia coli</i> (OmpC and OmpF); <i>Klebsiella pneumoniae</i> (OmpK35 and OmpK36); <i>Pseudomonas aeruginosa</i> (OprD); <i>Acinetobacter baumannii</i> (CarO)
Increased efflux	Efflux pumps	<i>E coli</i> (AcrAB-TolC); <i>P aeruginosa</i> (MexAB-OprM); <i>A baumannii</i> (AdeABC and AdeIJK)
Aminoglycosides²¹⁻²⁴ (eg, gentamicin, tobramycin, and amikacin)		
Antibiotic modification	Aminoglycoside-modifying enzymes (AMEs)	N-acetyltransferases (AAC); O-adenyltransferases (ANT); O-phosphotransferases (APH)
Decreased entry	Alterations in bacterial cell outer membrane	PhoPQ; PmrAB; ParRS; CprRS
Target alteration	16S-ribosomal RNA methyltransferases (RMTs)	ArmA; RmtA; RmtB; RmtC; RmtD; RmtE; RmtF; RmtG; NpmA; NpmB; NpmC
Target alteration	30S ribosome subunit binding site mutations	<i>rrs</i> and <i>rpsL</i> gene mutations
Tetracycline derivatives²⁵⁻²⁸ (eg, minocycline, tigecycline, and eravacycline)		
Antibiotic modification	Increased breakdown	Tet(X); Tet(X2); Tet(X3); Tet(X4)
Increased efflux	Major facilitator superfamily (MFS) of transporters	Tet(A); Tet(B); Tet(C); Tet(D)
Increased efflux	Resistance nodulation division (RND)-type multidrug efflux pumps	AdeABC; TmexCD1-ToprJ1
Target alteration	Ribosomal protein alteration	RpsJ

Fluoroquinolones^{29,30} (eg, ciprofloxacin)		
Antibiotic modification	Increased breakdown	AME: AAC(6′)-Ib-cr
Increased efflux	Efflux pumps	QepA; OqxAB
Target alteration	Topoisomerase substitution	GyrA; GyrB; ParC; ParE
Target alteration	DNA gyrase protection	QnrA
Polymyxins³¹ (eg, colistin and polymyxin B)		
Target alteration	Two component systems	<i>K pneumoniae</i> (PhoPQ, PmrAB, MgrB, and CrrAB); <i>P aeruginosa</i> (PhoPQ, PmrAB, ParRS, ColRS, and CprRS); <i>A baumannii</i> (PmrAB)
Target alteration	Mobile (plasmid-mediated) colistin resistance enzymes	Encoded by <i>mcr</i> genes
Fosfomycin³²		
Antibiotic modification	Fosfomycin modifying enzymes	Encoded by <i>fos</i> genes
Decreased entry	Alterations in fosfomycin transporters	GlpT; UhpT; CyaA; PtsI

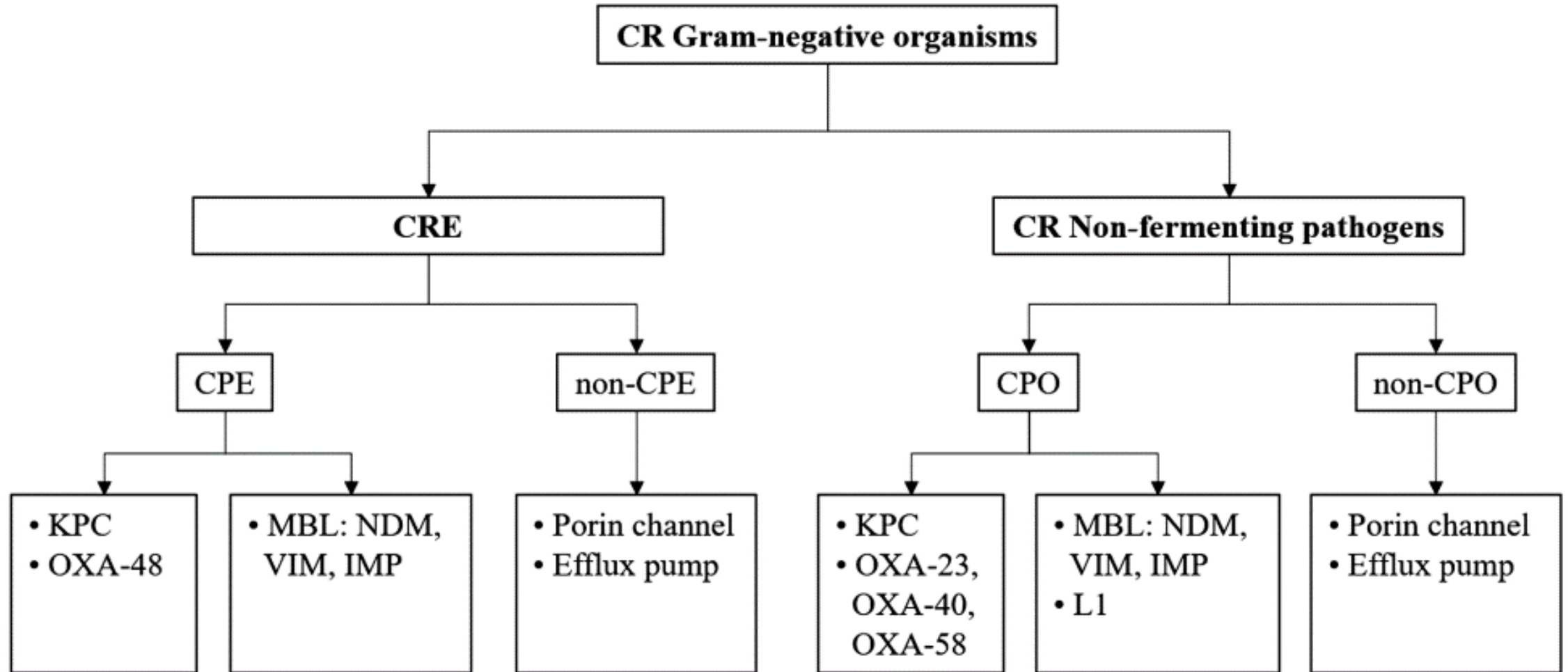


Classification of carbapenemases/ β -lactamases



* Some variants may possess carbapenemase activity

Mechanism of carbapenem resistance



台灣醫院感染管制與抗藥性監測管理系統(THAS系統)

2016~2025Q1 加護病房常見之醫療照護相關感染菌種

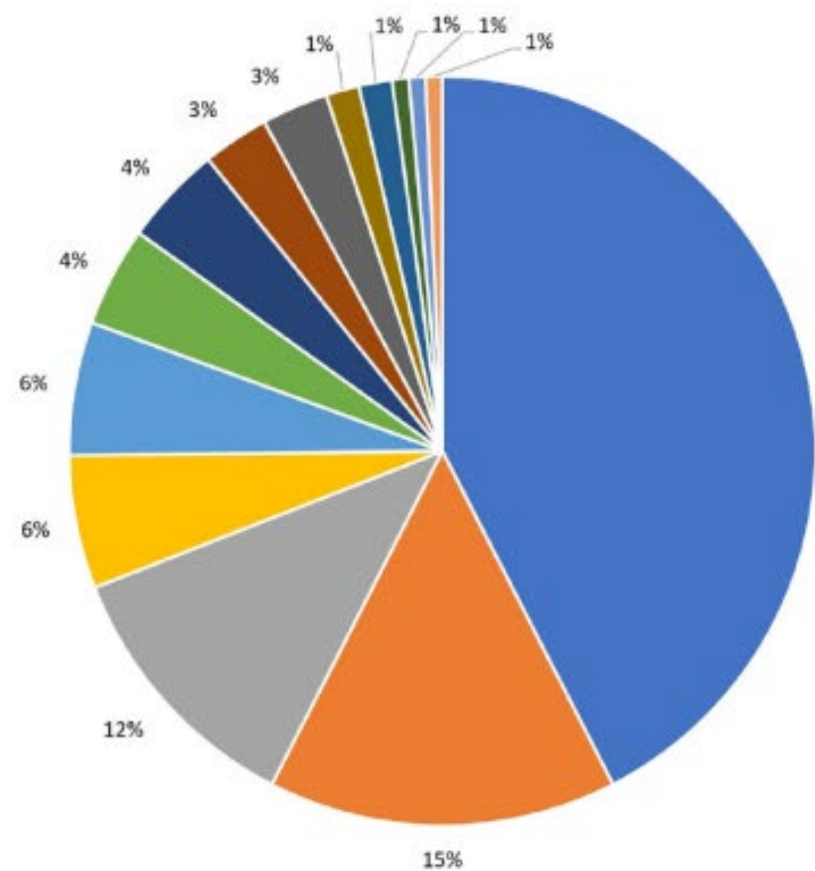
部位別: PNEUMONIA



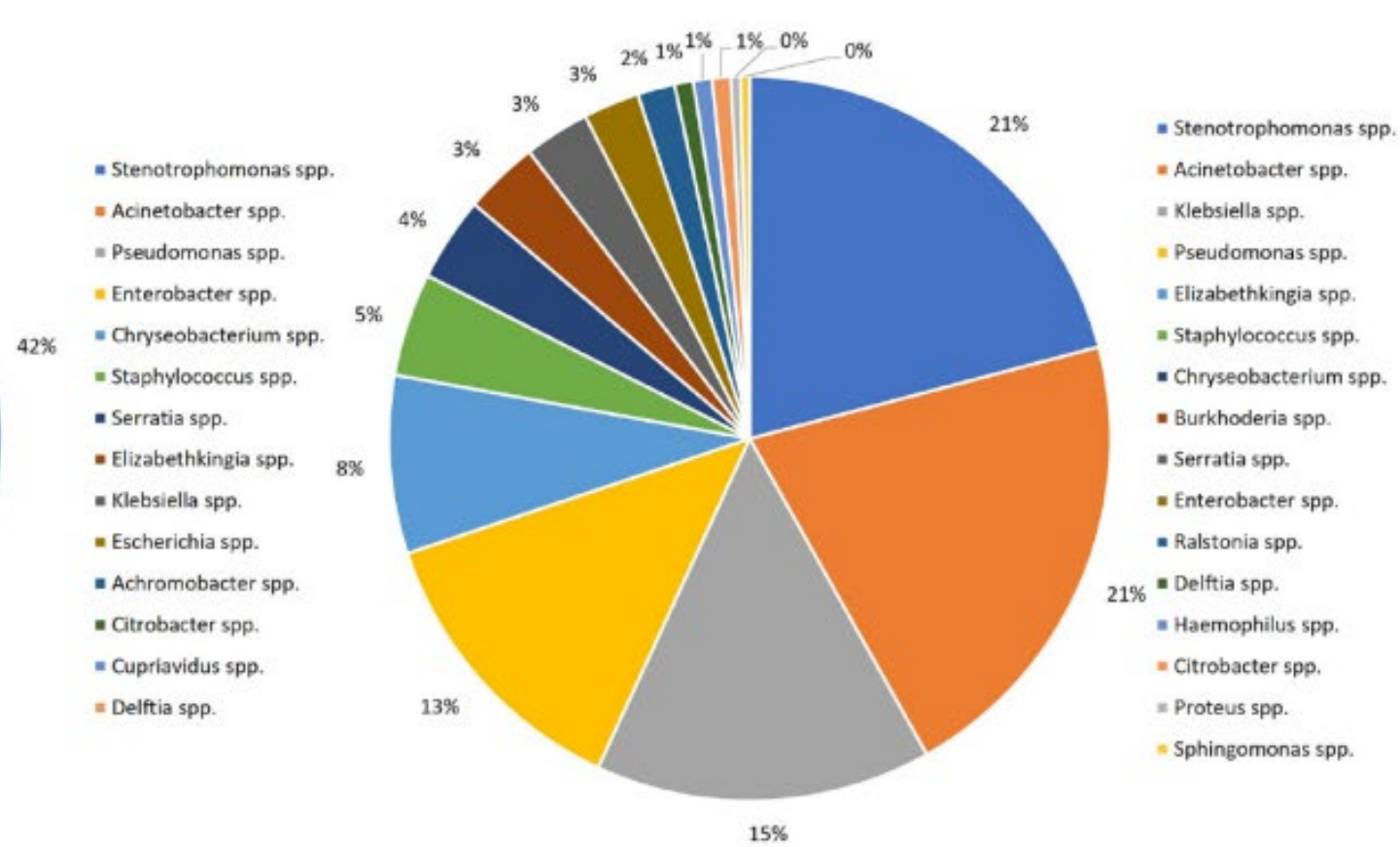
衛生福利部疾病管制署
TAIWAN CDC

醫學中心										
PNEU	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025Q1
菌種	排名	排名	排名	排名	排名	排名	排名	排名	排名	排名
<i>Pseudomonas aeruginosa</i>	1	1	1	1	1	1	2	2	1	1
<i>Klebsiella pneumoniae</i>	2	2	2	2	2	2	1	1	2	2
<i>Acinetobacter baumannii</i>	3	3	3	4	4	4	4	4	3	3
<i>Stenotrophomonas maltophilia</i>	5	5	4	3	3	3	3	3	4	3
<i>Staphylococcus aureus</i>	4	6	5	5	5	5	5	6	5	5
<i>Enterobacter species</i>	6	4	6	6	8	7	8	8	6	8
<i>Enterobacter cloacae complex</i>	-	PA > KP > AB				10	10	10	7	10
<i>Escherichia coli</i>	8					6	11	5	8	7
<i>Serratia marcescens</i>	7	8	8	11	9	10	6	10	8	6
<i>Chryseobacterium indologenes</i>	10	10	11	12	9	12	13	7	10	8
區域醫院										
PNEU	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025Q1
菌種	排名	排名	排名	排名	排名	排名	排名	排名	排名	排名
<i>Klebsiella pneumoniae</i>	3	3	3	3	3	1	1	2	1	1
<i>Pseudomonas aeruginosa</i>	1	2	2	1	2	2	2	1	2	2
<i>Acinetobacter baumannii</i>	2	1	1	2	1	3	3	3	3	3
<i>Stenotrophomonas maltophilia</i>	5	5	4	5	4	5	4	4	4	4
<i>Escherichia coli</i>	8	7	6	7	7	7	7	7	5	8
<i>Staphylococcus aureus</i>	4					4	5	5	6	5
<i>Enterobacter species</i>	6	KP > PA > AB				6	6	6	7	6
<i>Serratia marcescens</i>	9	9	9	9	10	9	10	9	8	6
<i>Candida albicans</i>	7	8	8	8	8	8	8	8	9	13 ¹⁰
<i>Acinetobacter nosocomialis</i>	-	-	22	22	-	22	16	17	10	13

COVID-19 MV with nosocomial pneumonia



Alpha variant with secondary pneumonia



Omicron variant with secondary pneumonia

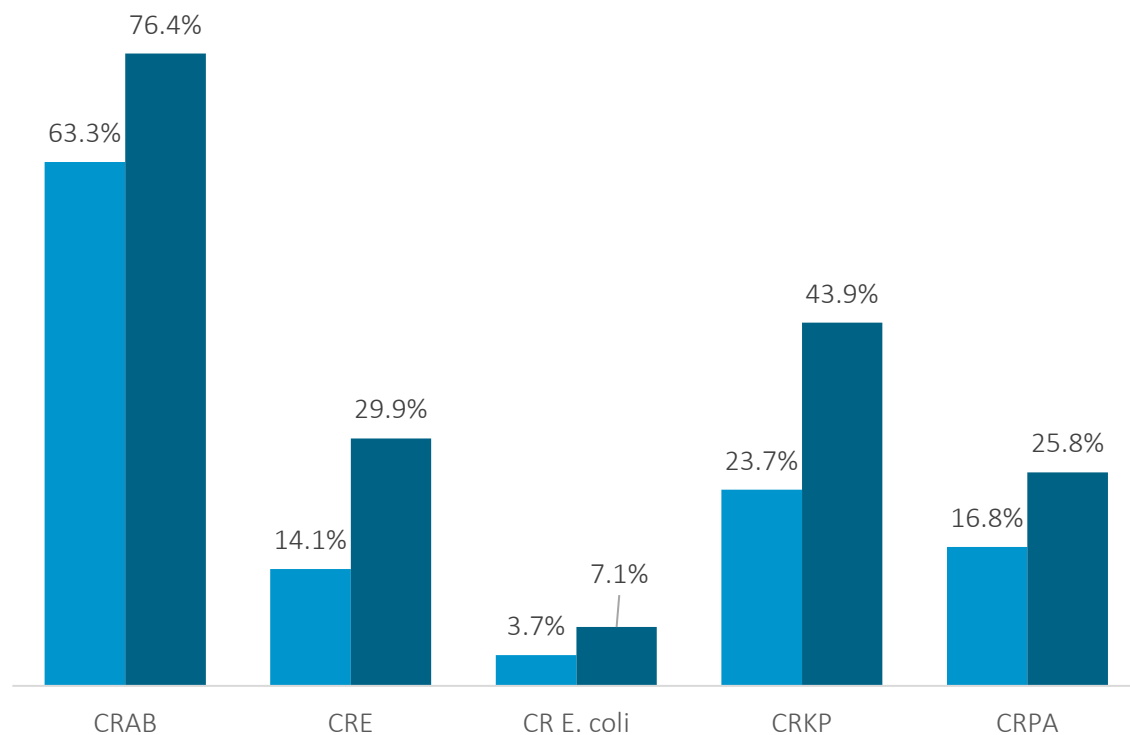
Carbapenem resistant GNB 比例居高不下

■ 2016 ■ 2025Q1

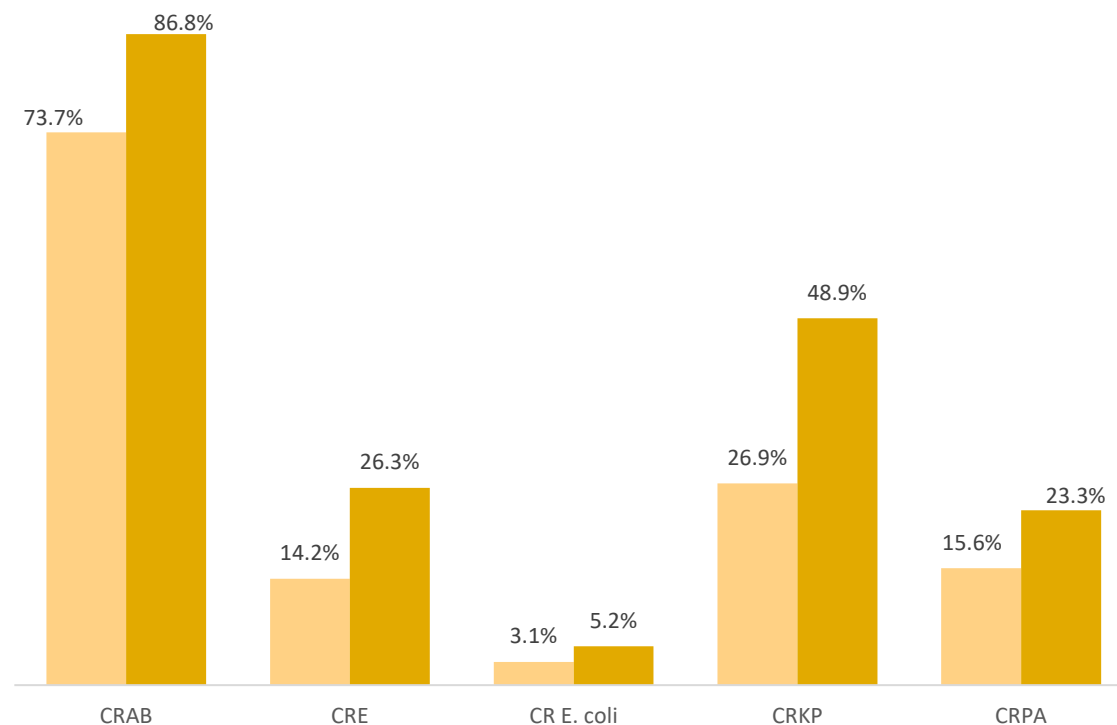
■ 2016 ■ 2025Q1



衛生福利部疾病管制署
TAIWAN CDC

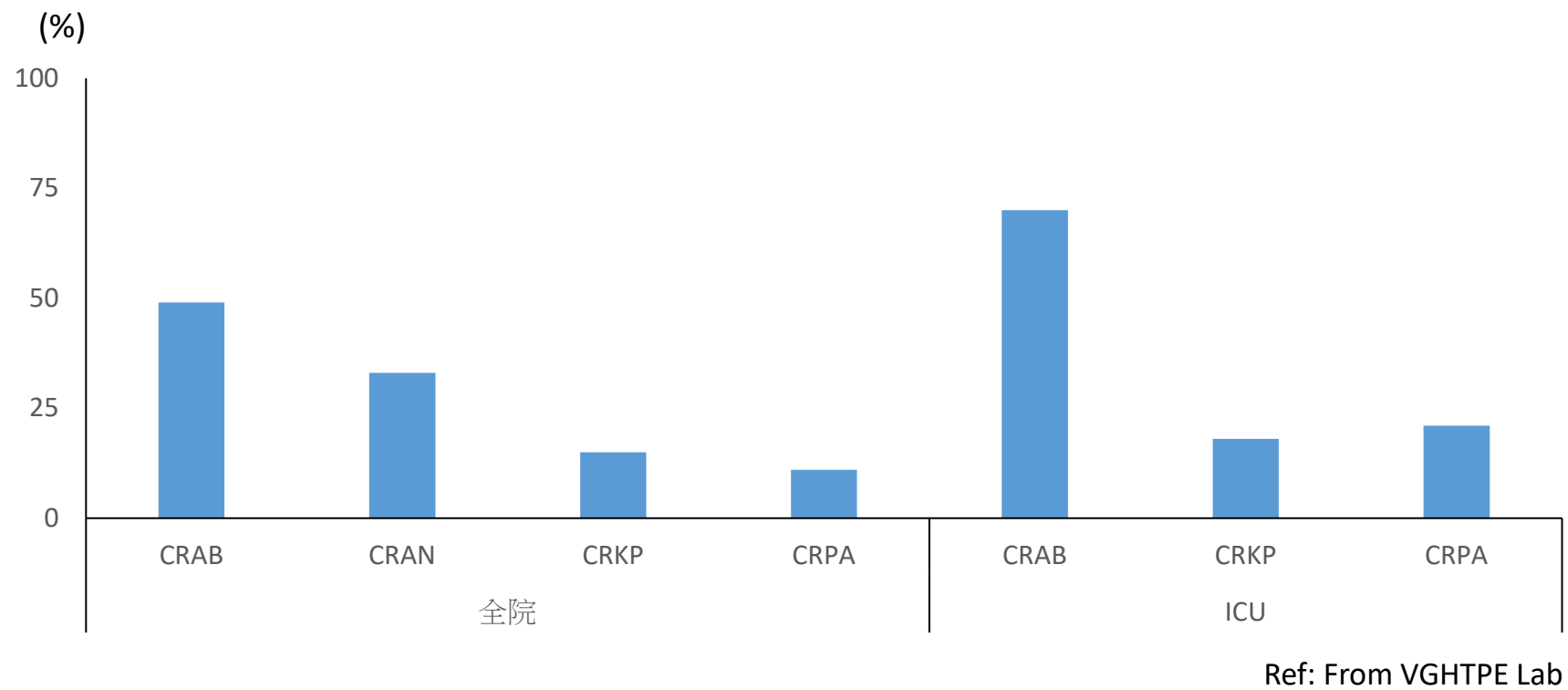


醫學中心



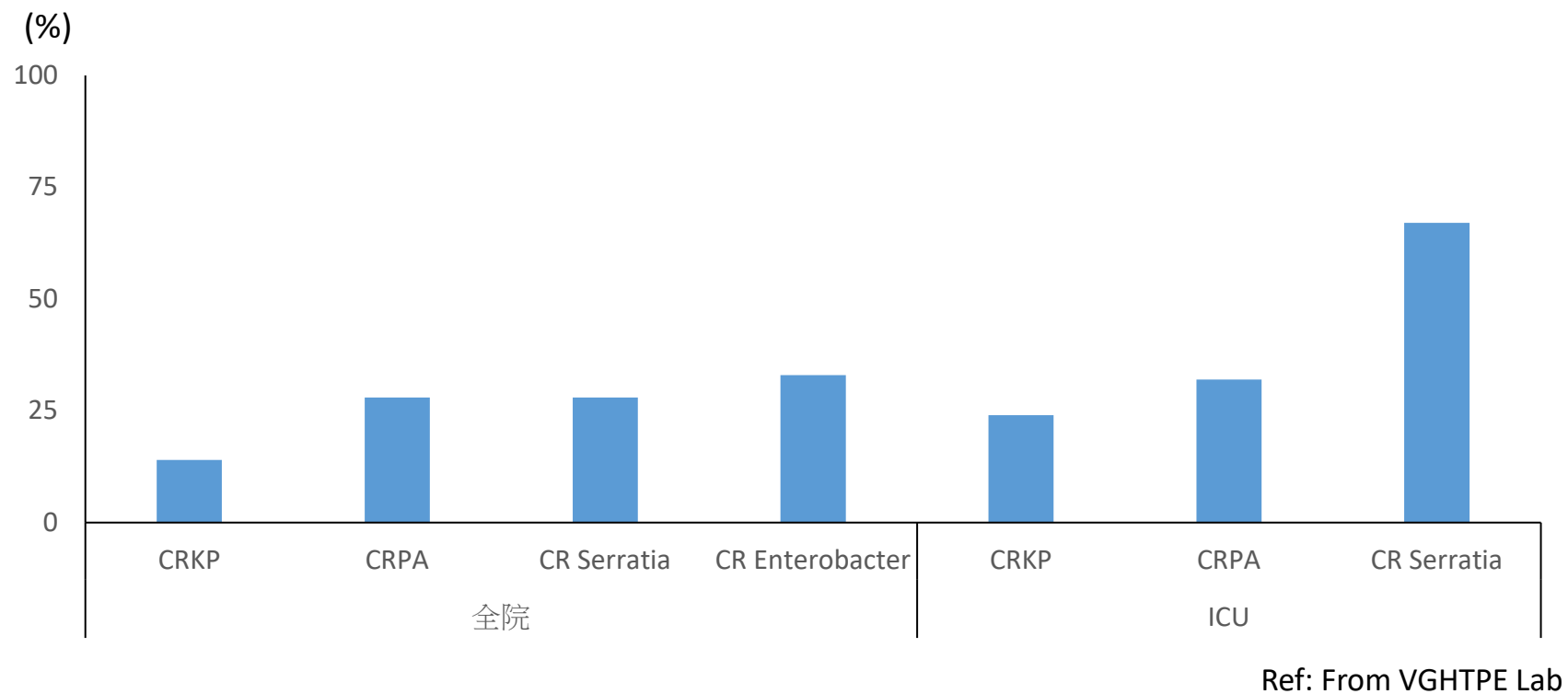
區域醫院

CRGNB in ordinary culture VGHTPE 2025/7-2025/12

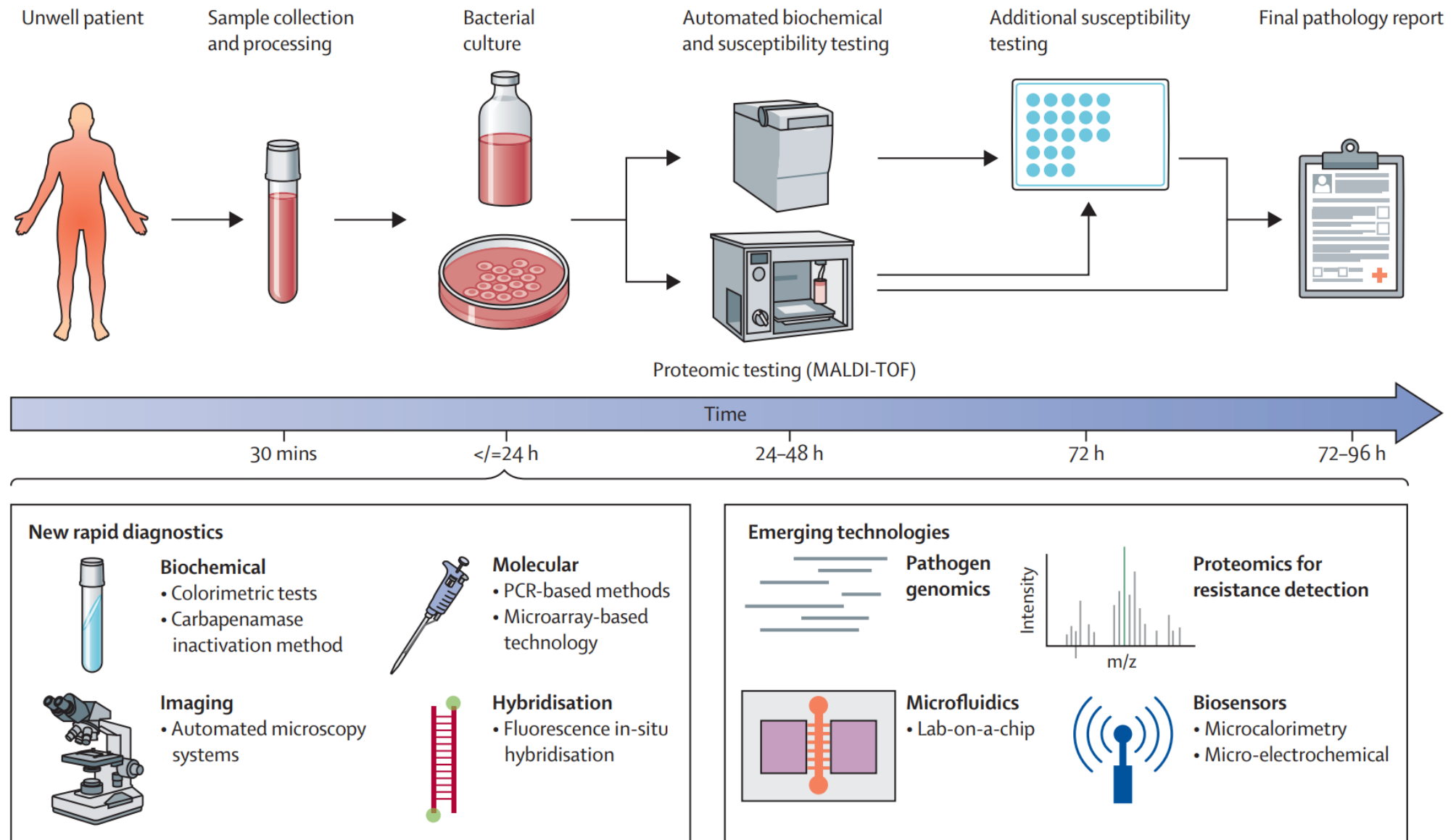


CRGNB in blood culture

VGHTPE 2025/7-2025/12



Diagnostic testing In MDR infection



Lancet. 2025 Jan 18;
405(10474):257-272.

The BioFire FilmArray Pneumonia Panel

BACTERIA:

(Semi-Quantitative)

- *Acinetobacter calcoaceticus-baumannii* complex
- *Enterobacter cloacae* complex
- *Escherichia coli*
- *Haemophilus influenzae*
- *Klebsiella aerogenes*
- *Klebsiella oxytoca*
- *Klebsiella pneumoniae* group
- *Moraxella catarrhalis*
- *Proteus* spp.
- *Pseudomonas aeruginosa*
- *Serratia marcescens*
- *Staphylococcus aureus*
- *Streptococcus agalactiae*
- *Streptococcus pneumoniae*
- *Streptococcus pyogenes*

ATYPICAL BACTERIA:

(Qualitative)

- *Chlamydia pneumoniae*
- *Legionella pneumophila*
- *Mycoplasma pneumoniae*

VIRUSES:

- Adenovirus
- Coronavirus
- Human metapneumovirus
- Human rhinovirus/enterovirus
- Influenza A virus
- Influenza B virus
- Parainfluenza virus
- Respiratory syncytial virus

ANTIMICROBIAL RESISTANCE GENES:

Carbapenemases:

- IMP
- KPC
- NDM
- OXA-48-like
- VIM

ESBL:

- CTX-M

Methicillin resistance:

- *mecA/C* and MREJ (MRSA)



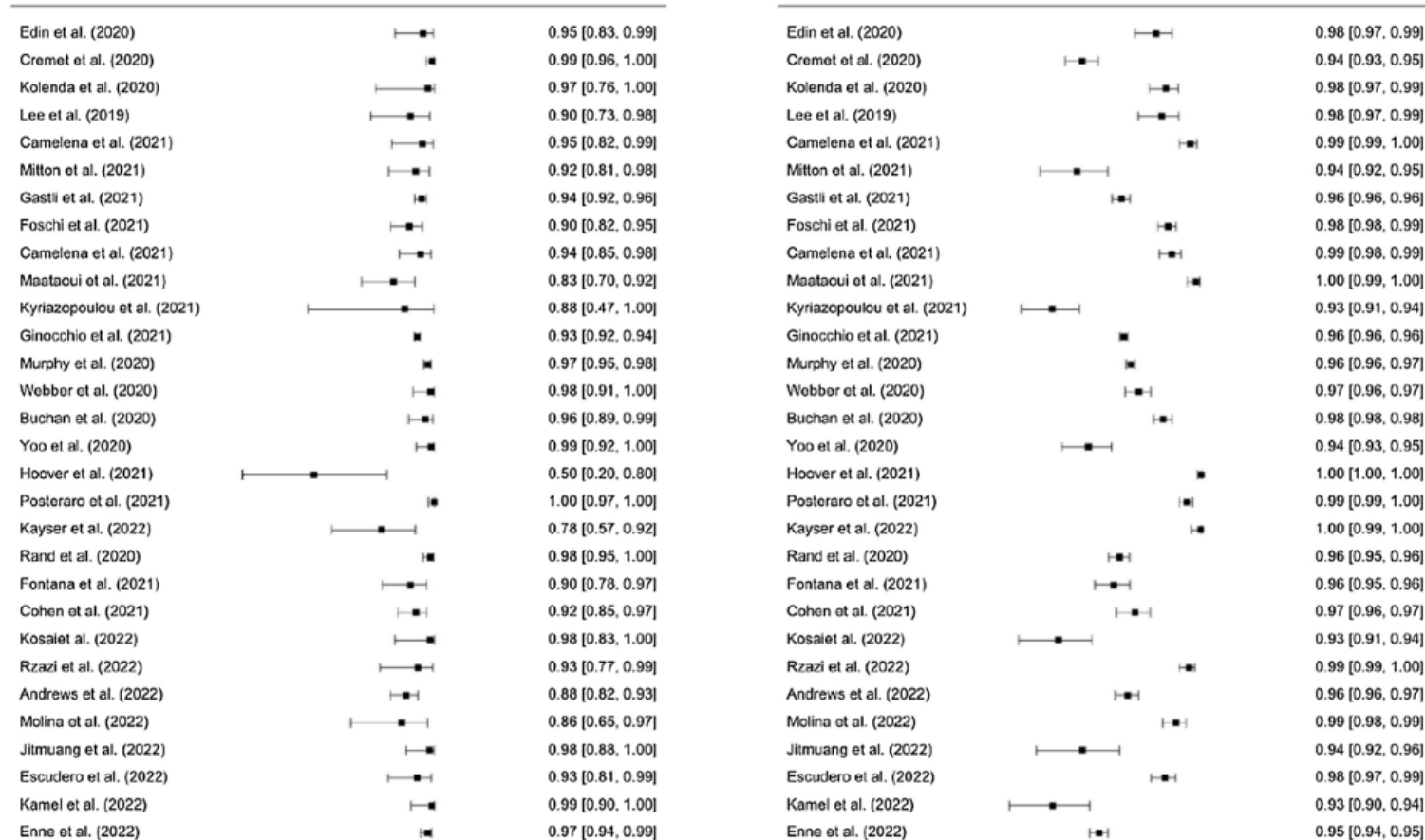
Diagnostic utility by multiplex PCR exam

50-99%

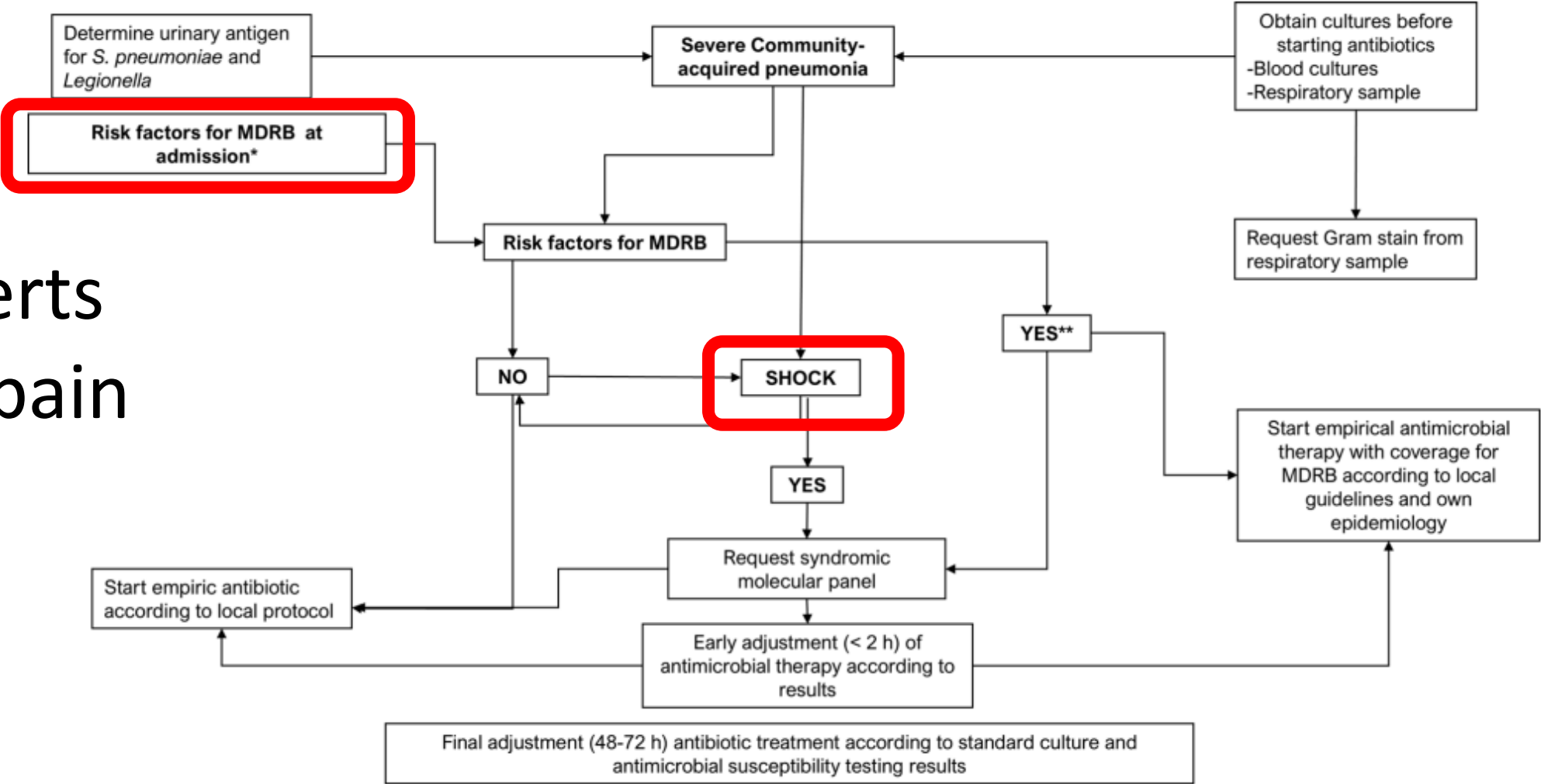
Sensitivity

Specificity

93-100%



If HAP/VAP, testing with syndromic panels is recommended regardless of the risk for MDR bacteria involvement or shock risk factors



11 experts
From Spain

Antibiotics activity Against MDRGNB

Active Variable Not recommended		Enterobacterales					Lactose non-fermenting organisms	
	Typical dosing regimen for serious infections ^{11,110,111}	Extended-spectrum β -lactamase-producing Enterobacterales	AmpC β -lactamase-producing Enterobacterales	Ambler class A carbapenemases (eg, KPC and IMI)	Metallo- β -lactamases (eg, NDM, VIM, and IMP)	Ambler class D carbapenemases (eg, OXA-48)	Difficult-to-treat resistant <i>Pseudomonas aeruginosa</i>	Carbapenem-resistant <i>Acinetobacter baumannii</i>
β-lactam								
Ceftolozane-tazobactam	3 g IV every 8 h, infused over 3 h							
Ceftazidime-avibactam	2.5 g IV every 8 h, infused over 3 h							
Meropenem-vaborbactam	4 g IV every 8 h, infused over 3 h							
Imipenem-relebactam	1.25 g IV every 6 h, infused over 30 min							
Cefiderocol	2 g IV every 8 h, infused over 3 h							
Ceftazidime-avibactam and aztreonam	Ceftazidime-avibactam: 2.5 g IV every 8 h, infused over 3 h plus aztreonam: 2 g IV every 8 h, infused over 3 h*							
Aztreonam-avibactam	2 g/0.67 g loading dose then 1.5 g/0.5 g every 6 h, infused over 3 h							
Cefepime-enmetazobactam	2 g/0.5 g every 8 h, infused over 4 h							
Sulbactam-durlobactam†	1 g of each drug IV every 6 h, infused over 3 h†							
Tetracycline derivative								
Eravacycline	1 mg per kg IV every 12 h							

Lancet. 2025 Jan 18;
405(10474):257-272.

Risk factors for carbapenem-resistant Gram-negative bacterial infections

Design

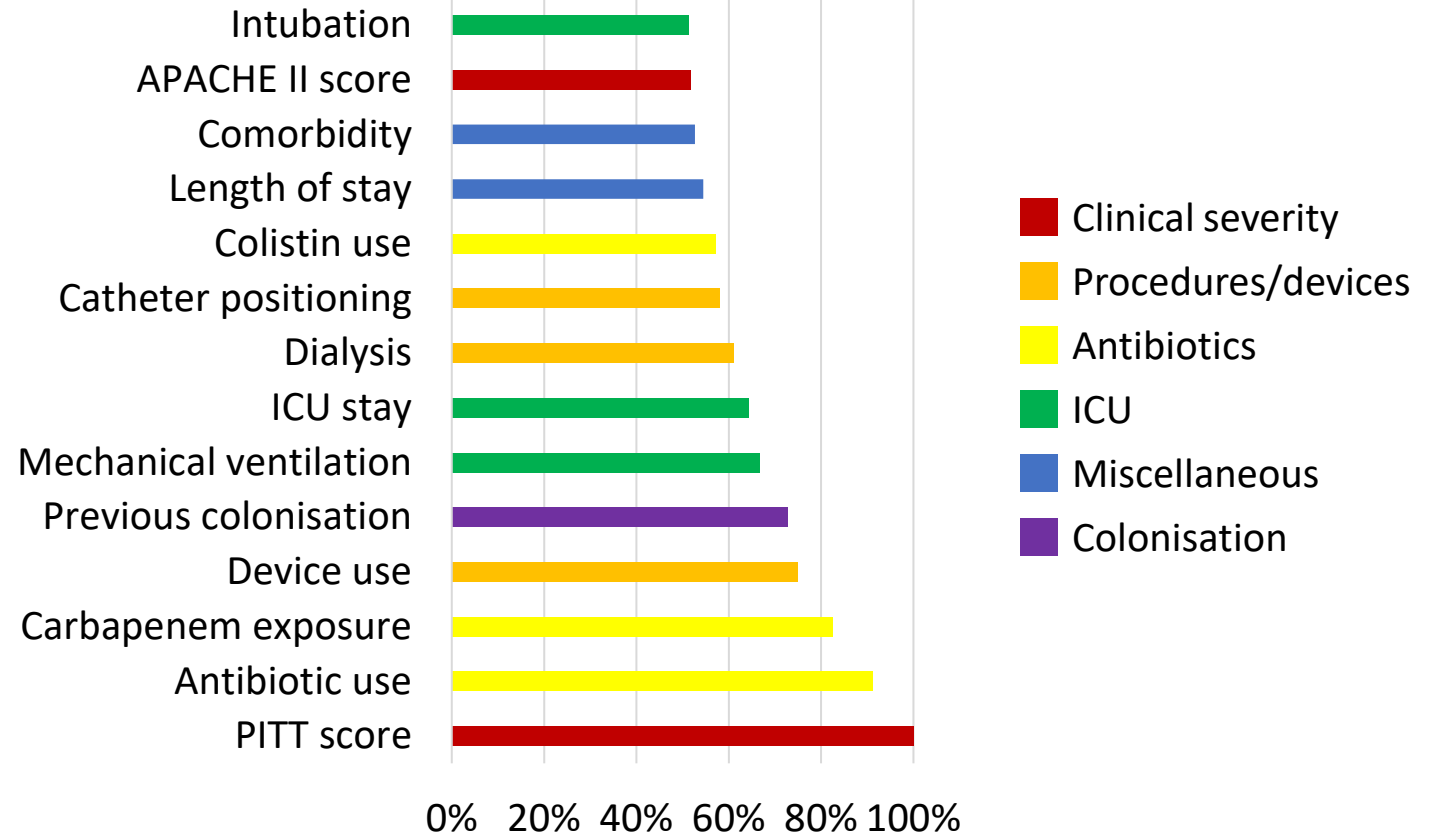
Systematic review of studies reporting risk factors associated with infections due to carbapenem-resistant Gram-negative pathogens in hospitalized patients

- 92 studies identified



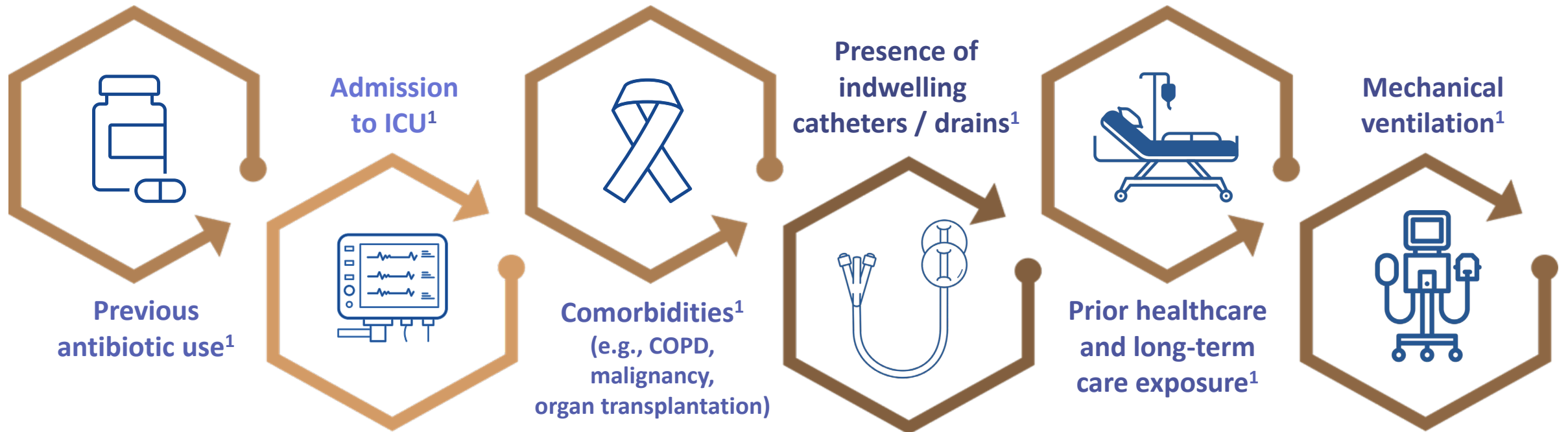
Outcomes

Figure 2 from Palacios-Baena Z 2021





Who is in danger?



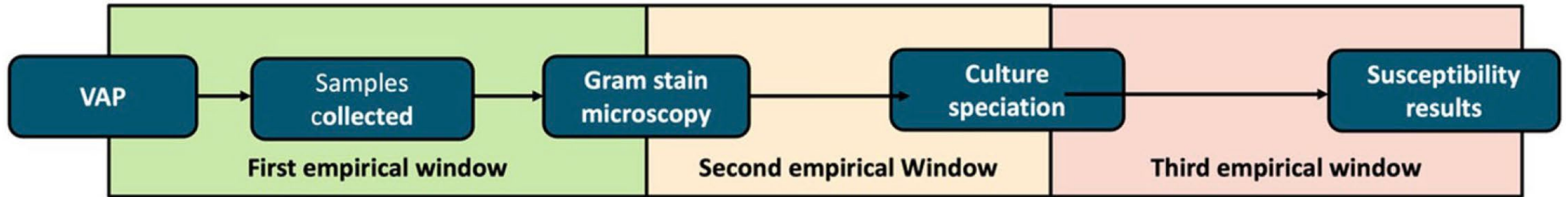
COPD, chronic obstructive pulmonary disease; ICU, intensive care unit.

1. Bassetti M, *et al.* Patient specific risk stratification for antimicrobial resistance and possible treatment strategies in Gram-negative bacterial infections. *Expert Rev Anti Infect Ther.* 2017;15(1):55–65.

Icons from The Noun Project.

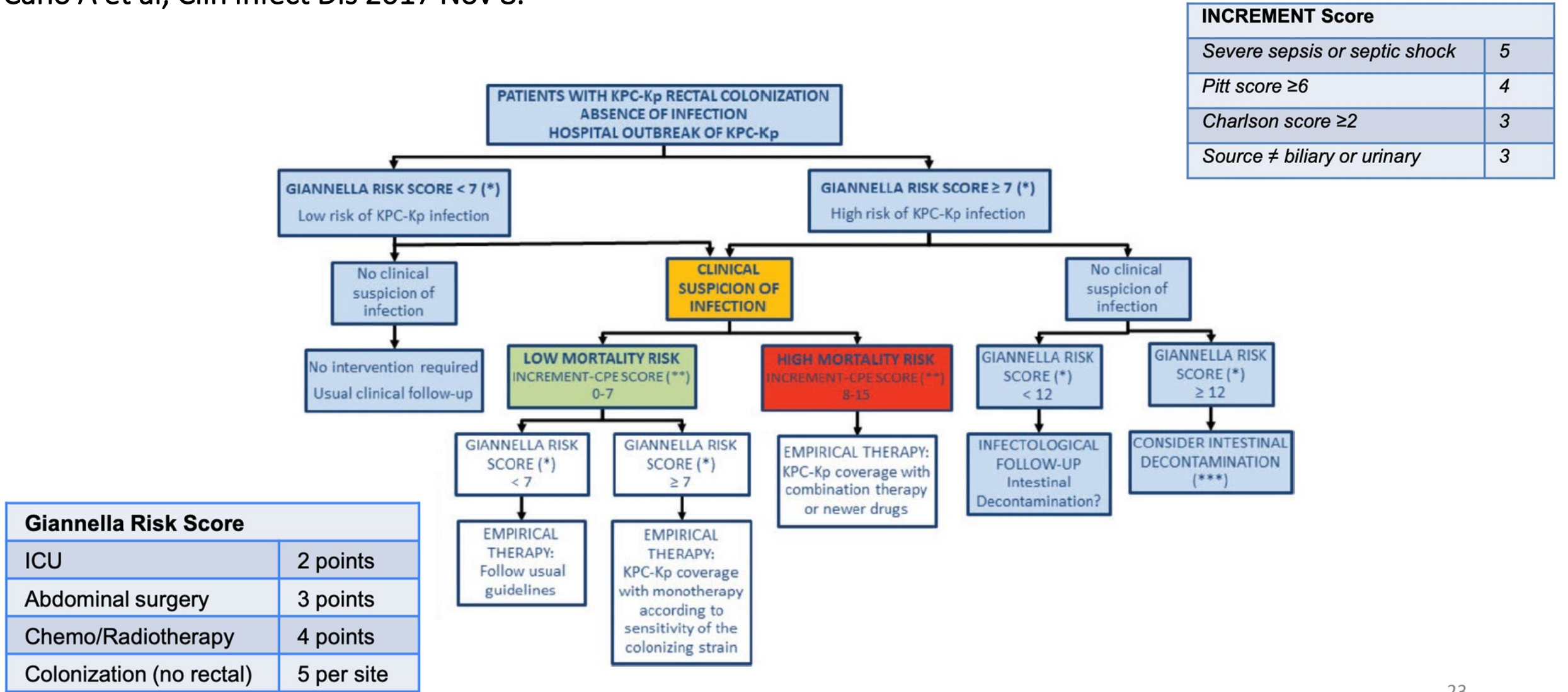
Empiric Therapeutic Window in VAP

Amani Alnimir Infect Dis Ther 2023



Risks of Infection & Mortality among Patients Colonized with KPC-Kp: Validation of Scores & Proposal for Management

Cano A et al, Clin Infect Dis 2017 Nov 8.



MDR colonization and immunocompromised patients

Article

Development of a Clinical Score to Stratify the Risk for Carbapenem-Resistant Enterobacterales Bacteremia in Patients with Cancer and Hematopoietic Stem Cell Transplantation

Variables	Univariate Analysis		Multivariate Analysis		Points
	OR (95%CI)	p-Value	OR (95%CI)	p-Value	
Recent carbapenems use	4.1 (2.3–7.7)	0.001			
➡ >7 days of antibiotic use prior to bacteremia	4.6 (2.6–4.1)	0.001	4.65 (2.29–9.46)	<0.001	2
Neutropenia	3.6 (1.6–8.2)	0.002			
➡ ≥10-days of hospitalization prior to bacteremia	4.4 (2.3–8.6)	0.001	4.03 (1.88–8.66)	<0.001	2
Recent intensive care unit admission	3 (1.4–6.8)	0.006			
Central venous catheter in place	3.4 (1.7–6.7)	0.002			
Previous colonization with KPC-CPE	10.6 (4.4–25.5)	0.001			
➡ Recent colonization with KPC-CPE	24.9 (1.3–55)	0.001	33.08 (11.7–93.25)	<0.001	5

Score	Sensitivity	Specificity	PPV	NPV
7	35%	98%	77%	91%

Impact of Inappropriate Initial Antibiotics (IIAT)

In critically ill patients with GN sepsis/septic shock (N=1076, St.Louis, U.S.A.)

- Retrospective, single-center study, adult ICU at Barnes-Jewish Hospital (2008-2012)
- **In-hospital mortality: 351, 29.2%**
- **Risk factors for mortality:**
 - Older (66.5 vs 63 ys)
 - Vasopressors (OR 2.79)
 - Ventilated (OR 3.06)
 - MDR infections (10.% vs 4%, $p < 0.001$)
 - IIAT (43.4% vs 14.6%, $p < 0.001$)

Inappropriate Initial Antimicrobial Therapy ↑ Mortality 3.8-fold

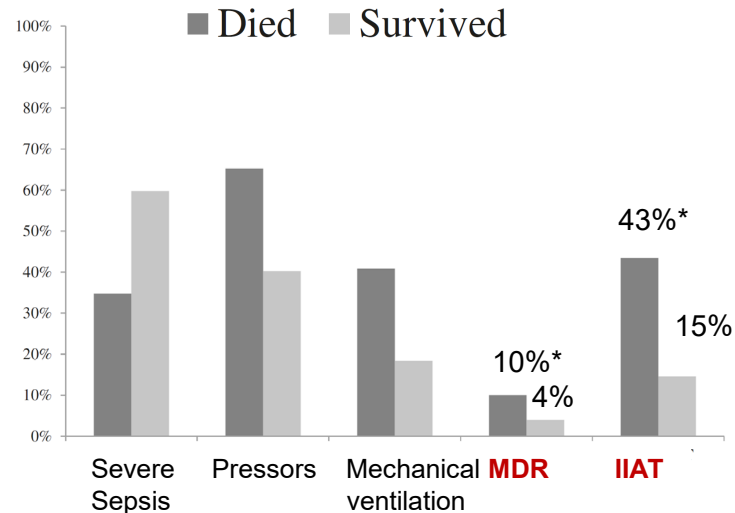


Figure 1 Sepsis severity, resistance and initial treatment. IIAT, initially appropriate antibiotic therapy; MDR, multidrug resistant.

* $P < 0.001$ for each comparison.

Multidrug Resistant ↑ IIAT 13-fold

Table 3 Predictors of hospital mortality^a

	Odds ratio	95% confidence interval	P value
Non-IIAT	3.872	2.770 to 5.413	<0.001
Chronic liver disease	1.942	1.319 to 2.860	0.001
Septic shock	1.846	1.335 to 2.553	<0.001
Pneumonia	1.766	1.237 to 2.522	0.002
Mechanical ventilation	1.669	1.172 to 2.376	0.005
APACHE II score (per 1 point)	1.076	1.047 to 1.105	<0.001
Surgery	0.701	0.560 to 0.879	0.002
Admitted from home	0.677	0.489 to 0.936	0.018
Urosepsis	0.675	0.469 to 0.972	0.034

IIAT: Initially appropriate antibiotic therapy

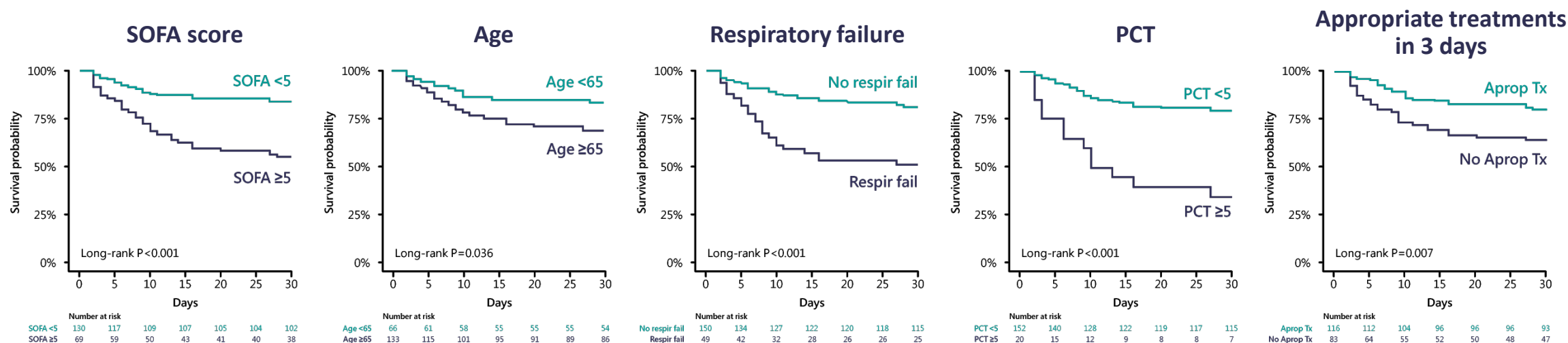
Table 4 Predictors of receiving initially inappropriate antibiotic therapy^a

	Odds ratio	95% confidence interval	P value
Multidrug resistant	13.05	7.00-24.31	<0.001
HIV	3.64	1.02-12.95	0.046
Transferred from another hospital	2.86	2.00-4.08	<0.001
Nursing home resident	2.28	1.35-3.84	0.002
Prior antibiotics	2.06	1.47-2.87	<0.001
Polymicrobial	1.90	1.30-2.77	0.001
Congestive heart failure	1.61	1.11-2.35	0.013
APACHE II score (per 1 point)	1.05	1.02-1.07	<0.001

Inadequate treatment within 3 days may increase the risk of 30-day mortality in patients with CRKP infection

- Primary outcome: the 7-, 15-, and 30-day mortality rates were 13.6% (26/191), 22.0% (42/191), and 26.7% (51/191).
- The risk factors for 30-day mortality among adult patients with CRKP infection were established to be the presence of **respiratory failure, high SOFA score, PCT >5 ng/mL, age >65 years, and non-appropriate treatments in 3 days.**

Kaplan–Meier survival plots of risk factor



A total 199 adult patients diagnosed with CRKP infection in a tertiary hospital in Shanghai between September 2019 and March 2021 were included. The primary outcome was the time to all-cause death up to 30 days from CRKP detection. The secondary outcome was the effect of treatment on CRKP infection, which was defined as effective (cure or improvement in the infection status) or noneffective (stable or deterioration of infection) within 7 and 15 days, based on the clinical, radiological, and laboratory findings.

Adapted from Chen J, et al. 2022

CRKP = carbapenem-resistant *Klebsiella pneumoniae*; SOFA = Sequential Organ Failure Assessment; PCT = procalcitonin.

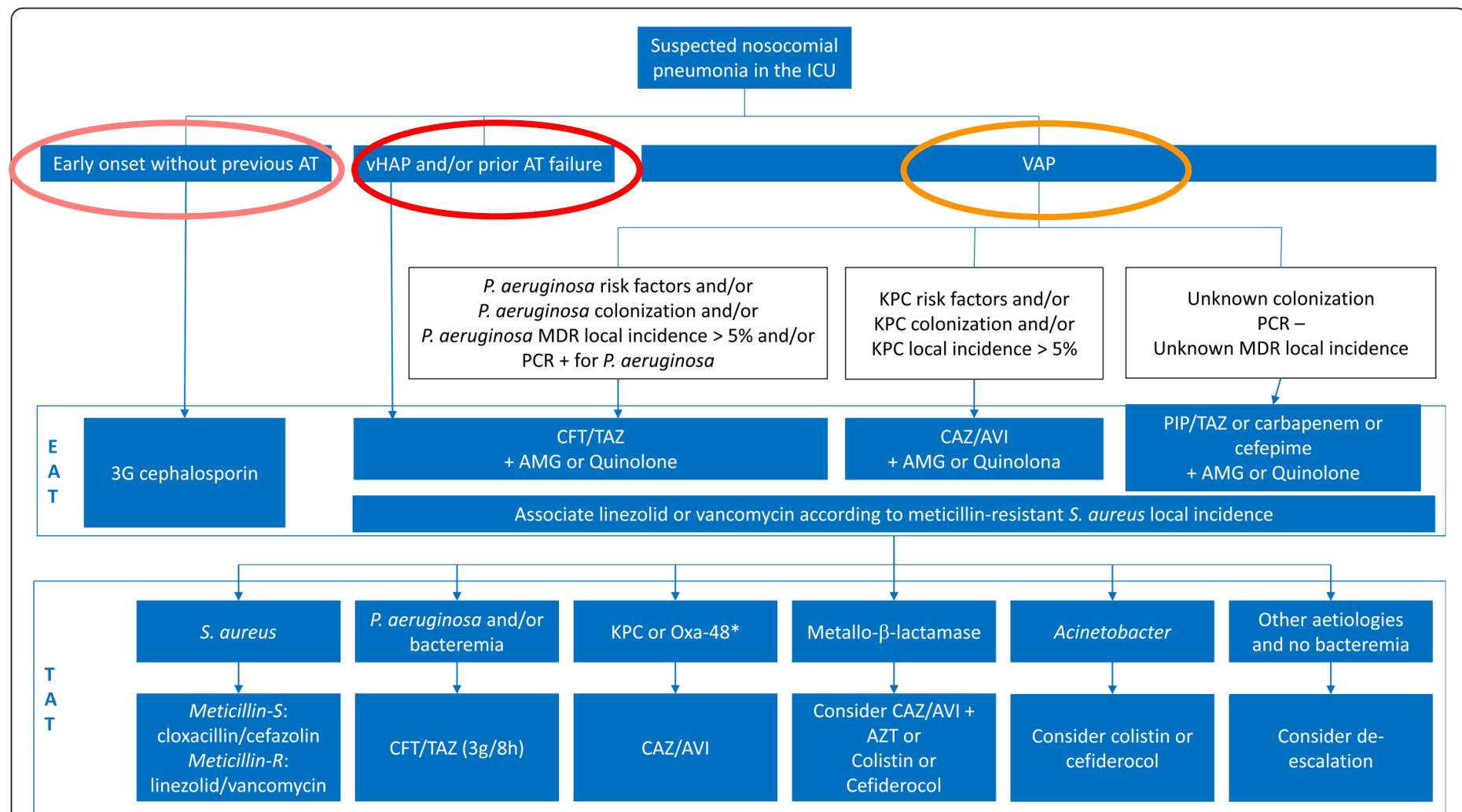


Fig. 2 PANNUCI algorithm. From empirical to targeted treatment on nosocomial pneumonia in ICU. After analyzing the onset, the previous use of antimicrobials or clinical condition (vHAP or VAP), empirical antimicrobial therapy is chosen based on risk factors, previous colonization, local flora and/or use of rapid techniques. Therefore, targeted therapy is selected depending on the type of microorganism isolated and the possible advantages of one antimicrobial over others. AT, antimicrobial therapy; vHAP, ventilated hospital-acquired pneumonia; VAP, ventilator-associated pneumonia; MDR, multidrug-resistant; PCR, polymerase chain reaction; CFT/TAZ, ceftolozane/tazobactam; CAZ/AVI, ceftazidime/avibactam; PIP/TAZ, piperacillin/tazobactam; AMG, aminoglycoside; AZT, aztreonam; EAT, empirical antimicrobial treatment; TAT, targeted antimicrobial treatment; OXA-48, OXA-48 carbapenemase; KPC, *Klebsiella pneumoniae* carbapenemase; R, resistance. *If Oxa-48 susceptible to CAZ/AVI

Septic patient

Yes

Yes

Clinical entities

Urinary tract infections
Intra-abdominal infections
Nosocomial pneumonia
Associated or suspected bacteremia

Yes

Risk factors for Carbapenemase- and/or ESBL-producing *Enterobacteriaceae*

- Previously known colonization
- Broad-spectrum antimicrobial therapy during previous 90 days (cephalosporins/fluoroquinolones)
- History of prolonged hospitalization and/or long-term care facilities
- Invasive devices
- Immunosuppression
- Current or prior ICU admission
- Local epidemiology, outbreak
- Travel from high endemic area (*Enterobacteriaceae*)

±

Associated comorbidities

- Diabetes
- COPD
- Moderate/severe renal/liver disease
- Immunosuppression/neutropenia
- Elderly patients
- Solid tumor
- Structural lung disease
- Organ transplantation
- Hemodialysis

Local epidemiological data in clinical isolates

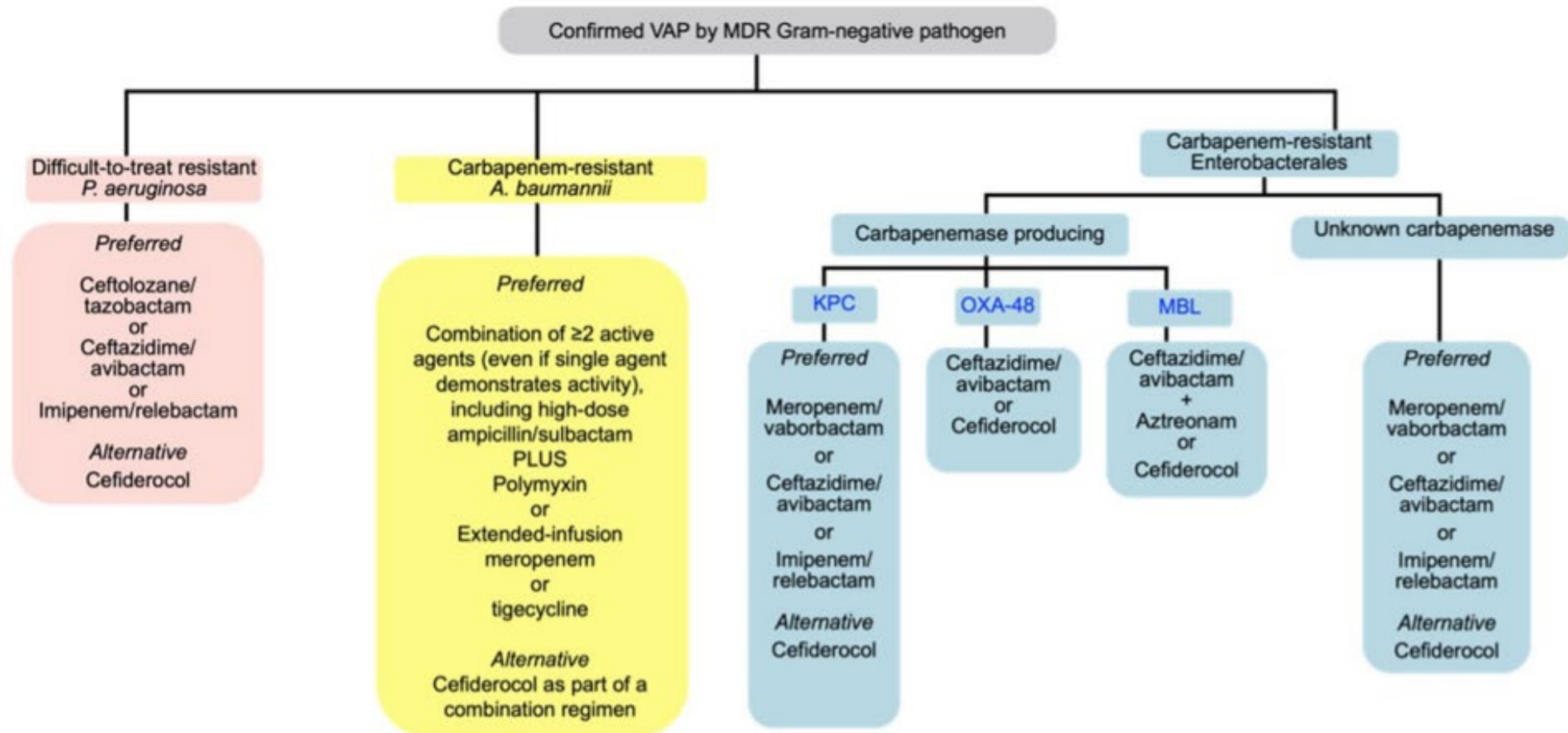
Carbapenem-resistant
Enterobacteriaceae >20%
ESBL-producing *Enterobacteriaceae*
>20% in *Escherichia coli* and/or
Klebsiella spp

Empiric therapy
Ceftazidime/avibactam

Obtain adequate microbiological sample for culture

Possible value
of rapid diagnostic tests

Treatment of MDRO in VAP by IDSA

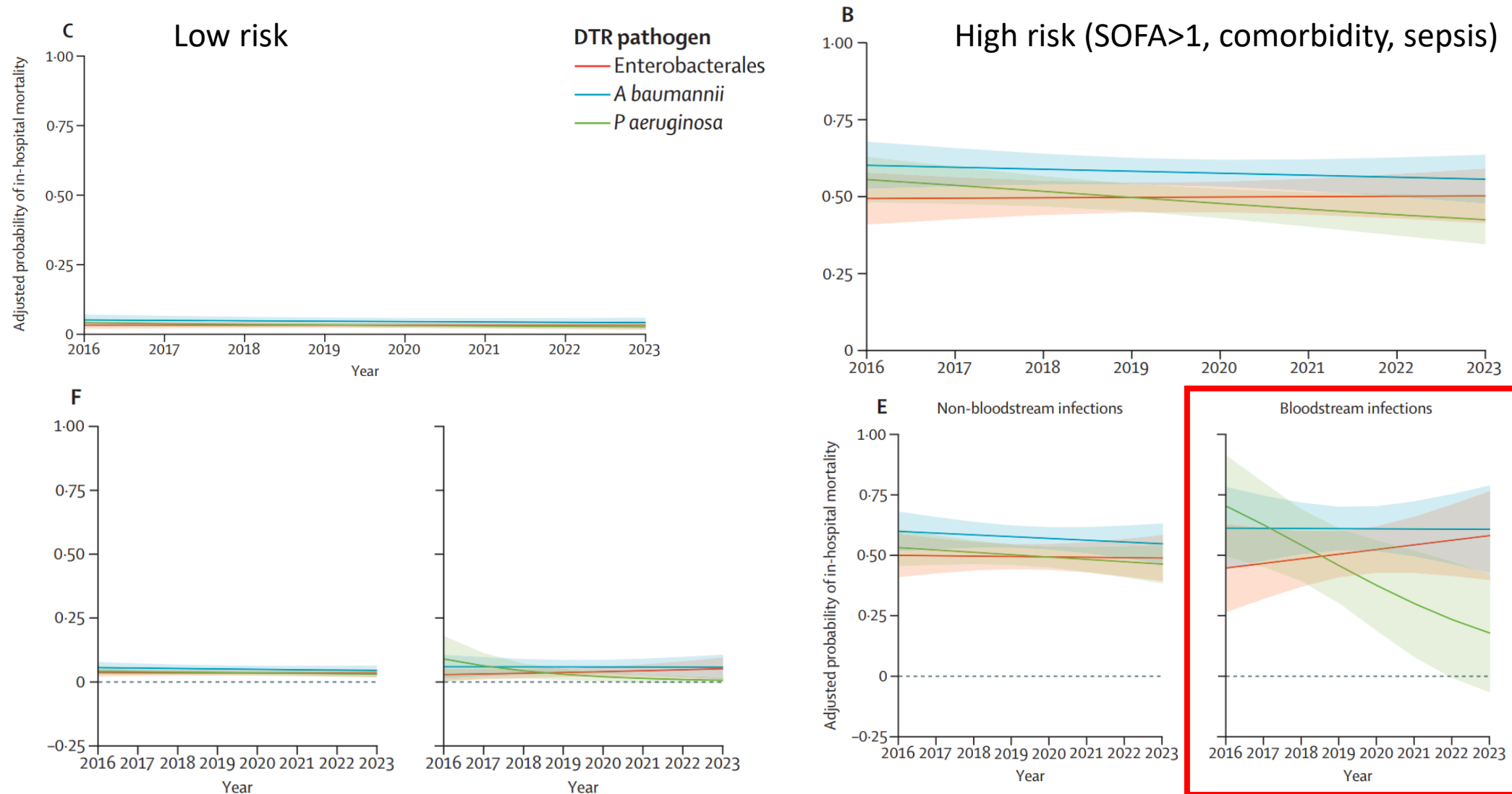


ESCMID 2022 & IDSA 2023 Guidance for the Treatment of Serious GNB infections

Agent	ESBL/AmpC producers	CRE - class A carbapenemases (e.g.; KPC)	CRE - class D carbapenemases (e.g.; OXA-48)	CRE - MBL-producers (e.g.; NDM)	DTR <i>P. aeruginosa</i> [#]
Carbapenems	preferred*	–	–	–	–
Ceftolozane-tazobactam	against (unless polymicrobial)	–	–	–	preferred*
Ceftazidime-avibactam [†]	optional* (stewardship)	preferred*	preferred*	–	preferred*
Meropenem-vaborbactam	optional* (stewardship)	preferred*	–	–	–
Imipenem-relebactam	optional* (stewardship)	preferred*	–	–	preferred*
Cefiderocol	optional* (stewardship)	optional (while waiting for AST)	optional (while waiting for AST)	preferred*	optional*
Polymyxins	–	–	–	–	–

Mortality from DTR-GNB in US (2016-2023)

Annual 4.5% mortality reduction in PA bacteremia



Abx prescription

AB

Cefiderocol

Old

Eravacycline

Enterobacterales

CAZ-AVI

Mer-Vab

Cefiderocol

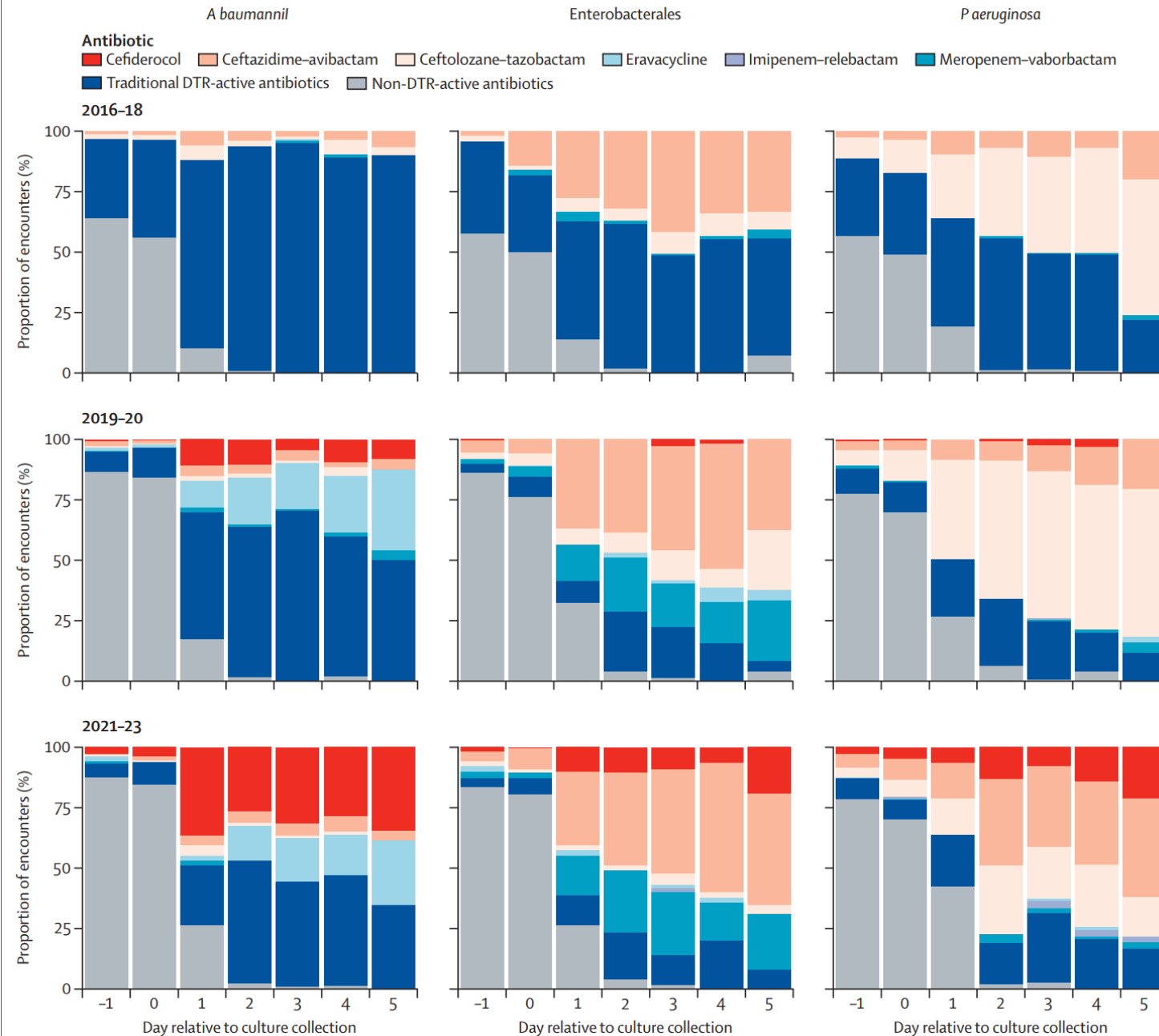
PA

CAZ-AVI

Old

Cefiderocol

CT



Comparison of official recommendations - CRE

Similarities	ESCMID	IDSA
Non-severe	Old antibiotics with in vitro S	Quinolones, TMP-SMX, single dose aminoglycoside
Metallo-beta-Lactamase (MBL)	Cefiderocol (conditional); ceftazidime-avibactam + aztreonam	Cefiderocol or ceftazidime-avibactam + aztreonam
Tigecycline	Not in BSI, HAP/VAP	Avoided in non-IAI
Combination therapy	Not routinely recommended	Not routinely recommended

Comparison of official recommendations - CRE

Differences	ESCMID	IDSA
Severe	Ceftazidime-avibactam or meropenem-vaborbactam (conditional); no recommendation on imipenem-relebactam, cefiderocol	Ceftazidime-avibactam, Meropenem-vaborbactam, Imipenem-relebactam, or cefiderocol for severe UTI
Complicated UTI	Aminoglycosides over tigecycline	Ceftazidime-avibactam, meropenem-vaborbactam, imipenem-relebactam, or cefiderocol
Eravacycline	No evidence	In IAI only
Combination therapy with carbapenem	Use when MIC $\leq$ 8 mg/L and with extended infusion	-

Ceftazidime-avibactam for Treatment of CRE BSI (Meta-analysis, 11 studies, 1205 patients)

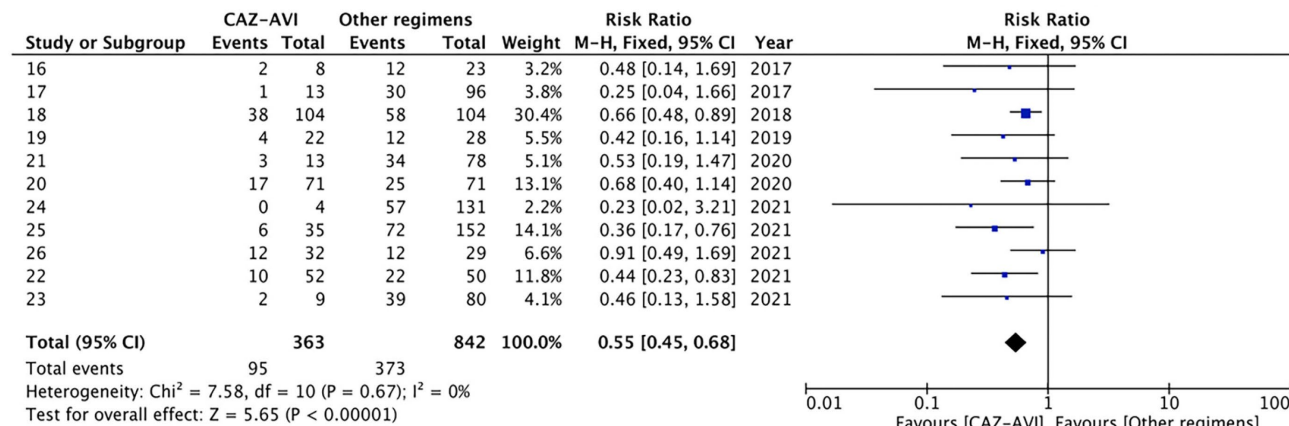
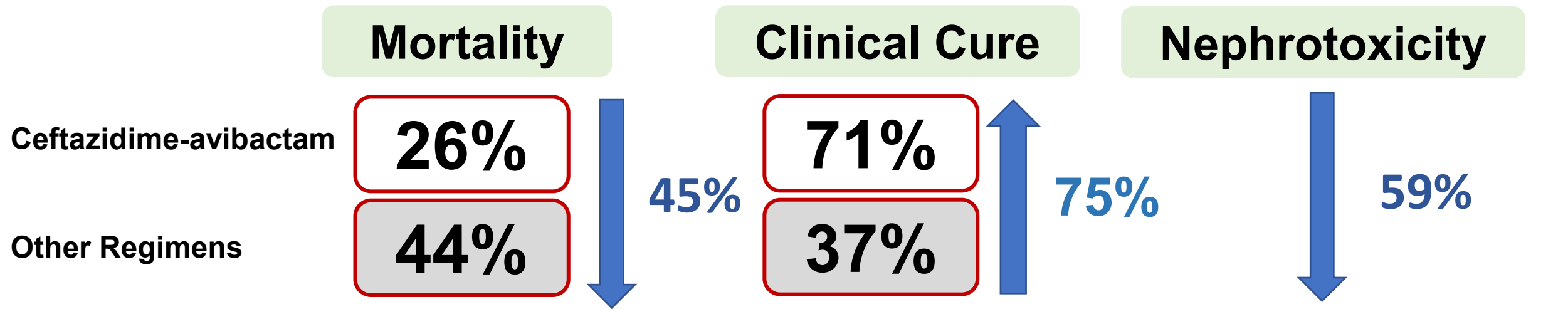


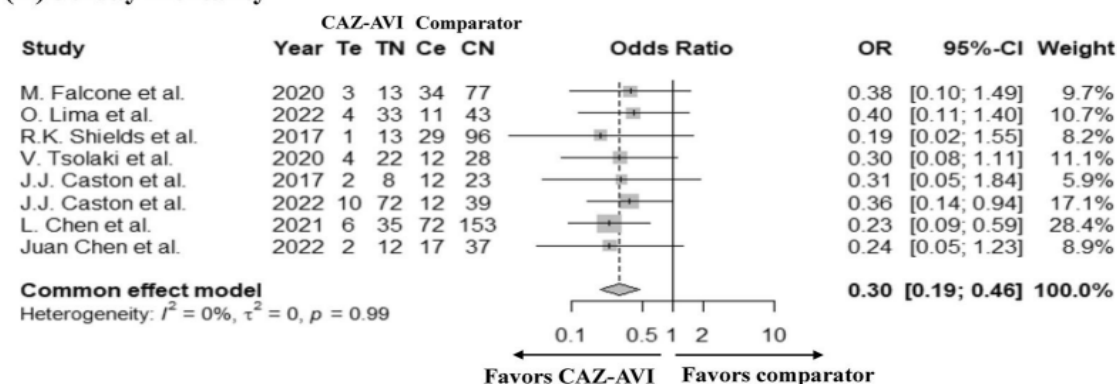
FIG 2 Thirty-day mortality of the CAZ-AVI regimens compared with controls in CRE BSI.

- **Lower 30-day mortality rate:** Overall: **45% reduction** (RR = 0.55, 95% CI:0.45-0.68, P < 0.00001)**vs Colistin-based regimen: 52% reduction** (RR = 0.48, 95% CI 0.33-0.69, P<0.0001).
- **Higher clinical cure rate** (RR = 1.75, 95% CI of 1.57-2.18, P < 0.00001)
- **Lower nephrotoxicity rate: 59% reduction** (RR = 0.41, 95% CI of 0.20-0.84, P = 0.02)

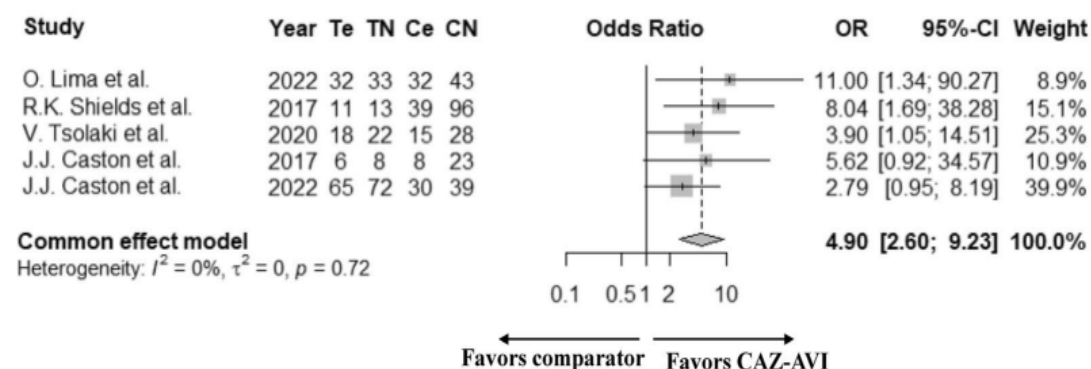
Ceftazidime-Avibactam in the Treatment of Patients with Bacteremia or Nosocomial Pneumonia: A Systematic Review and Meta-analysis

- The databases included in the search, until November 7, 2022, were Embase and PubMed. A total of 24 studies (retrospective: 22, prospective: 2) with separate outcomes for patients with bacteremia or pneumonia were included.
- This SLR and meta-analysis synthesized clinical outcome data and presents quantitative and qualitative evidence that may be considered while using CAZ-AVI for the treatment of patients with bacteremia and nosocomial pneumonia caused by **non-MBL producing CRE Infect Dis and/or MDR P. aeruginosa**

(A) 30-day mortality



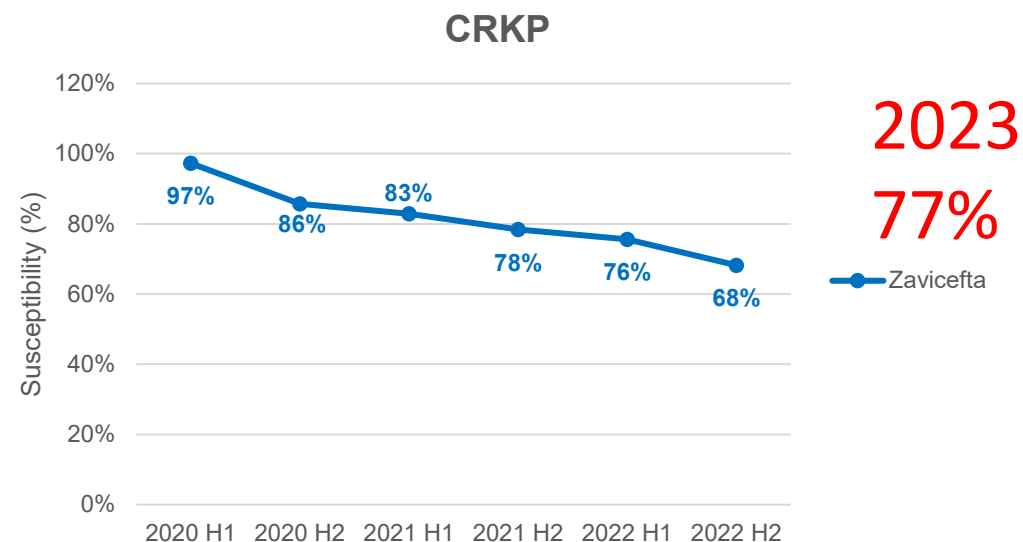
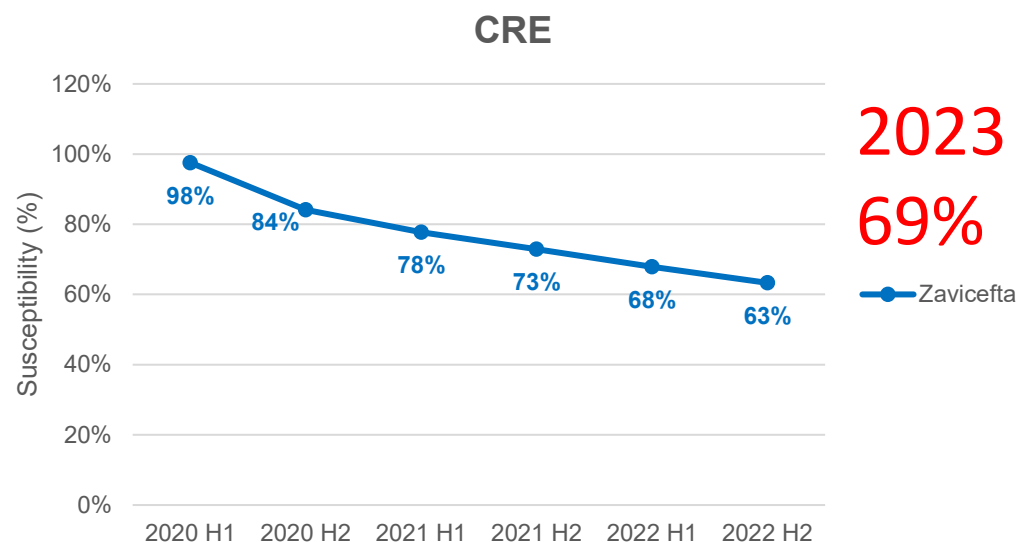
(B) Clinical cure



Question 3.7: What is the likelihood of the emergence of resistance of CRE isolates to the newer β -lactam agents when used to treat CRE infections?

- Suggested approach: The emergence of resistance is a concern with all β -lactams used to treat CRE infections. Available data suggest the frequency may be **highest for ceftazidime-avibactam**.

2020~2023年度台北榮總藥物敏感性報告趨勢圖



7. 全院MDRO (% Susceptible):

2025

菌名	株數	CT	CZA	MEM	MXF	TGC
<i>Klebsiella pneumoniae</i> (CRE)	446	9	73	38	18	75
<i>Escherichia coli</i> (CRE)	120	14	74	52	16	99
<i>Serratia marcescens</i> (CRE)	80	-	19	30	14	37
<i>Enterobacter hormaechei</i> (CRE)	52	6	46	44	31	52
<i>Klebsiella oxytoca</i> (CRE)	48	0	8	6	0	72
<i>Enterobacter cloacae</i> (CRE)	28	7	43	39	43	71

(CRE)：對 carbapenem 類中的 imipenem 或 ertapenem 任一抗生素具抗藥性。

菌名	株數	CT	CZA	MI	SFP
<i>Pseudomonas aeruginosa</i> (CR)	347	83	84	-	44
<i>Acinetobacter baumannii</i> (CR)	148	-	-	61	41
<i>Acinetobacter nosocomialis</i> (CR)	25	-	-	90	92

(CR)：對 carbapenem 類中的 imipenem 或 meropenem 任一抗生素具抗藥性。

菌名	株數	DAP^	LNZ	TGC
<i>Enterococcus faecium</i> (VRE)	255	99	98	99

^: %SDD

菌名	株數	LEV	MI	MXF
<i>Staphylococcus aureus</i> (MRSA)	25	14	76	14

抗生素名稱

CT: Ceftolozane/Tazobactam

CZA: Ceftazidime/Avibactam

DAP: Daptomycin

LEV: Levofloxacin

LNZ: Linezolid

MEM: Meropenem

MI: Minocycline

MXF: Moxifloxacin

SFP: Cefoperazone/Sulbactam

TGC: Tigecycline

- : Not available, or Method limitation

8. 全院Candida from BLOOD CULTURE (% Susceptible):

2025

菌名	株數	AMB	CAS	5FC	FCA	MF	VRC
<i>Candida albicans</i>	65	100	100	100	98	100	97
<i>Candida glabrata</i>	44	100	-	100	-	100	-
<i>Candida tropicalis</i>	40	100	100	95	85	100	83
<i>Candida parapsilosis</i>	37	97	100	100	70	100	70

抗真菌藥物名稱

AMB: Amphotericin B

CAS: Caspofungin

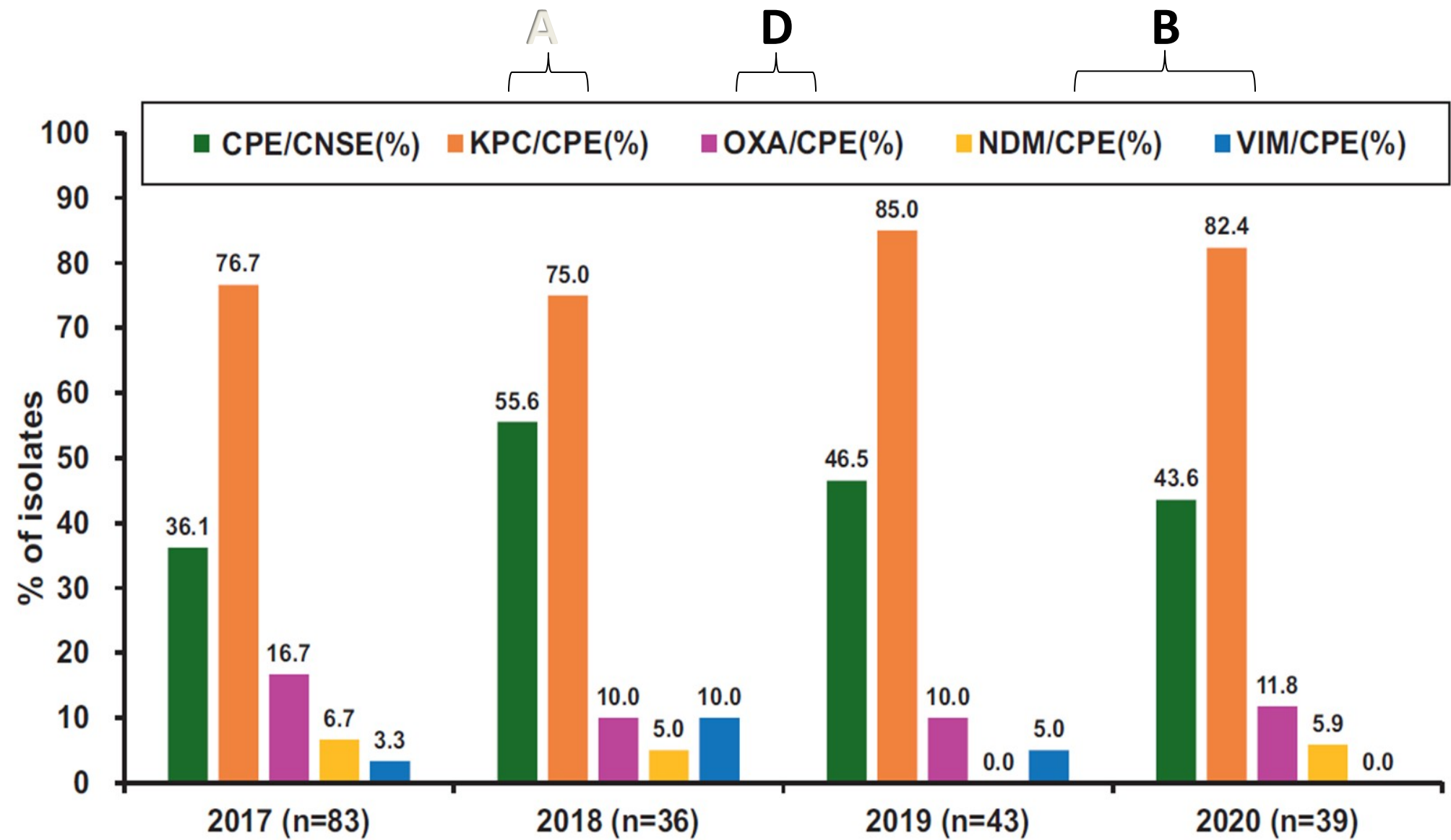
5FC: Flucytosine

FCA: Fluconazole

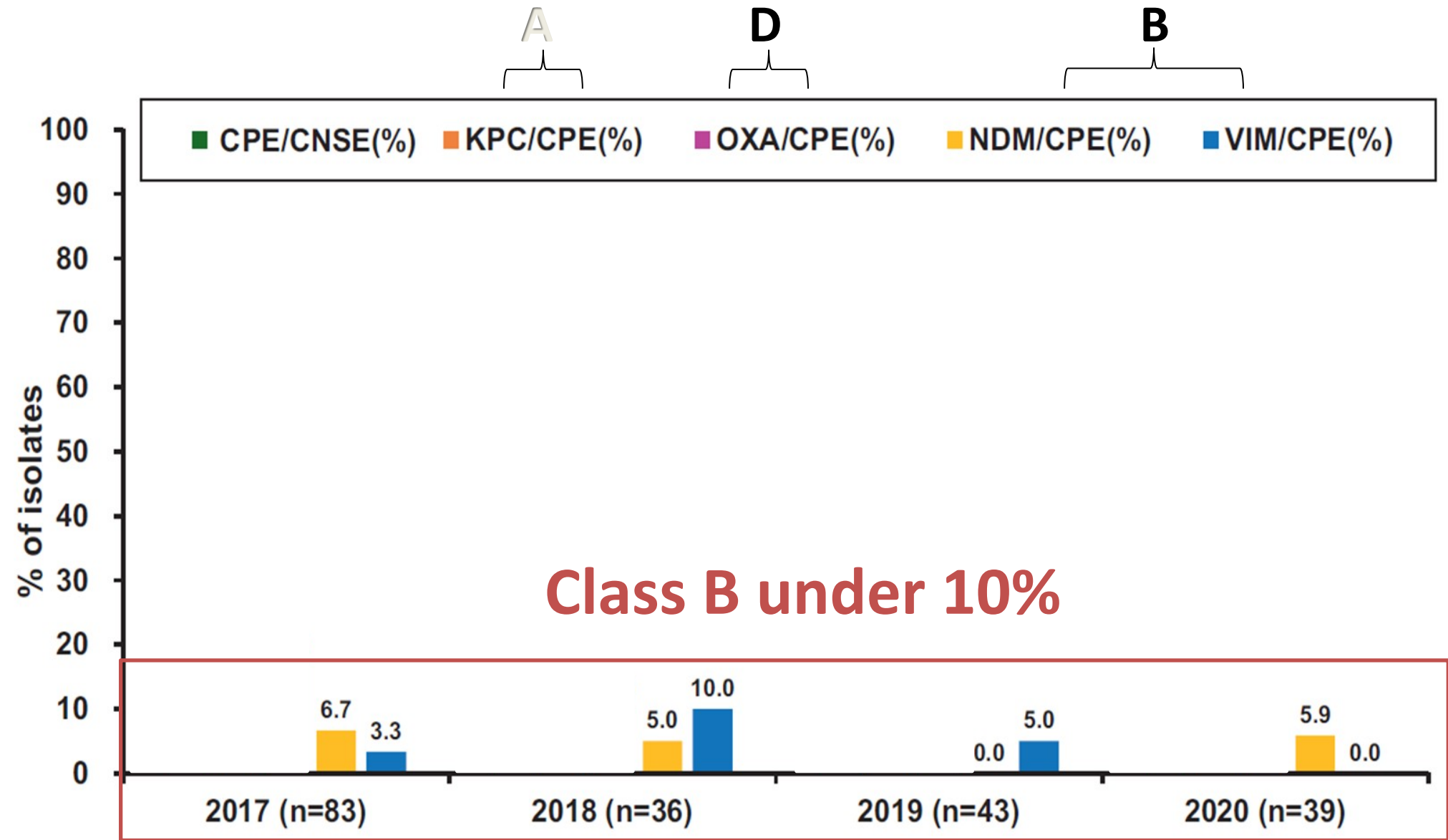
MF: Micafungin

VRC: Voriconazole

Epidemiological situation of CRE in Taiwan (SMART study)

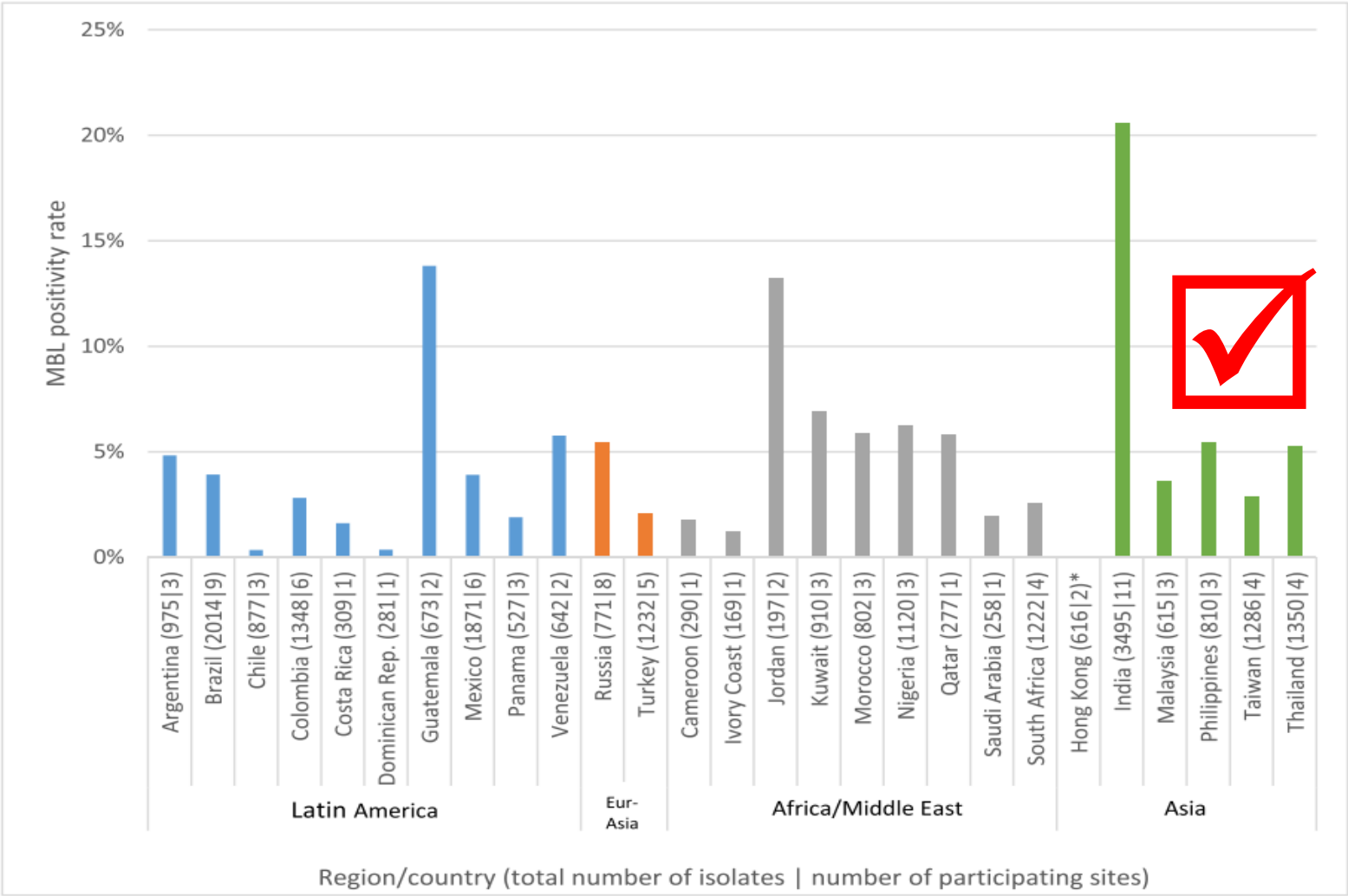


Epidemiological situation of CRE in Taiwan (SMART study)

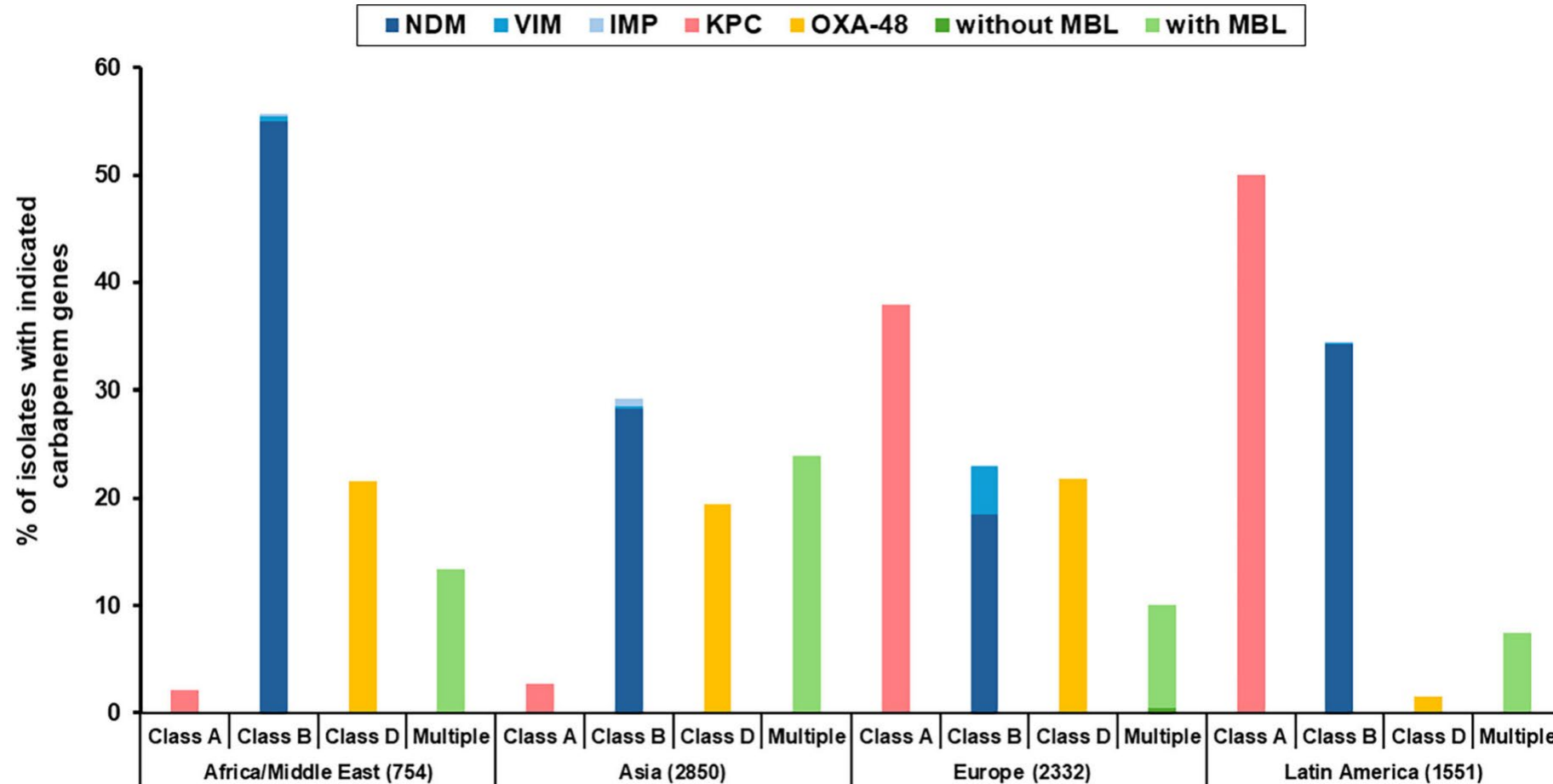


MBL prevalence: TW: around 2.5% (ATLAS study)

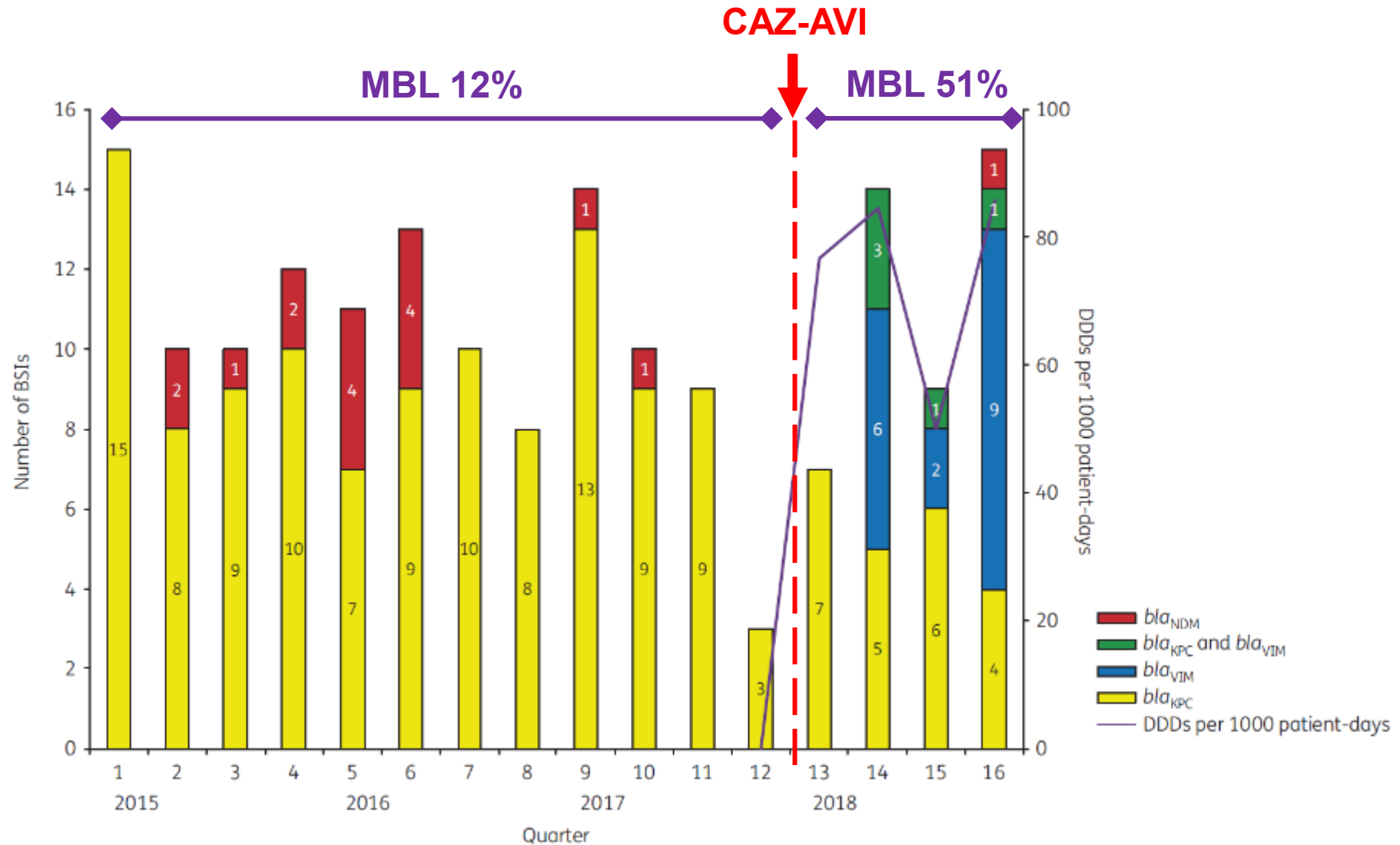
Fig. 2 Incidence of MBL positivity among isolates collected for the ATLAS surveillance project in 2019–2021 by country. Asterisk indicates no MBL producers collected in Hong Kong



Global CRE genes by ATLAS (2019-2023)



Ceftazidime-avibactam use and types of carbapenemase production



Risk factors for ceftazidime-avibactam clinical failure in CRE infections

Retrospective in US, clinical failure rate 45%

Risk factor ^a	Value ^b for patients with:		P value	Multivariate P value (OR, 95% CI) ^c
	Success (n = 42)	Failure (n = 35)		
Demographics and underlying conditions				
Male gender	29 (69)	18 (51)	0.16	
Age (yr)	64 (19–91)	59 (26–79)	0.22	
Solid-organ transplant recipient	9 (21)	9 (26)	0.79	
Charlson Comorbidity Index	4 (0–10)	5 (0–10)	0.86	
Severity of illness				
ICU at disease onset	17 (40)	27 (77)	0.001	Excluded
Renal replacement therapy	3 (7)	13 (37)	0.002	0.046 (4.78, 1.03–22.2)
SOFA score	5 (0–18)	8 (0–20)	0.0007	0.13
SAPS II score	37 (8–62)	43 (17–81)	0.04	0.98
Infection characteristics				
Infection type				
Pneumonia	12 (29)	21 (60)	0.01	0.045 (3.09, 1.03–9.34)
Primary bacteremia	15 (36)	5 (14)	0.04	
Urinary tract infection	7 (17)	1 (3)	0.07	
Intra-abdominal infection	3 (7)	4 (11)	0.70	
Skin/soft tissue infection	4 (10)	2 (6)	0.68	
Other	1 (2)	2 (6)	0.59	

- CRE: KP 78% (60/77), E coli 12% (9/77), Enterobacter cloacae 6% (5/77), and others.

The presence of KPC-3 variant was associated with ceftazidime/avibactam resistance ($P < 0.0001$)

- One KPC-3 variant-carrying isolate was resistant to both ceftazidime/avibactam and imipenem (MICs = 16 and 8 µg/ml, respectively).

Associations between imipenem, Imipenem/relebactam, ceftazidime, and ceftazidime/avibactam MICs against *KPC-K. pneumoniae* isolates

Factor	Median MIC (µg/mL) (range)											
	Imipenem			Imipenem/relebactam			Ceftazidime			Ceftazidime/avibactam		
	Factor absent	Factor present	P value	Factor absent	Factor present	P value	Factor absent	Factor present	P value	Factor absent	Factor present	P value
ESBL	8 (0.125-256)	16 (2-128)	0.03	0.25 (0.125-2)	0.5 (0.125-4)	0.39	128 (8-1,024)	256 (8-1,024)	0.002	2 (0.25-512)	1 (0.25-4)	0.03
KPC-3 variant	16 (0.125-256)	0.5 (0.25-8)	0.0007	0.5 (0.125-4)	0.25 (0.125-0.5)	0.31	256 (8-1,024)	256 (128-1,024)	0.26	2 (0.25-16)	128 (16-512)	0.0001
Major OmpK36 mutations	8 (0.125-64)	64 (2-256)	0.0001	0.25 (0.125-1)	1 (0.125-4)	0.0002	128 (8-1,024)	384 (8-1,024)	0.007	1.5 (0.25-256)	2 (0.25-512)	0.09

^aIsolates that carried MBL and OXA-carapenemases were excluded from analysis.

Adapted from Haidar G, et al. 2017

One hundred clinical CRE isolates were selected from previous studies or sequenced in this study, sourced from UPMC and Rutgers-New Jersey Medical School. Geographically, isolates were mostly from the US (88), with a few from India, China, and other countries. MICs for imipenem ± relebactam and ceftazidime ± avibactam were determined using broth microdilution. Antibiotic susceptibility was assessed per CLSI breakpoints, and CRE was defined using revised CDC criteria.

CRE = carbapenem-resistant Enterobacteriaceae; KPC = *Klebsiella pneumoniae* carbapenemase; KPC-K = KPC-producing *Klebsiella pneumoniae*; MIC = minimum inhibitory concentration; ESBL = extended-spectrum β-lactamases; CLSI = Clinical and Laboratory Standards Institute; UPMC = University of Pittsburgh Medical Center; CDC = Centers for Disease Control and Prevention.

Ceftazidime/avibactam resistant clinical strains expressing KPC-2 variants with substitutions in the Ω -loop are emerging

- The **KPC-2 D179N (Ω -Loop hydrogen bond networking changes)**¹ variant when expressed confers resistance to imipenem (MIC 4 $\mu\text{g/mL}$).
- Addition of avibactam overcomes the ceftazidime resistance mediated by KPC-2 but **not that mediated by the Asp179 variants**.

MICs of various BL, BLI and combinations against strains containing KPC-2 Asp179 variants

Strain	MIC			Imipenem/relebactam
	Ceftazidime	Ceftazidime/avibactam	Imipenem	
<i>K. pneumoniae</i> KPC-2	64 $\mu\text{g/mL}$	1 $\mu\text{g/mL}$	16 $\mu\text{g/mL}$	KPC-2 D179N variant MIC = 0.5 $\mu\text{g/mL}$
<i>E. Coli blaKPC-2 179 Asp (WT)</i>	128 $\mu\text{g/mL}$	1 $\mu\text{g/mL}$	8 $\mu\text{g/mL}$	
<i>E. Coli bla179 Asn (D179N)</i>	512 $\mu\text{g/mL}$	16 $\mu\text{g/mL}$	4 $\mu\text{g/mL}$	

Adapted from Barnes MD, et al. 2017

Reference1: Whole-cell viability assays. Mueller-Hinton (MH) agar-dilution MIC measurements were performed according to Clinical and Laboratory Standards Institute (CLSI) guidelines. The MICs are reported as the concentrations at which bacterial growth was no longer observed. Avibactam was tested at a constant 4 $\mu\text{g/mL}$ in combination with its respective antibiotic partners. All MIC measurements were performed at least three times.

BL/BLI = β -lactam/ β -lactamaseinhibitor combinations; D = Aspartic acid; KPC = *Klebsiella pneumoniae* carbapenemases; MIC = minimum inhibitory concentration; N = Asparagine; *E. coli* = *Escherichia coli*; *K. pneumoniae* = *Klebsiella pneumoniae*.

Question 3.8: What is the role of tetracycline derivatives for the treatment of infections caused by CRE?

- Suggested approach: Although β -lactam agents remain preferred treatment options for CRE infections, **tigecycline and eravacycline** are alternative options when β -lactam agents are either not active or unable to be tolerated. The tetracycline derivatives are not suggested for the treatment of CRE urinary tract infections or bloodstream infections.
- Limited clinical data are also available investigating the effectiveness of minocycline against CRE infections, but data suggest a **lower** proportion of CRE isolates are likely to be **susceptible to minocycline** compared to tigecycline or eravacycline. The panel suggests using minocycline with caution for the treatment of CRE infections.

Mechanisms of CZA-resistant KP in Taiwan (VGHTPE)

- 456 CRKP isolates were collected in VGHTPE in 2020
 - 31 (6.8%) are non-CP strains (CPE/CRE = 93.2%)
 - 301 (66%) KPC, 83 (18.2%) OXA-48, 41 (9.0%) MBL
- 35(7.6%) **CZA-resistant CRKP strains** with exposure to CZA were analyzed
 - 20 (57.1%) MBL producers
 - **10 (28.6%) KPC variants**, exhibiting MIC for CZA ranging from 64 to ≥ 256 mg/L.
 - KPC-33 (n = 3), KPC-31 (n = 1), KPC-39 (n = 1), KPC-44 (n = 1), KPC-58 (n = 1), KPC-90 (n = 1), and 2 novel KPC variants.
 - 5 wild type KPC with reduced susceptibility

Question 3.9: What is the role of polymyxins for the treatment of infections caused by CRE?

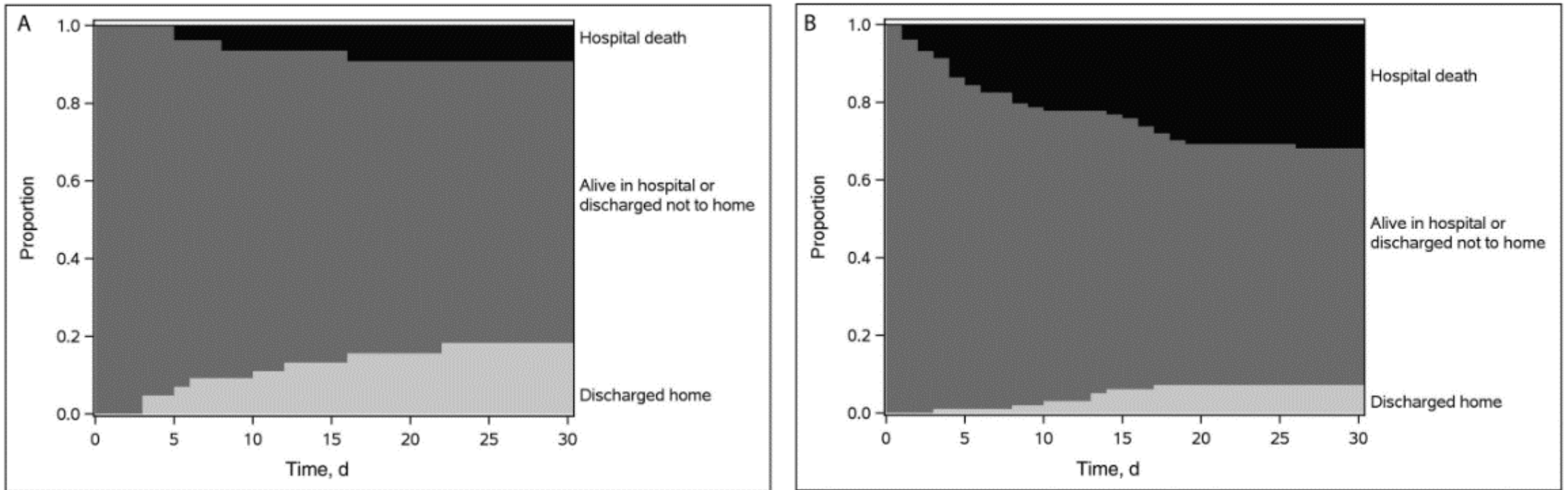
- Suggested approach: Polymyxin B and colistin are **not** suggested for the treatment of infections caused by CRE. Colistin is an alternative agent for **uncomplicated CRE cystitis**.
- **Polymyxin B should not be used as treatment for CRE cystitis**, due to its predominantly nonrenal clearance.

Question 3.10: What is the role of combination antibiotic therapy for the treatment of infections caused by CRE?

- Recommendation: **Combination** antibiotic therapy (ie, the use of a β -lactam agent in combination with an aminoglycoside, fluoroquinolone, or **polymyxin**) is **not routinely recommended** for the treatment of infections caused by CRE.

Ceftazidime-Avibactam(A) VS Colistin(B) in CRE: inverse probability of treatment weighting

137 Patient characteristics: 61 yrs, 47% male, 50% White, 34% critical
46% bacteremia, 21% pneumonia, **97% CRKP**



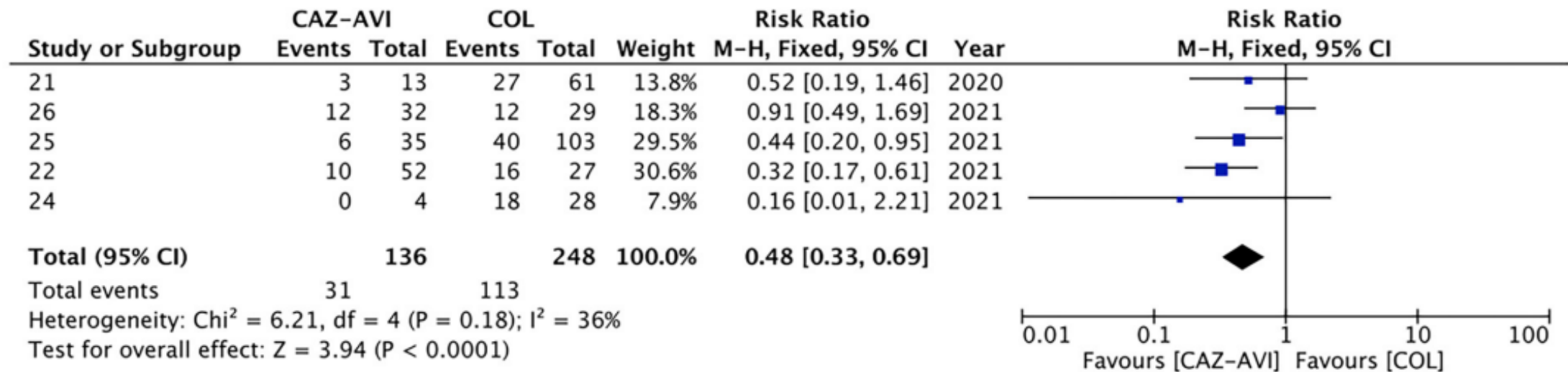
IPTW P=0.001

**CAZ-AVI reduced 30-day mortality (9% vs. 32%) with lower
incident renal failure (4% vs. 13%) vs. Colistin**

Clin Infect Dis. 2018 Jan 6;66(2):163-171.

Observational retrospective analyses are designed to evaluate associations among variables and cannot establish causality; they are not intended for direct comparison with clinical trials

CAZ-AVI VS Colistin in CRE BSI in 30-D mortality



Observational retrospective analyses are designed to evaluate associations among variables and cannot establish causality; they are not intended for direct comparison with clinical trials

CAV/AVI group vs. polymyxin B group had 75% lower 30-day mortality risk

A single-center retrospective observational study included patients with CRPA infection treated with CAZ/AVI or polymyxin B between January 2018 and December 2020.

- 51 patients were treated with ceftazidime/avibactam monotherapy while 85 patients received the therapy of **polymyxin B mostly combined with other antibiotics** (carbapenems and amikacin etc.)
- After adjustment by propensity score matching, the CAV/AVI group vs. polymyxin B group had
 - Lower 30-day mortality (14.3% vs 42.9%, $p=0.018$)
 - Higher bacterial clearance rate (42.9% vs 14.3%, $p=0.018$)

Table 5 Univariate and Multivariate COX Regression Analysis Associated with 30-Day Mortality After Adjustment

	Univariate Analysis		Multivariate Analysis	
	P value	HR (95% CI)	P value	HR (95% CI)
Age (1-year increments)	0.033	1.047 (1.004–1.093)	0.022	1.050 (1.007–1.094)
Central venous catheter	0.021	0.262 (0.084–0.816)	0.027	0.268 (0.084–0.859)
Duration of therapy (1-day increments)	0.027	0.871 (0.771–0.984)		
CAZ/AVI use	0.031	0.287 (0.092–0.890)	0.016	0.244 (0.078–0.765)

Abbreviations: HR, Hazard ratios; CI, Confidence interval; CAZ/AVI, Ceftazidime/avibactam.

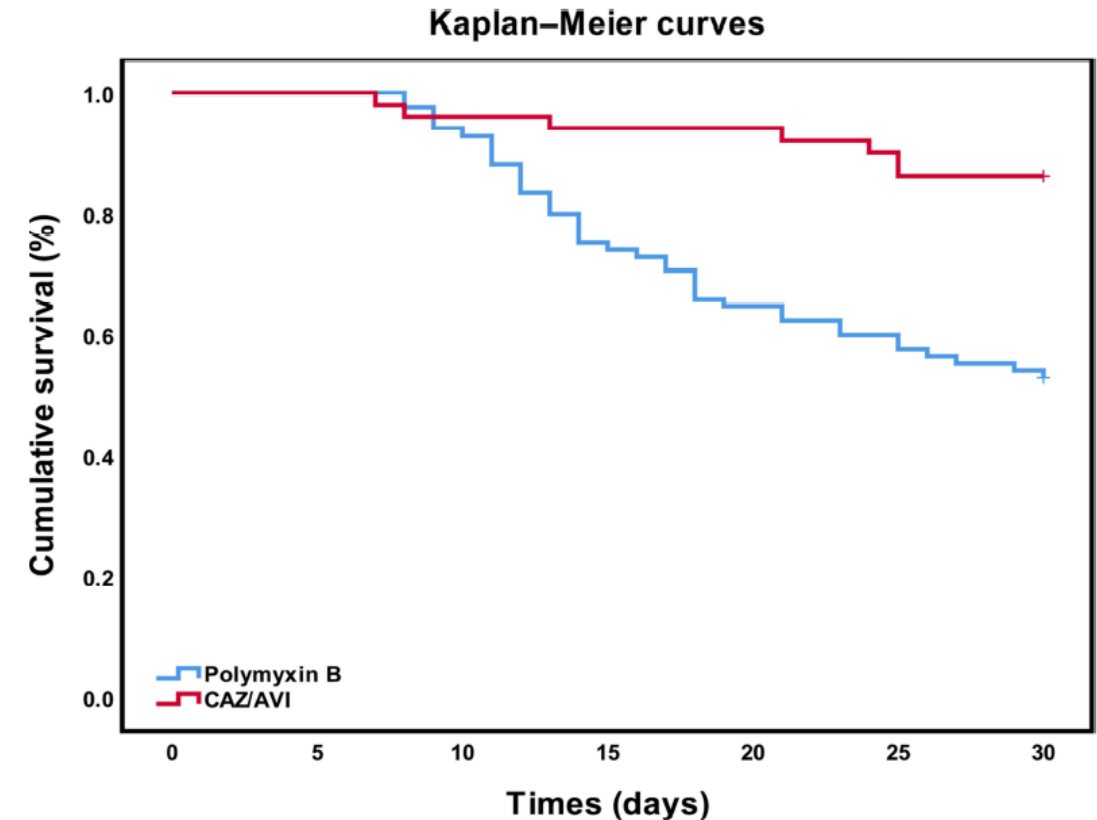


Figure 2 Kaplan–Meier curves of patients with CRPA infection treated with CAZ/AVI or polymyxin B. **Abbreviations:** CRPA, Carbapenem-resistant *Pseudomonas aeruginosa*; CAZ/AVI, ceftazidime/avibactam.

Cefiderocol in HAP/VAP treatment

	HAP/VAP		Overall	
	Cefiderocol (n=45)	Best available therapy (n=22)	Cefiderocol (n=101)	Best available therapy (n=49)
Day 14	11 (24%; 12.9–39.5)	3 (14%; 2.9–34.9)	19 (19%; 11.7–27.8)	6 (12%; 4.6–24.8)
Day 28	14 (31%; 18.2–46.6)	4 (18%; 5.2–40.3)	25 (25%; 16.7–34.3)	9 (18%; 8.8–32.0)
End of study	19 (42%; 27.7–57.8)	4 (18%; 5.2–40.3)	34 (34%; 24.6–43.8)	9 (18%; 8.8–32.0)

Table 5: All-cause mortality in the safety population

	Cefiderocol (n=101)	Best available therapy (n=49)
<i>Acinetobacter</i> spp*	21/42 (50%)	3/17 (18%)
<i>Acinetobacter baumannii</i>	19/39 (49%)	3/17 (18%)
<i>Klebsiella pneumoniae</i>	8/34 (24%)	4/16 (25%)
Without <i>Acinetobacter</i> spp	6/28 (21%)	4/15 (27%)
<i>Pseudomonas aeruginosa</i>	6/17 (35%)	2/12 (17%)
Without <i>Acinetobacter</i> spp	2/11 (18%)	2/11 (18%)
<i>Escherichia coli</i>	1/6 (17%)	0/3
Without <i>Acinetobacter</i> spp	0/3	0/1
<i>Stenotrophomonas maltophilia</i>	4/5 (80%)	NA
Without <i>Acinetobacter</i> spp	2/3 (67%)	NA

Data are n/N (%). NA=not available. *Includes *Acinetobacter baumannii* (for 39 patients assigned cefiderocol and 17 assigned best available therapy), *Acinetobacter nosocomialis* (for two patients assigned cefiderocol), and *Acinetobacter radioresistens* (for one patient assigned cefiderocol).

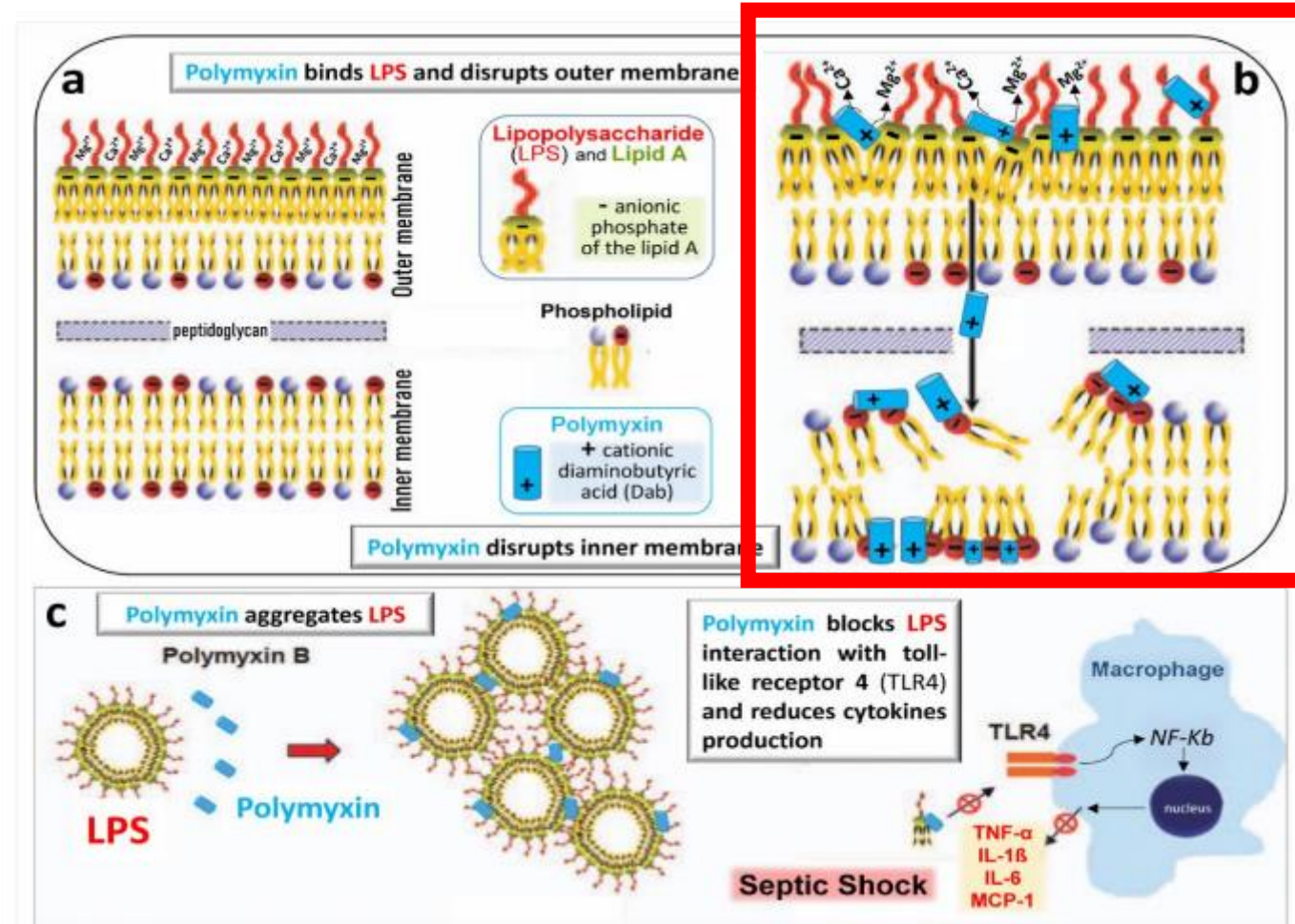
Table 6: All-cause mortality at the end of study by most frequent baseline pathogen in the safety population

In the **cefiderocol** group, 83% (66/80) of patients received **monotherapy**; in the best available therapy group, 71% (27/38) received combination therapy. 25 patients (**66%**) of 38 in the **best available therapy** group received **colistin-based treatment**.

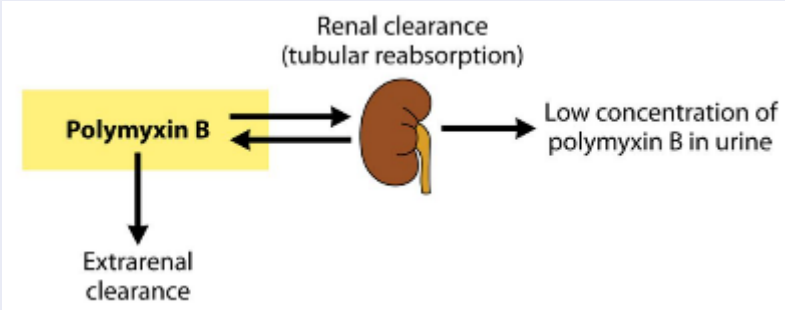
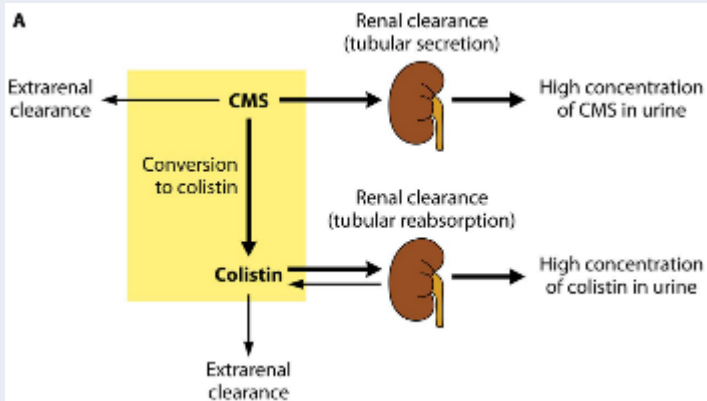
Despite the similarities in clinical and microbiological outcomes, the **all-cause mortality rate in the cefiderocol group was higher** than in the best available therapy group.

Schematic Representation of the Bactericidal Action of Polymyxins

- Cationic, cyclic decapeptide
- Binds anionic lipopolysaccharide (LPS) in outer membrane of GNB
- Displacing Ca^{2+} & Mg^{2+} from outer cell membrane
- Permeability change, leakage of cell content, cell death
- Polymyxin reduce the sepsis severity by blocking LPS interaction with macrophage receptor, TLR4.



The difference between Polymyxin B and Colistin

	Polymyxin B ¹	Colistimethate Sodium (Colistin, Polymyxin E) ¹
Brand Name	Bobimixyn 寶比黴素	Colimycin 克痢黴素
Commercial Formulations	50 mg/vial; 500,000(500K) units/vial	66.8 mg Colistin base/vial; 2,000,000 units/vials
Prodrug Form	No	Yes (Colistimethate sodium)
Pharmacokinetic Pathways	No renal metabolism, excreted unchanged 	Prodrug is metabolized in kidneys to active form; excreted by kidneys 
Dosing Regimen	Loading dose: 2.5mg/kg Maintain dose: 1.5mg/kg, q12h ²	Loading dose: 5mg/kg Maintain dose: 2.5 mg x (1.5 x CrCl + 30) IV divided for q12h ³

Bobimixyn Potential Adverse Effects

- **Nephrotoxicity**: Acute Tubular Necrosis (ATN)
 - Clinical Signs:
 - Rise in **serum creatinine (sCr)** and **blood urea nitrogen (BUN)**.
 - Decreased **urine output**.
 - Electrolyte imbalances (e.g., potassium, magnesium).
- **Neurotoxicity**: Interfere with ion channels on neurons and disrupt the release of the neurotransmitter acetylcholine at the neuromuscular junction.
 - Clinical Signs:
 - **Paresthesias**: A tingling or numbness, most commonly around the mouth, face, and extremities.
 - **Dizziness and Ataxia**: Feeling dizzy, unsteady, or having poor coordination.
 - **Neuromuscular Blockade**: It can cause generalized muscle weakness, which can progress to affect the respiratory muscles, leading to respiratory distress or failure. This risk is much higher in patients also **receiving anesthetics** or other **neuromuscular blockers**.
- **Skin Hyperpigmentation**: Subsequent melanocyte activation
 - Clinical Signs:
 - Predominantly of the head and neck in shades of brown



Clinical outcomes and safety of polymyxin B in the treatment of carbapenem-resistant Gram-negative bacterial infections: a real-world multicenter study

Xiaojuan Zhang¹, Shaoyan Qi², Xiaoguang Duan¹, Bing Han¹, Shuguang Zhang¹, Shaohua Liu¹, Haixu Wang¹, Haibo Zhang^{3*} and Tongwen Sun^{1*} 

- The real-world study included **100 patients treated with intravenous PMB for at least 7 days** during the period of October 2018 through June 2019.
- Temperature (38 °C vs 37.1 °C), white blood cells ($14.13 \times 10^9/l$ vs $9.28 \times 10^9/l$), C-reactive protein (103.55 ug/l vs 47.60 ug/l), procalcitonin (3.89 ng/ml vs 1.70 ng/ml) and APACHE II levels (17.75 ± 7.69 vs 15.98 ± 7.95) were significantly decreased after PMB treatment.
- **Bacteria eradication** rate was **77.65%**. **Overall mortality** at discharge was **15%**, and **28-day mortality** was **40%**. Major adverse reactions occurred in 16 patients. **Nephrotoxicity** was observed in 7 patients (**7%**).

Polymyxin B 可參考Colistin 藥敏作為處方依據 MIC $\leq$ 2可考慮使用



台灣感染症醫學會
The Infectious Diseases Society of Taiwan

26th Annual Meeting of Antimicrobial Resistance in Taiwan

*The Burden of Multidrug-resistant Organisms and the Potential Application of
Information and Bio-molecular Technologies*

Polymyxin B 在常見的MDR-GNB MIC $\leq$ 2 比例高(9成以上) ,
根據CLSI 及其他文獻論述：

- Polymyxin B 藥敏可參考Colistin ；
- Polymyxin B MIC 可參考Colistin ， MIC $\leq$ 2 即可考慮使用

In vitro MDR Enterobacterales sensitivity from ATLAS (2021-2022) with HAP/VAP and IAI

All 14564 isolates: CRE prevalence 10.4%, MBL 4% by EUCAST method

Phenotype		Antimicrobials	%S (EUCAST)	% R (EUCAST)	MIC ₉₀ (mg/L)/% susceptible (%S), EUCAST															
					MIC ₅₀	MIC ₉₀	MIC range	Aztreonam/ avibactam		Aztreonam		Colistin		Cefiderocol		Cefepime		Tigecycline ^a		
All Enterobacterales (n=14 564)																				
MDR (n=6217)	Amikacin	82.7	17.3	2	128	0.25–128	Genotypes	n	%S	MIC ₉₀	%S	MIC ₉₀	%S	MIC ₉₀	%S	MIC ₉₀	%S	MIC ₉₀		
	Aztreonam	30.5	69.5	32	128	0.03–128														
	Aztreonam/avibactam	98.9	1.0	0.12	0.5	0.015–128														
	Cefepime	43.9	56.0	8	64	0.12–64														
	Colistin	83.0	17.0	0.25	16	0.12–16														
	Meropenem	82.5	17.5	0.06	32	0.06–32														
	Tigecycline*	93.9	6.1	0.5	2	0.03–16														
ESBL (n=1688)	Amikacin	66.3	33.6	4	128	0.25–128	Overall (n=577)													
	Aztreonam	2.3	97.7	128	128	0.12–128	NDM	508	94.5	2	13.8	>64	74.8	>8	47.4	8	0.0	>32	98.0	2
	Aztreonam/avibactam	99.0	1.0	0.12	0.5	0.015–128	VIM	48	100.0	2	14.6	>64	72.9	>8	72.9	4	0.0	>32	81.0	4
	Cefepime	6.5	93.5	64	64	0.12–64	IMP	25	100.0	2	24.0	>64	60.0	>8	68.0	4	8.0	>32	68.0	>8
	Colistin	86.5	13.5	0.25	8	0.12–16	Co-carriage													
	Meropenem	62.0	38.0	0.5	32	0.06–32	OXA-48-like	145	98.6	0.5	6.2	>64	73.8	>8	61.4	4	0.0	>32	99.3	2
	Tigecycline*	97.2	2.8	0.5	2	0.06–16	KPC	30	100.0	1	0.0	>64	60	>8	40.0	16	0.0	>32	86.7	4
CRE (n=1088)	Amikacin	33.3	66.6	64	128	0.5–128	ESBL ^b (n=450)	353	98.9	1	0.6	>64	74.8	>8	50.1	8	0.0	>32	98.6	2
	Aztreonam	7.0	93.0	128	128	0.03–128	pAmpC ^b (n=450)	72	66.7	16	2.8	>64	84.7	8	37.5	>32	0.0	>32	94.4	1
	Aztreonam/avibactam	96.4	3.6	0.25	1	0.015–128	YRIK ^c (n=81)	21	57.1	32	0	>64	100.0	0.5	19.1	4	0.0	>32	100.0	0.5
	Cefepime	1.0	99.0	64	64	1–64	YRIN ^c (n=81)	40	52.5	16	2.5	>64	100.0	0.25	20.0	>32	0.0	>32	100.0	0.5
	Colistin	75.9	24.1	0.25	16	0.12–16	YRIK/YRIN + pAmpC	35	31.4	32	0	>64	100.0	0.25	17.1	>32	0.0	>32	100.0	0.5
	Meropenem	0	100	32	32	16–32														
	Tigecycline*	90.2	9.8	1	2	0.12–16														

MBL NDM S

Aztreonam/avibactam: 94.5% but poor activity

With RBP

MBL NDM S

Aztreonam/avibactam: 94.5% but poor activity
With PBP

Colistin 74.8%; Cefiderocol 47.4%; Tigecycline 98%

CRE S

Aztreonam/avibactam: 96.4%

Colistin 75.9%

Tigecycline 90.2%

In vitro MDR Enterobacterales bacteremia sensitivity from ATLAS (2021-2022)

All 10376 isolates (46% KP, 24% E coli, 11% Enterobacter)

: CRE prevalence 6.8% (74.3% KP, 11.1% E coli), MBL 3.3% by EUCAST method

Antimicrobial agent	Susceptibility (EUCAST)		MIC (mg/L)		No. of isolates inhibited at respective MIC (mg/L) (% cumulative inhibition)										
	%S	%R	MIC ₅₀	MIC ₉₀	≤0.06	0.12	0.25	0.5	1	2	4	8	16	32	>32
CRE isolates (N = 710) ^a															
Aztreonam/avibactam	98.9	1.1	0.25	1	62 (8.7)	140 (28.5)	266 (65.9)	158 (88.2)	44 (94.4)	14 (96.3)	18 (98.9)	5 (99.6)	0 (99.6)	0 (99.6)	3 (100)
Aztreonam	5.6	93.4	128	128	7 (1)	5 (1.7)	10 (3.1)	14 (5.1)	4 (5.6)	1 (5.8)	6 (6.6)	4 (7.2)	9 (8.5)	23 (11.7)	627 (100)
Cefepime	0.1	99.3	64	64	0	0	0	0	1 (0.1)	1 (0.3)	3 (0.7)	9 (2.0)	12 (3.7)	47 (10.28)	637 (100)
Colistin	80.1 ^c	19.9	0.25	16	0	183 (25.8)	315 (70.1)	36 (75.2)	21 (78.2)	14 (80.1)	19 (82.8)	33 (87.5)	89 (100)		
Meropenem	0	100	32	32	0	0	0	0	0	0	0	0	128 (18.0)	582 (100)	
Tigecycline ^b	93 ^c	1.3	1	2	0	15 (2.1)	58 (10.3)	193 (37.5)	257 (73.7)	137 (93.0)	41 (98.7)	7 (99.7)	2 (100)		
Amikacin	43.1	56.9	16	128	0	0	0	4 (0.6)	60 (9.0)	63 (17.9)	100 (32)	79 (43.1)	58 (51.3)	45 (57.6)	301 (100)

CRE S

Aztreonam/avibactam: 98.9%

Colistin 80.1%

Tigecycline 93%

Antimicrobial agent	Susceptibility (EUCAST)		No. of isolates inhibited at respective MIC (mg/L) (% cumulative inhibition)												
	%S	%R	≤0.06	0.12	0.25	0.5	1	2	4	8	16	32	>32		
NDM (N = 407) ^a															
Aztreonam/avibactam	96.8	3.2	74 (18.2)	66 (34.4)	103 (59.7)	99 (84.0)	32 (91.9)	8 (93.9)	12 (96.8)	7 (98.5)	4 (99.5)	0 (99.5)	2 (100)		
Aztreonam	10.1	87.5	12 (2.9)	3 (3.7)	12 (6.6)	6 (8.1)	8 (10.1)	5 (11.3)	5 (12.5)	6 (14.0)	5 (15.2)	27 (21.9)	318 (100)		
Cefepime	0.2	99.5	0	1 (0.2)	0 (0.2)	0 (0.2)	0 (0.2)	1 (0.5)	0 (0.5)	2 (1.0)	6 (2.5)	15 (6.1)	382 (100)		
Cefiderocol	51.1	48.9	0	3 (0.7)	2 (1.2)	7 (3.0)	49 (15.0)	147 (51.1)	146 (87.0)	26 (93.4)	7 (95.1)	3 (95.8)	17 (100)		
Colistin	81.3 ^d	18.7	0	5 (1.2)	225 (56.5)	83 (76.9)	12 (79.9)	6 (81.3)	3 (82.1)	20 (87.0)	53 (>8)	100			
Ertapenem	0.7	99.3	2 (0.5)	0 (0.5)	1 (0.7)	0 (0.7)	1 (1.0)	0 (1.0)	1 (1.2)	7 (3.0)	395 (>8)	100			
Minocycline ^b	34.9	39.8	0	0	2 (0.5)	6 (2.0)	33 (10.1)	53 (23.1)	48 (34.9)	103 (60.2)	74 (78.4)	88 (>16)	100		
Tigecycline ^c	97.5 ^d	0	2 (0.5)	25 (6.6)	86 (27.8)	103 (53.1)	132 (85.5)	49 (97.5)	10 (100)						
Tobramycin	10.1	89.9	0	1 (0.3)	12 (3.2)	18 (7.6)	7 (9.3)	3 (10.1)	11 (12.8)	50 (25.1)	305 (>8)	100			

MBL NDM S

Aztreonam/avibactam: 96.8%

Colistin 81.3%; Cefiderocol 51.1%; Tigecycline 97.5%

In vitro activity of aztreonam/avibactam in CRE from US 62 centers (2017-2022)

Agent	MIC ₅₀ (mg/L)	MIC ₉₀ (mg/L)	Range (mg/L)	%S	%I	%R
CREs with only class A carbapenemases (360 isolates)						
Aztreonam-avibactam	0.25	0.5	≤0.03–8	99.4	N/A	0.6
Ceftazidime-avibactam	1	2	<0.03–32	98.9	N/A	1.1
Meropenem-vaborbactam	0.03	0.5	<0.03–>32	98.3	0.6	1.1
Imipenem-relebactam	0.125	1	0.03–>8	92.8	2.2	5.0
Cefiderocol	0.5	2	<0.03–64	98.9	0.3	0.8
Tigecycline	0.5	2	≤0.06–8	96.7	1.9	1.4
Colistin	0.25	>8	≤0.06–>8	N/A	82.7	17.3
CREs with class B carbapenemases ^c (50 isolates)						
Aztreonam-avibactam	0.25	0.5	≤0.03–8	98.0	N/A	2.0
Ceftazidime-avibactam	>32	>32	0.125–>32	4.0	N/A	96.0
Meropenem-vaborbactam	16	>32	0.5–>32	16.0	6.0	78.0
Imipenem-relebactam	8	>8	1–>8	2.0	4.0	94.0
Cefiderocol	2	16	<0.03–32	74.0	14.0	12.0
Tigecycline	0.5	2	0.25–4	92.0	8.0	0.0
Colistin	0.25	16	0.06–>8	N/A	86.0	14.0
CREs with only class D carbapenemases (12 isolates)						
Aztreonam-avibactam	0.25	0.5	≤0.03–0.5	100.0	N/A	0.0
Ceftazidime-avibactam	1	2	0.125–2	100.0	N/A	0.0
Meropenem-vaborbactam	32	>32	1–>32	16.7	8.3	75.0
Imipenem-relebactam	4	8	1–>8	16.7	0.0	83.3
Cefiderocol	0.5	2	<0.03–4	100.0	0.0	0.0
Tigecycline	1	4	0.25–4	83.3	16.7	0.0
Colistin	0.125	0.25	≤0.06–>8	N/A	91.7	8.3
CREs without carbapenemases (89 isolates)						
Aztreonam-avibactam	0.5	4	0.03–>16	94.4	N/A	5.6
Ceftazidime-avibactam	1	8	<0.03–>32	96.6	N/A	3.4
Meropenem-vaborbactam	1	8	0.03–16	88.8	9.0	2.2
Imipenem-relebactam	0.25	2	0.06–4	89.9	5.6	4.5
Cefiderocol	1	8	0.03–64	88.8	5.6	5.6
Tigecycline	0.5	4	≤0.06–4	89.9	10.1	0.0
Colistin	0.25	2	≤0.06–>8	N/A	91.0	9.0

- All 54576 isolates: CRE prevalence 0.9%, MBL 3.7% in CRE
- Class A: S% 92.8-99.4%, colistin I% 82.7%
- Class B: ATM-AVI 98%S, Tige 92%S, Coli 86%I
- Class D: ATM-AVI 100%S, Cef-Avi 100%S, Cefiderocol 100%S, Coli 91.7%I

Papp-Wallace KM et al, Open Forum Infect Dis . 2025 Apr 25;12(5):ofaf250.

ATM-AVI R CRE by ATLAS (2019-2023)

Fig. 1

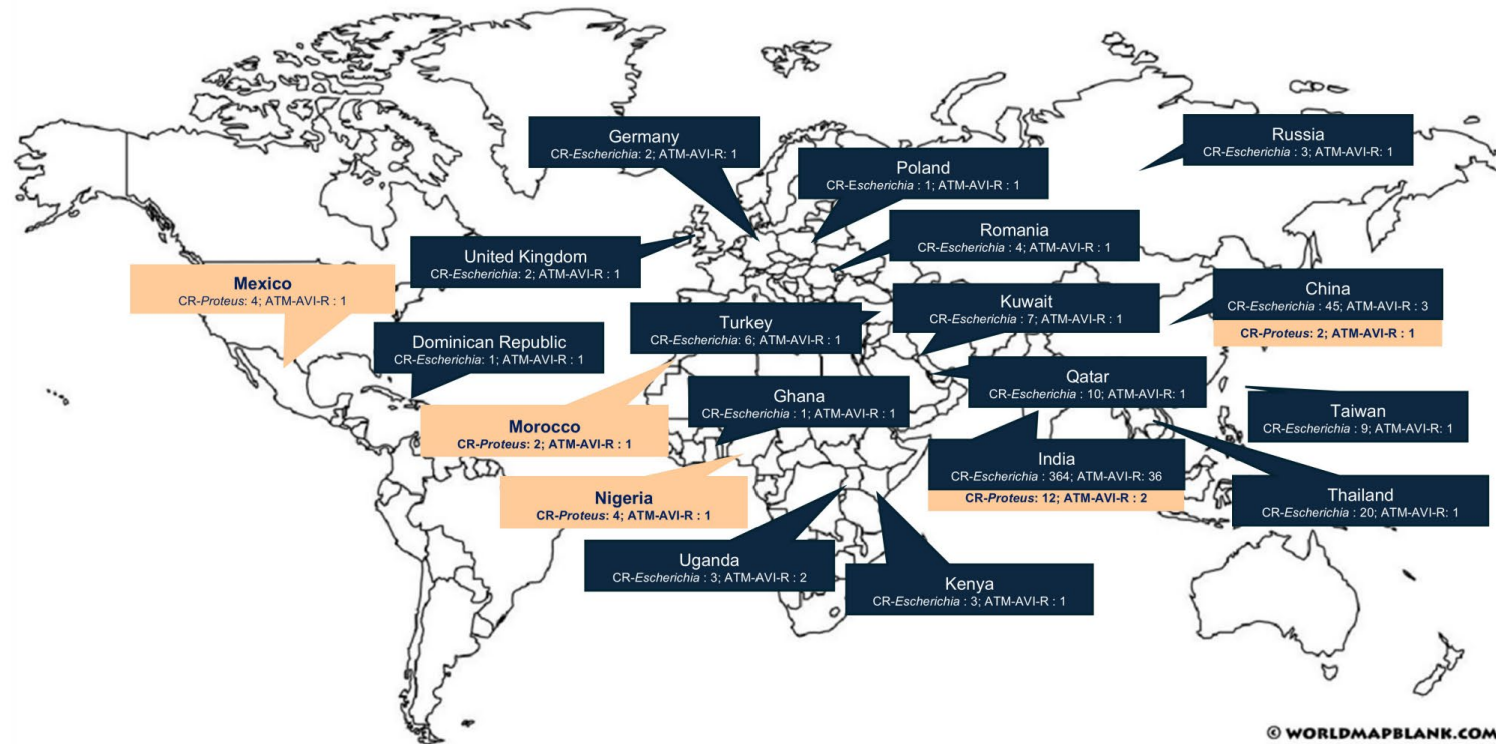


FIG 1 Geographic distribution of ATM–AVI-R carbapenem-resistant isolates belonging to *Escherichia coli* (CR-*Escherichia*) and *Proteus mirabilis* (CR-*Proteus*) collected worldwide in the ATLAS program, 2019–2023. Countries with CR-*Escherichia* and CR-*Proteus* but without ATM–AVI-R isolates are not shown, including Argentina, Australia, Belgium, Brazil, Canada, Colombia, Costa Rica, Croatia, Czech Republic, Denmark, France, Greece, Guatemala, Israel, Italy, Ivory Coast, Japan, Jordan, Latvia, Lithuania, Malawi, Malaysia, Panama, Philippines, South Africa, South Korea, Spain, Ukraine, United States, and Venezuela. Image source: World Map Blank (<https://worldmapblank.com/>).

Lee YL et al, Antimicrob Agents Chemother. 2026 Apr;70(4):e0154925.

Primary resistance mechanisms	Species	MIC (µg/mL)	Geographical source	Predominant MLST (ST)	Key references
Enterobacterales					
1. PBP3 Insertions + AmpC/ESBL Production					
PBP3 insertion (YRIN/YRIK) + CMY-42 AmpC	<i>E. coli</i>	8-32	Global	ST167, ST361, ST405, ST410, ST617, ST38	Sadek et al. [78]; Le Terrier et al. [79]; Mushtaq et al. [80]; Mendes et al. [81]; Tellapragada et al. [82]
PBP3 insertion (YRING) + CMY-145 AmpC	<i>E. coli</i>	8	Singapore	ST167, ST361	Chew et al. [83]
PBP3 insertion (YRIN/YRIK) + PER-2/PER-6/PER-7 ESBL	Engineered <i>E. coli</i>	8-256	NA	NA	Le et al. [84]
PBP3 insertion (YRIN) + KPC-21 mutation (W105R) + CMY-42 AmpC	<i>E. coli</i>	512	China	ST410	Ma et al. [85]
PBP3 insertion (YRIN/YRIK) + NDM5/NDM1/OXA+CMY-42 AmpC + porins loss	<i>E. coli</i>	8-32	Sweden	ST361, ST405, ST167, ST448, ST410, ST4450, ST617, ST1284	Tellapragada et al. [82]
PBP3 insertion (YRIN) and mutation (A417V) + CMY-145 AmpC mutation (L139R/N366Y)	<i>E. coli</i>	128		ST361	Haidar at al. [86]
2. PER-type β-Lactamases					
PER-2 + AmpC + ompF-like truncation	<i>E. cloacae</i>	8-16	Latin America	ST200, ST1324, ST88	Estabrook et al. [87]
PER-4 + OXA-1	<i>K. oxytoca</i>	16	Latin America	ST40	
PER-14 + SHV + TEM-1	<i>R. ornithinolytica</i>	16	Latin America	NA	
3. KPC Variants + Porin Defects					
KPC-2 S109P + OmpK36 truncation + Efflux pump upregulation	<i>K. pneumoniae</i>	128	USA	ST258	Xian et al. [88]
KPC overexpression + OmpK35/36/37 defects	<i>K. pneumoniae</i>	32-128	China	ST11	Yu et al. [89]
4. Novel PBP3 + Porin Mutations					
PBP3 Gly420dup + OmpC G151D	<i>P. hangzhouensis</i>	>64	Singapore	ST44	Chew et al. [90]
5. Efflux/Porins Alterations					
acrA efflux overexpression + OmpK35/36 truncation + DHA-1 AmpC	<i>K. pneumoniae</i>	8-16	Argentina, Taiwan, Thailand	ST872, ST15, ST273	Mendes et al. [81]
OmpC/OmpK36 variants (P177A, D188E) + PBP3 variant (S259insE) + AmpC hyperproduction	<i>E. cloacae</i> complex	16	Poland	ST78	Mendes et al. [81]
6. Altered β-Lactamase-Avibactam Interaction					
SHV-12 R244G substitution	<i>Enterobacter mori</i>	16	China	NA	Wu et al. [91]
CMY-16 T150S/N346H mutation + OmpK porin defects	<i>K. pneumoniae</i>	8-256	USA	ST101, ST147, ST377	Niu et al. [92]
Non-fermenting Gram-negative bacteria					
1. Efflux/Porins Alterations					
MexAB-OprM overexpression + OprD porin loss	<i>P. aeruginosa</i>	8	Sapin, Italy	ST175, ST235	Sastre et al. [93]; Gonzalez et al. [94]
2. Pyocyanin Overproduction					
PA4292 mutation or deletion-induced Pyocyanin overproduction	<i>P. aeruginosa</i>	128-256	China	NA	Zhao et al. [95]

Abbreviations: MIC, minimal inhibitory concentration; MLST, multilocus sequence typing; YRIN, an amino-acid insertion (Tyr-Arg-Ile-Asn) in the PBP3 sequence; YRIK, an amino-acid insertion (Tyr-Arg-Ile-Lys) in the PBP3 sequence; YRING, an amino-acid insertion (Tyr-Arg-Ile-Asn-Gly); *E. coli*, *Escherichia coli*; *E. cloacae*, *Enterobacter cloacae*; *K. oxytoca*, *Klebsiella oxytoca*; *R. ornithinolytica*, *Raoultella ornithinolytica*; *K. pneumoniae*, *Klebsiella pneumoniae*; *P. hangzhouensis*, *Providencia hangzhouensis*; *P. aeruginosa*, *Pseudomonas aeruginosa*; NA, data not available.

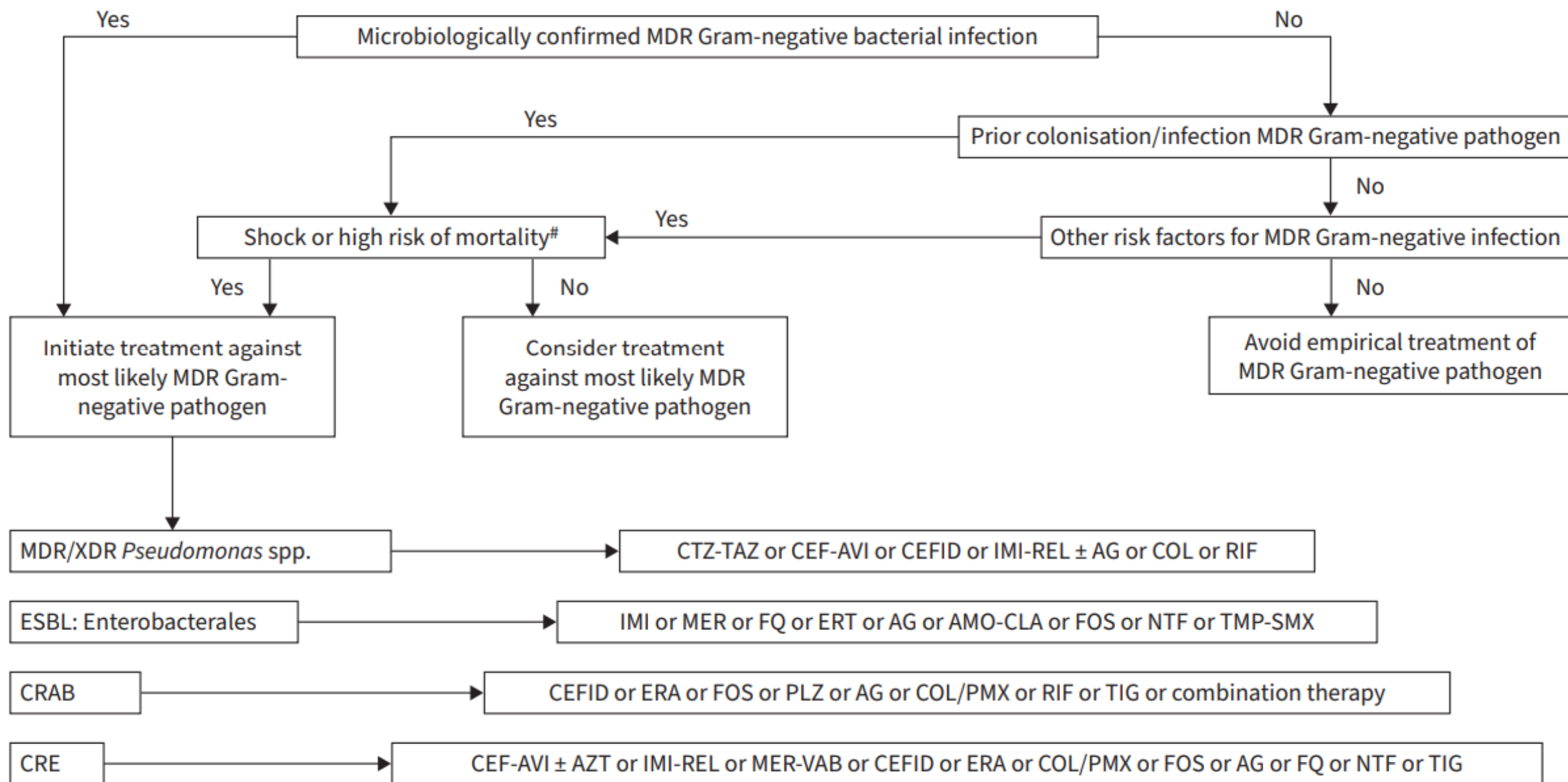
Antimicrobial stewardship programme

	Description	Outcomes
Pre-authorisation	Prescribers are required to gain approval prior to use of certain antimicrobials	Ensures appropriate antibiotic selection and dosing Prevents unnecessary initiation of antibiotics
Prospective audit and feedback	External review of antibiotics after initiation, with suggestions from experts on optimal use	Reduction in unnecessary use of broad-spectrum antibiotics
Facility-specific treatment guidelines	Specific treatment guidelines for common conditions, such as community-acquired pneumonia, UTI and surgical prophylaxis, taking into account national guidelines and local susceptibilities and formulary options	Simplifies antibiotic prescribing for common conditions Allows for consensus between stewardship team and prescribers
Antibiotic time-outs	Provider-led reassessment of the continuing need for antibiotics after additional clinical information has returned	Improves the appropriateness of antibiotic use Encourages antibiotic de-escalation when appropriate
Pharmacy-based interventions	Prescribing interventions by pharmacy to optimise antibiotic use May be incorporated into electronic health record	Dose optimisation, dose adjustments based on renal function, changing intravenous antibiotics to oral formulation
Rapid diagnostics	Early pathogen identification using technology such as PCR to identify bacterial infection prior to awaiting culture results	Earlier identification of causative organism can lead to antibiotic de-escalation and optimisation of antibiotics
Clinical decision support system	Incorporation of clinical and patient-specific data into electronic health record to provide providers with relevant information prior to prescribing antibiotics	Decreased overall antibiotic use and improved use of antibiotics in patients with sepsis

Recommendations from Taiwan IDST 2024

	CRAB		CRE		DTR-PA
Recommended treatment	Pneumonia	BSI	BSI	Pneumonia	• Ceftolozane-tazobactam • Ceftazidime-avibactam • Imipenem-relebactam • Cefiderocol • Polymyxin based therapy: Colistin or Polymyxin B
	• Tigecycline + β-lactams or Imipenem • Cefiderocol +/- Tigecycline or Fosfomycin or Carbapenem • Sulbactam-durlobactam + Imipenem • Sulbactam+Tigecycline • Polymyxins(Colistin or Polymyxin B) + Fosfomycin Any one of the above IV regimens + Adjunctive Colistin inhalation	• Polymyxins(Colistin or Polymyxin B) + Tigecycline or Sulbactam • Cefiderocol +/- Tigecycline or Fosfomycin	• Ceftazidime-avibactam • Meropenem-vaborbactam • Imipenem-relebactam • Polymyxin based combination: Colistin or Polymyxin B + Tigecycline or Amikacin	• Ceftazidime-avibactam • Polymyxin based combinations: Colistin or Polymyxin B • Imipenem-relebactam Any one of the above IV regimens + Adjunctive Colistin inhalation	
			cUTI	cIAI	
			• Ceftazidime-avibactam • Meropenem-vaborbactam • Imipenem-relebactam • Aminoglycosides • Colistin	• Ceftazidime-avibactam • Imipenem-relebactam • Tigecycline or Eravacycline • Polymyxin based combination: Colistin or Polymyxin B + Tigecycline • Cefiderocol	

IDST 2024.7 Prefinal Version



Take home messages

- *Carbapenem-resistant GNBs* are important nosocomial pathogens with high mortality.
- Choice of antibiotics against CR-GNB should consider **resistance mechanisms**.
- For **CRE** severe infection, **monotherapy with newer antibiotics**, such as Ceftazidime-avibactam, Meropenem-vaborbactam, Imipenem-relebactam, or cefiderocol was recommended.
- **KPC variant** is important in Ceftazidime-avibactam resistance and imipenem-relebactam preserves sensitivity in them.
- **Polymyxins** might be considered in CRE in local guidance with caution in adverse event and treatment efficacy.
- **Aztronam-Avibactam** has excellent in vitro activity against MBL pathogens.

Thank you for your attention.